New putative insights into Neprilysin (NEP)-dependent pharmacotherapeutic role of Roflumilast in treating COVID-19

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Abstract

Nowadays, COVID-19 represents the most serious inflammatory respiratory disease worldwide. Despite of many proposed therapies, no effective medication has been approved yet. Neutrophils appear to be the key mediator for COVID-19-associated inflammatory immunopathologic, thromboembolic and fibrotic complications. Thus, for any therapeutic agent to be more appropriate, it should greatly block the neutrophilic component of COVID-19. One of the effective therapeutic approaches investigated to reduce neutrophils-associated inflammatory lung diseases with less adverse effects was Roflumilast. Being a highly selective PDE4i, roflumilast acts by enhancing cAMP level, that probably potentiates its anti-inflammatory action via increasing NEP activity. Because activating NEP was previously reported to mitigate several airway inflammatory ailments; this review deeply discusses the proposed NEP-based therapeutic properties of roflumilast, which may be of great importance in curing COVID-19. However, further clinical studies are required to confirm this strategy and to evaluate its in-vivo preventive and therapeutic efficacy against COVID-19.

Abbreviations

AC: Adenylate cyclase ACE: Angiotensin-converting enzyme ACE2: Angiotensin-converting enzyme 2 AD: Atopic dermatitis Ang II: Angiotensin II cAMP: Cyclic adenosine monophosphate c-ATP: Cellular adenosine triphosphate cGMP: Cyclic guanosine mono-phosphate COPD: Chronic obstructive pulmonary disease CRP: C-reactive protein DM: Diabetes mellitus

ECs: Endothelial cells

ET-1: Endothelin-1

FEV1: forced expiratory volume in 1 s

GIT: Gastrointestinal tract

GLP-1: Glucagon like peptide-1

HCQ: Hydroxychloroquine

IBD: Inflammatory bowel diseases

IL-6: Interleukin-6

JAK: Janus kinase

NEP: Neutral endopeptidase, Neprilysin

NETs: Neutrophils extracellular traps

PAF: Patelet activating factor

PDE4i, Phosphodiesterase-4 inhibitor

PIC: Pulmonary Intravascular Coagulopathy

RA: Rheumatic arthritis

RNO: Roflumilast-N-oxide

SARS-CoV-2: Severe acute respiratory syndrome coronavirus 2

SFK-PI3K: Src family kinases phosphoinositide 3-kinase

STAT-3: Signal transducer and activator of transcription-3

WHO: World Health Organization

3CLpro: 3 chymotrypsin like protease

1. Introduction

Coronavirus disease 2019 (COVID-19) is a global infectious illness that results in a huge number of deaths. For restricting its spread, there is an urgent need to evok the most effective therapy. (Li et al., 2020b). Recently, a study hypothesizes that using the anti-inflammatory phosphodiesterase-4 inhibitor (PDE4i) for modulating COVID-19 may be beneficial (Bridgewood et al., 2020). Among PDE4i, roflumilast exhibits the highest potency for targeting and blunting airway inflammation via enhancing the level of cAMP (Rabe, 2011), which in turn may prolong its anti-inflammatory effect by activating Neprilysin (NEP) (Graf et al., 1995). As NEP is lately supposed to be a new potential target for COVID-19 therapy (El Tabaa and El Tabaa, 2020), roflumilast-enhanced NEP activity may have a prominent significance. Thus, we aim to review the proposed NEP-dependent pharmacological mechanisms by which roflumilast can block COVID-19-associated inflammatory, coagulopathy and fibrotic cascades.

2. COVID-19 challenges

COVID-19 is a contagious fatal respiratory disease caused by a novel virus called severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2). It was first recognized at the end of 2019 in Wuhan, China until being now an ongoing pandemic (Huang et al., 2020). As of 30 June 2020, more than 10.3 million cases have been reported across 188 countries and territories, resulting in more than 507,000 deaths and more than 5.28 million people have recovered (CSSE, 2020).

2.1 Clinical manifestations of COVID-19

Being one of severe airway diseases, COVID-19 patients usually show typical symptomatic respiratory presentations, such as cough, tiredness, muscle aches, headache, sore throat with sometimes fever and chills (Singhal, 2020). In such cohort, some patients may suffer from other worsened symptoms, such as profound acute shortness of breath combined with persistent chest pain, increasing the emergency need for oxygen therapy and mechanical ventilation (Yang et al., 2020). On the contrary, there are asymptomatic carrier states, who experience no symptoms or even only very mild symptoms; increasing thereby the risk of disease transmission (Lai et al., 2020).

Case reports declare that some people may display other unusual non-respiratory manifestations such as diarrhea which is recognized to be an initial sign of COVID-19 infection, in addition to taste or olfactory disorders which are especially identified in young people infected with SARS-CoV-2 (Luërs et al., 2020; Song et al., 2020).

Early clinical studies report that critically ill COVID-19 patients may associate with cardiovascular insults including myocardial injury, myocarditis, cardiac arrhythmias and heart failure with increased risk for thromboembolism as pulmonary embolus because of COVID-19-induced hypercoagulable state (Driggin et al., 2020).

Other cases with COVID-19 may also exhibit some neurological symptoms including dizziness, ataxia, altered mental state or even seizures (Mao et al., 2020). As well, some common COVID-19-related complications have been detected involving elevated liver enzymes, acute kidney injury (AKI) as well as an increased risk of developing fatal bacterial infections (Cox et al., 2020; Yang et al., 2020). Lately, ocular abnormalities such as conjunctival hyperemia, chemosis, and increased secretions are additionally reported in COVID-19 infected patients (Wu et al., 2020).

2.2 High-risk groups of COVID-19

As documented, COVID-19 can infect different groups of people, where most of them will recover without hospitalization, but others will develop sever illness. People with high risk for contracting COVID-19 infection include older people, usually over 60 to 70 years old and those who have weakened immune response either due to administering chemotherapy, radiation or medication for an autoimmune disease, undergoing an organ or stem cell transplant, losing a spleen or having non-functioning one. Moreover, adults (over 18 years old) with underlying chronic medical conditions such as high blood pressure, diabetes, chronic heart, lung and kidney diseases are more vulnerable to succumbed to COVID-19 infection (Vishnevetsky and Levy, 2020). Similarly, pregnant women appear to be more susceptible to COVID-19 with the potential of developing maternal and fetal complications (Liu et al., 2020a). As well, there is also an increased risk for overweight people and heavy cigarettes smokers (Tamara and Tahapary, 2020; van Zyl-Smit et al., 2020).

On the other hand, all children, even those with underlying medical problems, did not show a high risk of severe illness from COVID-19 (Lyu et al., 2020).

3. Pathophysiology of COVID-19

Since the prevalence of COVID-19 has nowadays become a major global burden around the world, there has been a necessity to perform the precious pathophysiological researches that will aim at recognizing the involved biological markers and the clear mechanisms through which the disease pathogenicity induced by SARS-CoV-2 can be explained.

Obviously, the coronavirus genome cannot be replicated outside the cytoplasmic membranes, so it continuously seeks to penetrate living cells for ensuring its survival. For viral replication, polyproteins should be firstly hydrolyzed into functional proteins by a variety of proteolytic enzymes, which are more commonly known to RNA viruses such as RNA dependent RNA polymerase (RdRp), 3 chymotrypsin like protease (3CL protease), papain like protease and helicase (Ziebuhr, 2005).

At present, several studies showed that penetrating pneumocytes is considered as the main way for SARS-CoV-2 replication within the human body. The finding which is ensured from the evidence of utilizing angiotensin-converting enzyme 2 (ACE-2) enzyme as receptors for viral entry, **Figure 1** (Zhang et al., 2020a). ACE-2 was found to be highly expressed in alveolar and bronchial membranes, in type II pneumocytes and

possibly on vascular endothelial cells (EC) within lungs (Jia, 2016); explaining why the common signs and symptoms of respiratory infection will develop in coinciding with COVID-19 disease.

Simultaneously, ACE-2 protein was also detected to be distributed in various human organs other than lungs involving oral and nasal mucosa, GIT, skin, heart, liver, kidney, and brain (Hamming et al., 2004); elucidating the reason for developing other extra-pulmonary manifestations associated with COVID-19 infection.

Binding of SARS-CoV-2 with ACE-2 may downregulate ACE-2 and subsequently, inhibit the ACE-2regulated generation of angiotensin (1–7) peptide which can, via MAS-receptor (MasR), perform several beneficial activities as vasodilator, anti-inflammatory, anti-hypertrophy, anti-proliferative, anti-fibrosis and antioxidant (Kuba et al., 2005).

Concerning the pulmonary RAS, cutting off the ACE-2/Ang (1–7)/MasR axis will activate the vasopressor ACE/Angiotensin (Ang) II /Ang II type 1 receptor (AT1) axis on the other side. The axis which may dive the airway inflammatory cascades, as a result of significant increase in Ang II level. Ang II, through activating the AT1 receptor, could promote the release of multiple inflammatory cytokines especially TNF- α , IL-6, GM-CSF and MCP-1 (Sprague and Khalil, 2009).

3.1 Cytokine storm in COVID-19

In response to massive amounts of released cytokines, a form of inflammatory reaction would be triggered, namely cytokine storm syndrome, that is characterized by higher IL-6 levels. This is obviously detected in seriously ill COVID-19 patients and may further weaken their immune response (Dal Moro and Livi, 2020).

Because of the positive correlation between high IL-6 level and COVID-19 severity, IL-6 is specifically suggested to be the master marker used for monitoring disease progression (Liu et al., 2020b). There a growing evidence that IL-6 can play a crucial part in the uncontrolled intestinal inflammatory process, proving its role in the pathogenesis of COVID-19-asociated diarrhea. However, another causing factor may be attributed to the direct viral invasion of gut epithelial cells via ACE-2 (Mudter and Neurath, 2007).

As previously reported, IL-6 could prohibit the olfactory signal pathway; proposing that anosmia detected in COVID-19 patients may be due to IL-6-mediated inflammation of the nasal mucosa (Henkin et al., 2013; Luërs et al., 2020). Besides, other additional elements supporting that SARS-CoV-2 may have a neuroinvasive propensity to invade the central olfactory pathway causing an olfactory dysfunction (Marinosci et al., 2020). Jointly, IL-6 was also found to be extremely involved in promoting the ocular inflammation; matching with conjunctivitis that is recently reported to be linked with COVID-19 infection (Ghasemi, 2018).

3.2 IL-6-induced endothelial dysfunction and coagulopathy in COVID-19

In addition to the direct role of SARS-CoV-2/ACE-2 interaction in inducing the endothelial dysfunction (Zhang et al., 2020b), IL-6 was also reported to interrupt the normal function of endothelial cells (ECs) through inactivating the endothelial nitric oxide synthase (eNOS) which in turn could decrease NO production with subsequent induction of an oxidative stress state leading to impairment in endothelial responses (Hung et al., 2010).

As a consequence, disrupting the endothelial cells function either by SARS-CoV-2 itself or IL-6 could activate the platelets and stimulate their adhesion and aggregation; resulting in a pulmonary specific vasculopathy termed Pulmonary Intravascular Coagulopathy (PIC) (Aird, 2003; Levi and van der Poll, 2017; McGonagle et al., 2020).

Most anatomical studies of COVID-19 victims demonstrate the formation of blood thrombus (fibrin clot) in their pulmonary vessels, in addition to deep vein thrombosis that increases the risk for developing pulmonary embolism (Cui et al., 2020; Klok et al., 2020). These clots result in a compensatory increase of plasminogen (fibrinolysin) but, with disease progression, it fails to break down these fibrin deposits reflected in elevated D-dimer (DD) levels, which is reported to be associated with the severity of COVID-19 infection and may be also correlated with activation of the pro-inflammatory cytokine cascade (Belen-Apak and Sarialioğlu, 2020; Leonard-Lorant et al., 2020).

Emerging data suggest that COVID-19-associated endothelial dysfunction could induce several structural and functional changes resulting in leukocyte trafficking, which in turn, may shift the vascular equilibrium towards triggering more inflammation (Aird, 2003). Although leukocyte trafficking was known to play an essential part in the protective responses against any infection or injury, it may also lead to extensive tissue damage as shown in numerous inflammatory disorders (Chen et al., 2018). One of the most abundant leukocytes been assured in COVID-19 are neutrophils that represent the first line of defense in the innate immune system.

3.3 Neutrophils-mediated inflammation in COVID-19

With the continual reduction detected in lymphocytes count of COVID-19 patients, they become more prone for secondary infections with the risk of high mortality rate. This occurs due to loss of all lymphocyte effector cells that possess the essential antiviral activity, including CD8+ or cytotoxic lymphocytes and natural killer cells, as well as B cells, which able to form the specific antibodies targeted for inactivating the virus (Dallan et al., 2020; Remy et al., 2020).

Therefore, developing severe lymphopenia will effectively inhibit the stimulation of adaptive cell-mediated immune response and consequently, facilitate the inflammatory-mediated neutrophils response which could be started with their chemotaxis and recruitment, followed by degranulation (Hyun and Hong, 2017; Didangelos, 2020). Neutrophils possess an arsenal of proteases such as (elastase, proteinase-3 and cathepsin G), inflammatory mediators such as (TNF- α and IL-6), and toxic oxidants that do not kill phagocytosed pathogens only, but also can damage the host tissue (Gernez et al., 2010).

3.4 Inflammatory sepsis in COVID-19

In response to high neutrophilia with progressive lymphopenia established in COVID-19, viral sepsis may be promoted as a result of systemic uncontrolled inflammation induced by neutrophils with further worsening of tissue injury (Li et al., 2020a), that is in consistence with the final diagnosis emphasized the existence of a septic shock among COVID-19 patients with profound lymphopenia (Dallan et al., 2020).

Sepsis is a syndrome that has attracted the attention worldwide because of its high mortality rate of about 50–80 %. It is widely recognized as a kind of systemic inflammatory response that caused by severe bacterial infections and tissue injuries (Annane et al., 2005). Sepsis itself may share in the subsequent release of inflammatory factors (IL-6 and TNF- α) that could eventually aggravate the existing inflammation (Molano Franco et al., 2019).

Developing sepsis could lead to multiple organ dysfunction, shock, and even death, which are not caused directly by the invading pathogens; but as a result of inflammation (Crowther, 2001; Mantzarlis et al., 2017). During sepsis, there is an extensive crosslink between increased inflammation, endothelial dysfunction and hyper-coagulopathy, in which the microvascular dysfunction was documented to be one of important sepsis hallmarks (Schouten et al., 2008).

3.5 TGF-β1-induced pulmonary fibrosis in COVID-19

Giving the reported evidence of induced endothelial dysfunction, pulmonary fibrosis may be also prompted as a substantial problem during COVID-19 infection, to the extent that pulmonary post-mortem findings in fatal cases of COVID-19 revealed the presence of extensive fibrotic features as myofibroblastic proliferation or organizing pneumonia (George et al., 2020). The vascular endothelial dysfunction could stimulate the fibrotic consequences via secreting a peptide, namely endothelin-1 (ET-1) (Elshazly et al., 2013), which could induce the release of transforming growth factor- $\beta 1$ (TGF- $\beta 1$), a fibrogenic cytokine mainly implicated in driving the pulmonary fibrosis development (Wermuth et al., 2016).

3.6 ET-1-reduced cAMP in COVID-19

Surprisingly, ET-1 is also suggested to exaggerate the inflammation via inhibiting adenylate cyclase (AC) activity and thereby, cAMP accumulation (Insel et al., 2012).Within the immune system, cAMP is synthesized from ATP by the action of AC to regulate the anti-inflammatory effects (Gentile et al., 1988). As reported, cAMP could decrease the production of pro-inflammatory mediators as well as enhance the production of anti-inflammatory factors in various immune cells (Raker et al., 2016). Meanwhile, cAMP was concluded to promote ATP production that is described to potentially improve the efficiency of innate and adaptive immune systems for fighting off COVID-19 (De Rasmo et al., 2016; Taghizadeh-Hesary and Akbari, 2020).

Consistent with these findings, it was reported that COVID-19 may be more fatal in the elderly-population than in children, as with increasing the age, there is a gradual decline in the cellular ATP and subsequent ATP-induced cAMP accumulation (Srivastava, 2017). Furthermore, tobacco smokers, who suffer from a decreased content of ATP in immune cells, are also found to be more susceptible for COVID-19 infection (Malińska et al., 2019).

Regardless of age, males are generally more prone to die by COVID-19 than females (Jin et al., 2020). The finding which can be attributed to sex hormone differences, since estrogen was recorded to potentially induce ATP production during the inflammation than androgens (Kassi and Moutsatsou, 2010). Additionally, the same strategy could be particularly relevant for patients with seriously medical conditions, who showed an immune dysregulation as a result of ATP-depletion (Zhou et al., 2020).

With increasing the number of SARS-CoV-2- infected cases globally, there is unfortunately no time chance for discovering a newly therapeutic agent. Taken together, directing most efforts towards vaccine production may be of no avail at least nowadays, since millions of people everywhere have been already infected with COVID-19, and they are in urgent need for rapid treatment in order to prevent the disease progression. In addition, developing anti-viral drugs needs a long way to go. Therefore, the best choice may be repurposing the currently available drugs which may greatly save time and money as well as secure many people from death.

4. COVID-19 therapies

World Health Organization (WHO) reported that COVID-19 now becomes much more than a health crisis. Till present, curing COVID-19 remains elusive, despite of the great efforts directed by the researchers towards understanding and identifying the disease mechanisms. There is no doubt that COVID-19 can trigger airway inflammatory reactions, in which neutrophils play the major role in increasing the severity by inducing COVID-19-associated coagulopathy (Zuo et al.). In that context, several therapeutic strategies have been proposed to control COVID-19 (Cascella et al., 2020).

4.1 Current therapies

The most commonly one involves the use of hydroxychloroquine (HCQ) as the first-line therapy because of its anti-inflammatory and immunomodulatory effects (Hu et al., 2017). Based on the international guidelines, HCQ is reported to be utilized either alone or in combination with other drugs including, systemic corticosteroids, Tocilizumab (TCZ), macrolide Aazithromycin, antiviral Lopinavir/Ritonavir and anticoagulant Enoxaparin (Mehra et al., 2020; Rosenberg et al., 2020). However, the use of HCQ is lately recorded to have many restrictions due to increased risk of serious cardiac arrhythmias. Additionally, either HCQ or CQ is no longer authorized by FDA to treat COVID-19 (Joyce et al., 2013).

Moreover, current COVID-19 treatment protocol also recommends the use of oral anti-inflammatory steroids, however they may paradoxically exaggerate COVID-19-associated neutrophilia (Fukakusa et al., 2005). In addition, due to their local and systemic harmful adverse effects, steroids may be inappropriate drug of choice for vulnerable patients with pre-existing hypertension, diabetes, or cardiovascular diseases, which represent the most susceptible group to be infected with COVID-19 (Varga et al., 2020). That pushed clinicians to search for additional or alternative anti-inflammatory treatments that can efficiently control the neutrophilic component of COVID-19 apart from steroid related complications.

TCZ, a humanized monoclonal antibody acted by blocking IL-6 receptors, has been suggested for COVID-

19 patients to suppress the inflammatory storm and minimize the mortality (Fu et al., 2020). However, some studies showed that TCZ may effectively reduce both fever and inflammatory markers, but with no satisfactory clinical outcomes inferred for the critically ill COVID-19 patients (Campochiaro et al., 2020; Dastan et al., 2020). As documented, this medication may also raise both blood pressure and lipid levels, which are considered the main risk factors exaggerating the severity in COVID-19 patients of cardiovascular (CV) diseases (Rao et al., 2015). Furthermore, anti-interleukin therapy is expected to worse the post-COVID-19 pulmonary fibrosis (George et al., 2020; Silva et al., 2020).

As regards azithromycin, various clinical evidences revealed that it could exert a great role against both Acute Respiratory Distress Syndrome (SARS) and Middle East Respiratory Syndrome (MERS), that prompted scientists to strongly suggest it as a potential treatment for COVID-19. Azithromycin was detected to possess anti-inflammatory and immunomodulating actions in addition to antiviral properties because of its ability to minimize the production of pro-inflammatory cytokines particularly IL-6 and TNF- α , noxious oxidative radicals as well as to improve T-helper functions. However, the preliminary studies have demonstrated that using azithromycin should be in caution due to its potential arrhythmogenic threat, especially in highly risk COVID-19 patients (Pani et al., 2020).

Moreover, provision should be also taken to mitigate the cardiac risk, especially after adding lopinavir/ritonavir into the current treatment protocol for COVID-19 (Gérard et al., 2020). Lopinavir acts as anti-HIV protease inhibitor via inhibiting the action of 3CLpro, thus disrupting the viral replication and release from host cells. Recent *in-vitro* study indicates that lopinavir can also exhibit antiviral activity against SARS-CoV-2, with which ritonavir can be added as a booster. However, there is a contradictory survey concluded that the use of lopinavir/ritonavir shows no significant reduction in the mortality rate within the severely ill COVID-19 patients (Owa and Owa, 2020).

4.2 Potential COVID-19 therapies

Considering ACE-2 the only viral receptors, a new study has been proposed that Lactoferrin, an orally nutritional supplement, may be potentially useful against COVID-19. In addition to its unique immunomodulatory and anti-inflammatory effects, Lactoferrin has been stated to possibly occupy ACE-2 receptors preventing SARS-CoV-2 from attaching to the host cells (Kell et al., 2020), however it is not ensured clinically till now.

Most of repurposed drugs used for treating COVID-19 are directed mainly towards blocking the induced cytokine storm, however this COVID-19-related sepsis argues now for investigating a different therapeutic approach (Remy et al., 2020).

Since, the morbidity/mortality rate in septic patients was reported to be correlated with the plasma level of ET-1, reducing its level will minimize all unwanted reactions mediated by ET-1 receptors. The observation that may explain why anti-inflammatory drugs like anti- TNF- α and IL-1-based therapies have been failed in treating sepsis, opposite to clinical trials that suggested the application of ET-1 receptor blockers as an effective strategy (Kowalczyk et al., 2015). In addition, decreasing ET-1 level may interrupt the fibrotic pathway regulated by TGF- β 1; inhibiting the induction of pulmonary fibrosis.

Because ET-1 was previously reported to be one of the substrates that could be potentially degraded by endogenous NEP (neutral endopeptidase) (Abassi et al., 1992), that pushed us to predict that enhancing NEP activity may become a prerequisite to defeat COVID-19 ghost (El Tabaa and El Tabaa, 2020).

NEP is a type II integral transmembrane metallopeptidase, which was clearly detected in various tissues like lung, kidney, brain, intestine, and vascular endothelium (Li et al., 1995) as well as in many inflammatory cells including neutrophils (Connelly et al., 1985). In the airways, NEP has been found to be expressed in the epithelium (Sont et al., 1997), smooth muscle cells (Di Maria et al., 1998), and fibroblasts (Kletsas et al., 1998).

NEP also has a high cleaving affinity towards some potent inflammatory and vasoactive peptides other than ET-1 including bradykinins (BKs), N-formyl-L-methionyl- L-leucyl-L-phenylalanine (fMLP) and atrial natriuretic peptide (ANP); that emphasize on its role in alleviating the airway inflammatory processes (Connelly et al., 1985; Shimamoto et al., 1994).

Several studies ensured that destroying or down regulating NEP may lead to further pathophysiological changes. This involves an increase in vascular permeability, recruitment and activation of inflammatory cells, particularly neutrophils. Neutrophils chemotaxis will lead to the release of neutrophil elastase enzymes (e.g., Cathepsin G), which may exert further destructive effects on airway tissues, leading to worsening and progression of the disease (Borson, 1991).

Therefore, reducing NEP activity either by cigarette smoking (Dusser et al., 1989), hypoxia (Carpenter and Stenmark, 2001) or respiratory pathogens like parainfluenza virus type 1, rat corona-virus, and Mycoplasma pulmonis (Jacoby et al., 1988; Borson et al., 1989), will be a clear explanation for their associated inflammatory cascades. Considering multiple activities of NEP in regulating local inflammatory neuropeptides within alveolar microenvironment and nearby vascular cells (Wick et al., 2011), it may exhibit a good target for counteracting the airway inflammation, coagulopathy and pulmonary fibrosis associated with COVID-19 infection.

Referring to the studies searching for agents that may up-regulate NEP gene expression; enhancing its activity and promoting its action (Borson, 1991), a variety of selective enhancers are pre-clinically developed involving drugs (glucocorticoids) (Borson and Gruenert, 1991), hormones (androgens (Yao et al., 2008) and estrogen (Xiao et al., 2009)) or natural products (Apigenin, Luteolin, and Curcumin, (-)-Epigallocatechin-3-gallate and Resveratrol) (Ayoub and Melzig, 2008; Chang et al., 2015; El-Sayed and Bayan, 2015).

Along with, Rolipram, an investigative PDE4i, has also been examined; suggesting that increasing the levels of intracellular cAMP was directly correlated with enhanced NEP activity, which in turn may prolong and potentiate the cAMP-mediated short-term anti-inflammatory mechanism (Graf et al., 1995; Ayoub and Melzig, 2008).

That outcome may declare that roflumilast, a highly selective PDE4i, can exert its efficient anti-inflammatory effect via enhancing cAMP level as well as NEP activity. Accordingly, we predict that roflumilast may be one of the most useful drugs that is expected to play a great role in treating COVID-19. However, until this moment, no study declares the potential fundamental pathways contributing to relying roflumilast on NEP activity.

5. Roflumilast overview

Roflumilast is recorded to be a highly selective long-acting inhibitor of PDE4 isoenzyme, to which its use will be surely accompanied with an increase in the level of intracellular cAMP (Rabe, 2011).

5.1 Phosphodiesterase enzymes (PDEs)

Phosphodiesterase enzymes (PDEs) are a large superfamily of enzymes that catalyze the hydrolysis of secondary messengers such as cAMP and cyclic guanosine mono-phosphate (cGMP) into their inactive 5' monophosphate; thus regulating their intracellular level as well as the amplitude and duration of their signaling (Hertz et al., 2009).

Based on amino acid sequences, tissue distribution and pharmacological properties, PDEs could be classified into 11 families, namely PDE1-PDE11. Similarly, PDEs can be also grouped into three categories according to their substrate specificities including, cAMP-selective hydrolases (PDE4, 7 and 8), cGMP-selective hydrolases (PDE5, 6, and 9) and hydrolases for both cAMP and cGMP (PDE1, 2, 3, 10, and 11) (Azevedo et al., 2014).

Regarding PDE4, it was accounted to represent the predominant isoenzyme responsible for regulating cAMP levels in many cell types within the lung including airway epithelial cells, airway smooth muscle cells and pulmonary vascular endothelium. PDE4 was also noticed to be widely distributed in various inflammatory cells, like neutrophils, T lymphocytes, eosinophils, monocytes and basophils (van Schalkwyk et al., 2005; Halpin, 2008).

Notably, cAMP has a direct significant role in different inflammatory pathways via inhibiting ROS generation and pro-inflammatory cytokines production, mainly TNF- α and IL-6; (Shames et al., 2001; Isoni et al., 2009). cAMP could also promote the production of anti-inflammatory mediators such as IL-10 which was identified as a "cytokine synthesis inhibitory factor", and acted as a principle regulator in the JAK-STAT signaling pathway (Redford et al., 2011). Therefore, elevating cAMP level within the pulmonary tissue, vascular and inflammatory cells can provide an efficient anti-inflammatory action (Li et al., 2018).

On the other hand, it was found that the capacity of PDEs for cAMP hydrolysis is greater than the maximum rate of its synthesis. Therefore, minute reduction in PDEs activity can result in high elevation in cAMP level with significant changes in the activity of its dependent protein kinase (Halpin, 2008). The notice that pushed scientists since 1970 to investigate the potential therapeutic importance of inhibiting PDE4 activity (Weiss and Hait, 1977).

5.2. Selective and non-selective PDE4i

Because of the involvement of cAMP signaling in the pathophysiology of many inflammatory diseases, it has been proved that targeting PDE4 will resemble an effective therapeutic strategy for different inflammatory conditions, as chronic obstructive pulmonary disease (COPD), asthma, atopic dermatitis (AD), inflammatory bowel diseases (IBD), rheumatic arthritis (RA), lupus and neuroinflammation (Li et al., 2018).

Early, nonselective PDE inhibitors were discovered including theophylline and doxofylline, but, because of their associated significant adverse effects, their use had been limited.

Given that PDE4 is the only cellular pathway available for cAMP degradation (Fertig, Bracy A., 2018), therapeutic studies have been directed to develop the most selective PDE4 inhibitors, among which, Apremilast and Roflumilast are currently available (Boswell-Smith et al., 2006; Kumar et al., 2013).

6. Pharmacotherapeutic effects of Roflumilast

Since 2011, roflumilast has been approved by FDA as an anti-inflammatory drug specifically designed for many respiratory disorders mainly COPD and asthma. By time, roflumilast has been reported to exert different pharmacological activities, **Figure 2 and Table 1** (Li et al., 2018).

6.1 Roflumilast and lung inflammation

Clinical trials reported that oral administration of roflumilast could suppress airway inflammation and improve lung function of COPD patients. In addition, it is documented to be effective in reducing the frequency of disease exacerbations when given as add-on to inhaled therapy in patients with moderate or severe COPD (Shen et al., 2018). As regards asthmatic patients, roflumilast could also significantly increase the forced expiratory volume in 1 s (FEV₁) and improved airway inflammation (Bateman et al., 2006).

The anti-inflammatory mechanisms of roflumilast can be contributed to its PDE4 inhibiting activity; leading to an increase in cAMP concentration and signaling within the epithelial airway and inflammatory cells. The action which in turn will enable roflumilast to suppress the expression of pro-inflammatory cytokines as IL-6 and TNF- α (Feng et al., 2017). Moreover, another study of cigarette smoke-induced pulmonary inflammation in guinea pigs showed that roflumilast could effectively reduce the numbers of neutrophils, lymphocytes and eosinophils in bronchoalveolar lavage fluid (Fitzgerald et al., 2006).

For COPD patients, roflumilast was represented to exert a significant role in reducing eosinophil cell counts within their bronchial biopsy samples and sputum (Rabe et al., 2018), in addition to its direct suppressing effect on neutrophils function and their ROS production. As a result of elevating cAMP level, roflumilast could inhibit neutrophil chemotaxis and degranulation. cAMP could directly activate protein of Epac1, which in turn could suppress neutrophil migration as well as oxidative burst. Furthermore, cAMP could also activate protein kinase A (PKA) in neutrophils, leading to a decline in their phagocytic activity (Dunne et al., 2019).

Some *in-vivo* and *in-vitro* studies revealed that roflumilast can potently reduce the endothelial permeability

and suppress the leukocyte–endothelial cell interactions through altering the expression of adhesion molecules and attenuating the upregulation of PMNL surface CD11b, that may be stimulated either by fMLP or platelet-activating factor (PAF). The action that could inhibit neutrophils adhesion to endothelial cells (Sanz et al., 2007). Additionally, results from *in-vitro* studies of human neutrophils showed that roflumilast could prevent the release of neutrophil elastase, matrix metalloproteinase and myeloperoxidase, inhibiting neutrophil function (Jones et al., 2005)

A synergistic effect of roflumilast with other anti-inflammatory agents such as corticosteroids or long-acting β 2-agonists have been demonstrated (Kawamatawong, 2017). It was concluded that roflumilast-N-oxide (RNO), the active metabolite of roflumilast, could enhance the anti-inflammatory effect of dexamethasone in airway smooth muscle cells *in-vitro* (Patel et al., 2017). At the same time, roflumilast was reported to reverse the corticosteroid-associated insensitivity towards neutrophils in COPD patients (Milara et al., 2015b). As well, other study revealed the great value of roflumilast in restoring the glucocorticoid sensitivity in glucocorticoid-resistant patients through blocking the downregulation of glucocorticoid receptor alpha (GR α), which was known to be responsible for glucocorticoid resistance (Reddy et al., 2020).

6.2 Roflumilast and hypercoagulable states

Neutrophils and platelets have been identified as crucial factors for thrombus initiation and progression. Both animal models and human diseases increased the evidence that neutrophils extracellular traps (NETs) possess a significant role in the pathogenesis of thrombosis. NETs were detected to be released from the activated neutrophils in a process called NETosis, which can be mediated by recruitment of both platelets and PMN into the endothelial wall. Then, NETs could stimulate platelets adhesion, activation and aggregation with subsequent activation of coagulation cascades to trigger thrombosis (Fuchs et al., 2010; Kimball et al., 2016).

Accordingly, inhibiting the prothrombotic function of neutrophils and interfering with NETs formation, by roflumilast, could reduce the risk of thrombosis in COPD as well as in other inflammatory diseases. Moreover, RNO (an active metabolite of roflumilast) was recorded to affect NETs via inhibiting Src family kinases phosphoinositide 3-kinase (SFK–PI3K) pathway in PMNs. In addition, RNO could block the key biochemical mechanisms regulating PMN–platelets adhesion (*Totani et al., 2016*).

6.3 Roflumilast and inflammatory sepsis

Previous findings stated that Janus kinase (JAK)/Signal transducer and activator of transcription-3 (STAT-3) was a key cellular signal transduction pathway proved to mediate the expression of many inflammatory cytokines produced during sepsis (Cai et al., 2015). That pathway resembles a positive feed-back signal for exacerbating the inflammatory response, resulting in uncontrolled systemic inflammation (Chang et al., 2019).

Moreover, during sepsis, there is also an inflammation-induced activation of coagulation as a result of the concomitant impairment of endothelial function, anticoagulant and fibrinolytic systems, indicating that systemic inflammation will be the main pathological reaction of sepsis and the major cause for associated multiple organ failure (Schouten et al., 2008). Therefore, reducing inflammation could be the key for treating sepsis.

Regarding the role of roflumilast in suppressing the mRNA expression of JAK/STAT-3 signaling pathway with subsequent inhibition of inflammatory cytokine release (e.g. IL-6 and TNF- α) in the lung tissue of septic mice model (Chang et al., 2019), there is a proof of its protecting effect against sepsis through the above-referred anti-inflammatory and anti-thrombotic activities.

6.4. Roflumilast and lung fibrosis

Because of the potential effect of anti-inflammatory treatment to mitigate airway fibrotic remodeling, roflumilast might have anti-fibrotic role due to its well-known anti-inflammatory action (Hatzelmann et al., 2010). Roflumilast was found to have the ability to prevent the progressive airways fibrosis, as a result of antagonizing fibroblasts activity, which could be mediated by TGF- β 1, an essential regulator of immune responses related to fibrosis (Togo et al., 2009). Anti-fibrotic profile of roflumilast could be also explained by its ability to reduce the expression of upregulated NADPH oxidase 4 (NOX4) (Milara et al., 2015c), which was indicated to be critical for pulmonary fibrotic remodeling (Amara et al., 2010).

Within this regard, roflumilast could also normalize most of increased metabolic changes like alterations in oxidative equilibrium, increased collagen and protein synthesis; resulting in decline in the fibrotic score. Simultaneously, reduced lung tissue pH has been proposed as a risk factor for lung fibrosis development, which was also reported to be corrected by roflumilast in bleomycin model of pulmonary fibrosis (Milara et al., 2015a).

7. Adverse effects and safety of Roflumilast

Roflumilast can be safely administered as it is not associated with the parlous induction of adverse effects involving seizures and cardiac arrhythmias; in addition, its elimination is not significantly altered by several drug classes or even by food and tobacco smoking (Gupta and O'Mahony, 2008).

Results from clinical trials demonstrated that the only use limitations reported for roflumilast were nausea, vomiting, diarrhea, weight loss and headache (Baye, 2012). These effects were appeared to be dose dependent and transient which in turn did not need treatment discontinuation (van Schalkwyk et al., 2005).

Great efforts have been made to limit the gastrointestinal adverse reactions and to provide a better benefit (Li et al., 2018). Thus, for improving patient tolerability, a study in the allergen-challenged Brown Norway rats, has been performed to evaluate the efficacy of inhaled roflumilast given either intratracheally or by nasal inhalation. As concluded, the inhaled form showed a powerful effect on improving the lung function (Chapman et al., 2007), supporting the therapeutic importance of using inhaled PDE4i against inflammatory lung diseases, which may be then more efficacious and less adverse effects than its oral forms, however it is still under clinical trial (Rhee and Kim, 2020).

8. Roflumilast in aging and diabetic comorbidities

During physiological aging process, a low-grade chronic systemic inflammation, called inflammaging, develops and impairs the maintenance of immunological homeostasis, in which there are high levels of C-reactive protein (CRP), proinflammatory cytokines as IL-6, in addition to low level of anti-inflammatory cytokines as IL-10 (Franceschi et al., 2018). PDE4 enzymes play a major role against inflammaging by increasing cAMP which in turn stimulates AMP-activated Protein Kinase (AMPK), exerting an anti-inflammatory effect. Since PDE4 enzyme activity in elderly individuals is greater than their youngers, using roflumilast, can experience a relatively more increase in cAMP level and as a consequence, potentiate its anti-inflammatory action in old age people (Muo et al., 2019).

Given the essential role of PDE4 in glucose and fat metabolism, roflumilast, through PDE4 inhibition, could prevent the disease progression in diabetes mellitus (DM) type 2 patients via improving the glycemic index. Roflumilast could encourage the secretion of intestinal glucagon like peptide-1 (GLP-1), which is a main incretin and potent insulinotropic agent for stimulating insulin secretion from the β -cells of pancreas (Wouters et al., 2012). In addition, it was documented that a deficiency in PDE4B could attenuate high-fat diet-induced adiposity and adipose tissue inflammation in mice (Vollert et al., 2012), referring to the role of roflumilast in reducing weight and improving insulin sensitivity in adults with prediabetes and/or obesity (Muo et al., 2019).

9. Roflumilast and COVID-19 infection

The rationale for selecting PDE4i for COVID-19 may be based on the previous findings demonstrating that inhibiting the activity of PDE4 will suppress a myriad of pro-inflammatory responses (Press and Banner, 2009). Inhibiting PDE4 will specifically prevent cAMP degradation, which in turn will decrease airway inflammation via preventing the activation and recruitment of inflammatory cells, specifically neutrophils as well as cytokines production (Barnette, 1999). The observation that drives scientists to attractively target PDE4 for treating COVID-19.

In addition to its anti-inflammatory, anti-coagulant ant anti-diabetic roles, roflumilast could be used safely in a combination with corticosteroids, recommended to be used effectively against COVID-19 infection, by improving their compromised anti-inflammatory properties and their resistance effect (Milara et al., 2015b; Wang et al., 2016).

At the same time, azithromycin, a macrolide antibiotic suggested for COVID-19 treatment, was documented to exhibit a lower affinity for cytochrome P-450A (CYP) 3A4 CYP 3A4. Thus, azithromycin would poorly interact with roflumilast because this cytochrome member resembles the main metabolic pathway for roflumilast (Westphal, 2000).

A little while ago, roflumilast is predicted to exert anti-viral effect similar to that of lopinavir/ritonavir via binding very close to the middle pocket of SARS-CoV-2 3CLpro and thereby, interfering with its activity (Hu et al., 2020). Then, roflumilast can deprive the virus from hydrolyzing the polyprotein into functional proteins required for its replication, **Figure 3** (He et al., 2020). However, the preventive and therapeutic effectiveness of roflumilast against COVID-19 and its pharmacological mechanisms have not been yet extensively studied.

10. NEP-based strategy for treating COVID-19 by

Roflumilast

One of the proposed NEP-dependent mechanisms for blocking the airway inflammation is to cleave the neutrophil-released cathepsin G, that is documented to convert both Angiotensinogen and Ang I into Ang II,**Figure 4** (Wintroub et al., 1984; Pham, 2006; Meyer-Hoffert, 2009). In response to severe COVID-19 infection, Ang II is reported to be continuously generated and probably motivates the systemic cytokine storm (Xiong et al., 2020). Among the released cytokines, IL-6 will play a vital role in the progression of numerous inflammatory reactions as well as endothelial dysfunction and platelets activation (Funakoshi et al., 1999; Liu et al., 2020c). Therefore, cleaving cathepsin G by NEP with reducing associated Ang II formation may be a logical commentary for the suppressed IL-6 expression detected following roflumilast treatment (Feng et al., 2017).

Postulating that IL-6 may be a key regulator of COVID-19 pathogenesis (Liu et al., 2020b), decreasing its level by roflumilast will be of great importance. Firstly, roflumilast can stop IL-6-mediated intestinal, olfactory and ocular inflammation and consequently, inhibit the induction of anosmia, diarrhea and conjunctivitis, respectively. Secondly, roflumilast may suppress the endothelial activation and inflammatory thrombocytosis prompted by IL-6 release.

As a result of the endothelial dysfunction, neutrophils trafficking has also been implicated in the pathogenesis of COVID-19, since their activation and accumulation are reported to be associated with tissue damage, exaggerated inflammation and disordered tissue repair (Tay et al., 2020). As such, NEP can degrade the chemoattractant fMLP, which was known to be involved in neutrophils chemotaxis. Hence, NEP may specifically prevent the recruitment of neutrophils across endothelial barrier from the blood circulation into the infected tissues (Sato et al., 2013). In particular, the potential role of roflumilast in inhibiting the adhesion and transmigration of neutrophils and their subsequent inflammatory sepsis may be attributable to increased NEP activity (Sanz et al., 2007; Li et al., 2020a).

Additionally, NEP was recorded to effectively breakdown the endothelium-derived ET-1; preventing the activation and aggregation of platelets as a result of prohibiting the synthesis of PAF (Rao and White, 1982; Mustafa et al., 1995), which was previously demonstrated to be also suppressed by the action of PDE4i (Tenor et al., 1996). Accordingly, this observation may reflect the potential NEP-dependent anti-coagulant role of roflumilast against the thromboembolic events in COVID-19; empowering it to restrain the development of PIC which is the initial step for evolving stroke in COVID-19 patients (Avula et al., 2020).

In line, it was also shown that COVID-19 patients may show pulmonary fibrosis, from which NEP may

protect lungs by stopping the ET-1-induced TGF- β 1; ensuring the concept that roflumilast may have the potential to attenuate the fibroblast activities and thereby, the ability to function as anti-fibrotic agent via blocking the fibrosis driven by TGF- β 1 (Dunkern et al., 2007; Togo et al., 2009).

Because cAMP is underscored to play an important role in improving the immune system of highly risk COVID-19 groups, breaking ET-1 by NEP will also maintain the high level of cAMP which may contribute for long-term anti-inflammatory effect of roflumilast (Graf et al., 1995; Raker et al., 2016).

Accordingly, we recommend that future clinical efforts should be driven towards ensuring the NEP-mediated pharmacotherapeutic mechanisms of roflumilast proposed for counteracting COVID-19 infection.

11. Conclusion

Reducing the patient's risk of COVID-19 progression is assumed to be biologically linked with suppression of neutrophilic component that predisposes to increased systemic inflammation and coagulopathy associated with COVID-19 infection. Therefore, management of COVID-19 should focus on modulating neutrophils function and their response. According to the underlying guidelines, recommended anti-inflammatory therapies for COVID-19 do not provide treatment satisfaction and effectiveness until now.

As the search continues, PDE4i has been suggested to offer an intriguing new class of COVID-19 treatment, since inhibiting PDE4 is thought to exhibit effective anti-inflammatory and anti-platelets activities. Among the clinically used PDE4i, roflumilast has been reported to be the most selective and potent drug submitted for treating many neutrophils-mediated airway inflammatory disorders. Furthermore, roflumilast has been recently reported to behave as a potential inhibitor of 3CLpro, which is a proteolytic enzyme required for viral replication within the host cells.

Considering COVID-19 treatment, roflumilast may also have additive advantages to the concurrent protocol, since it had been reported to be used safely in combination with either corticosteroids, azithromycin and recommended vitamins (C, E and Zinc) without showing any dangerous adverse effects up till now. As well, via attenuating the airway neutrophilic inflammation, roflumilast can enhance the compromised anti-inflammatory properties of corticosteroids and improve their resistance effect.

Additionally, as a result of increasing cAMP level, we suppose that roflumilast can prolong its antiinflammatory effect and display other therapeutic properties via enhancing NEP activity, which is proposed to be an important target for managing COVID-19.

Therefore, taken into our consideration that this review is the first one to discuss the NEP-mediated therapeutic properties of roflumilast and its role in facing the inflammatory, coagulopathy and fibrotic cascades driven by COVID-19, we hope that our hypothesis will serve as a stimulus for further confirmation about the therapeutic impact of roflumilast in COVID-19 management and consequently, may provide physicians with a novel repurposed treatment option against COVID-19.

Nomenclature of targets and ligands

Key protein targets and ligands in this article are hyperlinked to corresponding entries in http://www.guidetopharmacology.org, the common portal for data from the IUPHAR/BPS Guide to PHAR-MACOLOGY (Harding et al., 2018), and are permanently archived in the Concise Guide to PHARMACOL-OGY 2019/2020 (Alexander et al., 2019).

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Conflict of interest

The authors declare no conflict of interest. The authors and their institutions are the only responsible for the financial support and the content of this work in the submitted manuscript. All other authors have no conflict of interests to disclose.

References

Abassi, Z.A., Tate, J.E., Golomb, E., and Keiser, H.R. (1992). Role of neutral endopeptidase in the metabolism of endothelin. Hypertension 20 : 89–95.

Aird, W.C. (2003). The role of the endothelium in severe sepsis and multiple organ dysfunction syndrome. Blood 101 : 3765–3777.

Alexander, S.P.H., Christopoulos, A., Davenport, A.P., Kelly, E., Mathie, A., Peters, J.A., et al. (2019). THE CONCISE GUIDE TO PHARMACOLOGY 2019/20: G protein-coupled receptors. Br. J. Pharma-col. 176 : S21–S141.

Amara, N., Goven, D., Prost, F., Muloway, R., Crestani, B., and Boczkowski, J. (2010). NOX4/NADPH oxidase expression is increased in pulmonary fibroblasts from patients with idiopathic pulmonary fibrosis and mediates TGFbeta1-induced fibroblast differentiation into myofibroblasts. Thorax 65 : 733–738.

Annane, P.D., Bellissant, P.E., and Cavaillon, J.M. (2005). Septic shock. In Lancet, pp 63–78.

Avula, A., Nalleballe, K., Narula, N., Sapozhnikov, S., Dandu, V., Toom, S., et al. (2020). COVID-19 presenting as stroke. Brain. Behav. Immun.

Ayoub, S., and Melzig, M.F. (2008). Influence of Selected Natural Products on Neutral Endopeptidase Activity and β-Amyloid Production in SK-N-SH Cells. Pharm. Biol. 46 : 425–432.

Azevedo, M.F., Faucz, F.R., Bimpaki, E., Horvath, A., Levy, I., Alexandre, R.B. de, et al. (2014). Clinical and molecular genetics of the phosphodiesterases (PDEs). Endocr. Rev. 35 : 195–233.

Barnette, M.S. (1999). Phosphodiesterase 4 (PDE4) inhibitors in asthma and chronic obstructive pulmonary disease (COPD). Prog. Drug Res. 53 : 193–229.

Bateman, E.D., Goehring, U.-M., Richard, F., and Watz, H. (2016). Roflumilast combined with montelukast versus montelukast alone as add-on treatment in patients with moderate-to-severe asthma. J. Allergy Clin. Immunol. 138 : 142–149.

Bateman, E.D., Izquierdo, J.L., Harnest, U., Hofbauer, P., Magyar, P., Schmid-Wirlitsch, C., et al. (2006). Efficacy and safety of roflumilast in the treatment of asthma. Ann. Allergy, Asthma Immunol. *96* : 679–686.

Baye, J. (2012). Roflumilast (daliresp): a novel phosphodiesterase-4 inhibitor for the treatment of severe chronic obstructive pulmonary disease. P T 37 : 149–61.

Belen-Apak, F.B., and Sarialioğlu, F. (2020). Pulmonary intravascular coagulation in COVID-19: possible pathogenesis and recommendations on anticoagulant/thrombolytic therapy. J. Thromb. Thrombolysis *May* : 1–3.

Borson, D.B. (1991). Roles of neutral endopeptidase in airways. Am. J. Physiol. - Lung Cell. Mol. Physiol. 260 : L212-25.

Borson, D.B., Brokaw, J.J., Sekizawa, K., McDonald, D.M., and Nadel, J.A. (1989). Neutral endopeptidase and neurogenic inflammation in rats with respiratory infections. J. Appl. Physiol. *66* : 2653–2658.

Borson, D.B., and Gruenert, D.C. (1991). Glucocorticoids induce neutral endopeptidase in transformed human tracheal epithelial cells. Am. J. Physiol. - Lung Cell. Mol. Physiol. 260 : L83-9.

Boswell-Smith, V., Cazzola, M., and Page, C.P. (2006). Are phosphodiesterase 4 inhibitors just more theophylline? J. Allergy Clin. Immunol. 117 : 1237–1243.

Bridgewood, C., Damiani, G., Sharif, K., and Quartuccio, L. (2020). Rationale for use of PDE4 inhibition for severe inflammation in COVID-19 Pneumonia. *323* : 1824–1836.

Burgess, J.K., Oliver, B.G.G., Poniris, M.H., Ge, Q., Boustany, S., Cox, N., et al. (2006). A phosphodiesterase 4 inhibitor inhibits matrix protein deposition in airways in vitro. J. Allergy Clin. Immunol. 118 : 649–657.

Cai, B., Cai, J., Luo, Y., Chen, C., and Zhang, S. (2015). The specific roles of JAK/STAT signaling pathway in sepsis. Inflammation 38 : 1599–1608.

Campochiaro, C., Della-Torre, E., Cavalli, G., Luca, G. De, Ripa, M., Boffini, N., et al. (2020). Efficacy and safety of tocilizumab in severe COVID-19 patients: a single-centre retrospective cohort study. Eur. J. Intern. Med. 76 : 43–49.

Carpenter, T.C., and Stenmark, K.R. (2001). Hypoxia decreases lung neprilysin expression and increases pulmonary vascular leak. Am. J. Physiol. - Lung Cell. Mol. Physiol. 281 : L941-48.

Cascella, M., Rajnik, M., Cuomo, A., Dulebohn, S.C., and Napoli, R. Di (2020). Features, Evaluation and Treatment Coronavirus (COVID-19) (StatPearls Publishing).

Chang, X., Hu, L.F., Ma, X.J., Yin, J., Liu, X.Y., and Li, J.B. (2019). Influence of roflumilast on sepsis mice through the JAK/STAT signaling pathway. Eur. Rev. Med. Pharmacol. Sci. 23 : 1335–1341.

Chang, X., Rong, C., Chen, Y., Yang, C., Hu, Q., Mo, Y., et al. (2015). (-)-Epigallocatechin-3-gallate attenuates cognitive deterioration in Alzheimer's disease model mice by upregulating neprilysin expression. Exp. Cell Res. 334 : 136–145.

Chapman, R.W., House, A., Jones, H., Richard, J., Celly, C., Prelusky, D., et al. (2007). Effect of inhaled roflumilast on the prevention and resolution of allergen-induced late phase airflow obstruction in Brown Norway rats. Eur. J. Pharmacol. 571 : 215–221.

Chen, L., Deng, H., Cui, H., Fang, J., Zuo, Z., Deng, J., et al. (2018). Inflammatory responses and inflammation-associated diseases in organs. Oncotarget 9 : 7204–7218.

Connelly, J.C., Skidgel, R.A., Schulz, W.W., Johnson, A.R., and Erdös, E.G. (1985). Neutral endopeptidase 24.11 in human neutrophils: Cleavage of chemotactic peptide. Proc. Natl. Acad. Sci. U. S. A. 82 : 8737–8741.

Cox, M.J., Loman, N., Bogaert, D., and O'Grady, J. (2020). Co-infections: potentially lethal and unexplored in COVID-19. The Lancet Microbe 1 : e11.

Crowther, M.A. (2001). Continuing Challenges of Sepsis Research. JAMA 286 : 1894.

CSSE, J.H. (2020). Index @ Gisanddata.Maps.Arcgis.Com.

ArcGIS. Johns Hopkins University. Retrieved 30 June 2020.

Cui, S., Chen, S., Li, X., Liu, S., and Wang, F. (2020). Prevalence of venous thromboembolism in patients with severe novel coronavirus pneumonia. J. Thromb. Haemost.

Dal Moro, F., and Livi, U. (2020). Any possible role of phosphodiesterase type 5 inhibitors in the treatment of severe COVID19 infections? A lesson from urology. Clin. Immunol. 214 : 108414.

Dallan, C., Romano, F., Siebert, J., Politi, S., Lacroix, L., and Sahyoun, C. (2020). Septic shock presentation in adolescents with COVID-19. Lancet Child Adolesc. Heal.

Dastan, F., Nadji, S.A., Saffaei, A., and Tabarsi, P. (2020). Tocilizumab administration in a refractory case of COVID-19. Int. J. Antimicrob. Agents 106043 :.

Didangelos, A. (2020). Neutrophil involvement in Covid-19. Preprints.

Driggin, E., Madhavan, M. V., Bikdeli, B., Chuich, T., Laracy, J., Biondi-Zoccai, G., et al. (2020). Cardiovascular Considerations for Patients, Health Care Workers, and Health Systems During the COVID-19 Pandemic. J. Am. Coll. Cardiol. 75 : 2352–2371.

Dunkern, T.R., Feurstein, D., Rossi, G.A., Sabatini, F., and Hatzelmann, A. (2007). Inhibition of TGF- β induced lung fibroblast to myofibroblast conversion by phosphodiesterase inhibiting drugs and activators of soluble guanylyl cyclase. Eur. J. Pharmacol. 572 : 12–22.

Dunne, A.E., Kawamatawong, T., Fenwick, P.S., Davies, C.M., Tullett, H., Barnes, P.J., et al. (2019). Direct Inhibitory Effect of the PDE4 Inhibitor Roflumilast on Neutrophil Migration in Chronic Obstructive Pulmonary Disease. Am. J. Respir. Cell Mol. Biol. 60 : 445–453.

Dusser, D.J., Djokic, T.D., Borson, D.B., and Nadel, J.A. (1989). Cigarette smoke induces bronchoconstrictor hyperresponsiveness to substance P and inactivates airway neutral endopeptidase in the guinea pig. Possible role of free radicals. J. Clin. Invest. 84 : 900–906.

El-Sayed, N.S., and Bayan, Y. (2015). Possible role of Resveratrol targeting Estradiol and Neprilysin pathways in Lipopolysaccharide model of Alzheimer disease. Adv. Exp. Med. Biol. 822 : 107–118.

Elshazly, M., Hosny, H., Abdel-Hafiz, H., Zakaria, A., Elkaffas, K., and Okasha, N. (2013). Assessment of endothelial dysfunction in idiopathic pulmonary fibrosis. Egypt. J. Chest Dis. Tuberc. 62 : 589–592.

Feng, H., Chen, J., Wang, H., Cheng, Y., Zou, Z., Zhong, Q., et al. (2017). Roflumilast reverses polymicrobial sepsis-induced liver damage by inhibiting inflammation in mice. Lab. Investig. 97 : 1008–1019.

Fertig, Bracy A., and G.S.B. (2018). PDE4-Mediated cAMP Signalling. J. Cardiovasc. Dev. Dis. 5:8.

Fitzgerald, M.F., Spicer, D., McAulay, A.E., Wollin, L., and Beume, R. (2006). Roflumilast but not methylprednisolone inhibited cigarette smoke-induced pulmonary inflammation in guinea pigs. Eur. Respir. J. Suppl.

Franceschi, C., Garagnani, P., Parini, P., Giuliani, C., and Santoro, A. (2018). Inflammaging: a new immune-metabolic viewpoint for age-related diseases. Nat. Rev. Endocrinol. 14 : 576–590.

Fu, B., Xu, X., and Wei, H. (2020). Why tocilizumab could be an effective treatment for severe COVID-19? J. Transl. Med. 18 : 164.

Fuchs, T.A., Brill, A., Duerschmied, D., Schatzberg, D., Monestier, M., Myers, D.D.J., et al. (2010). Extracellular DNA traps promote thrombosis. Proc. Natl. Acad. Sci. U. S. A. 107 : 15880–15885.

Fukakusa, M., Bergeron, C., Tulic, M.K., Fiset, P.O., Dewachi, O. Al, Laviolette, M., et al. (2005). Oral corticosteroids decrease eosinophil and CC chemokine expression but increase neutrophil, IL-8, and IFN-γ-inducible protein 10 expression in asthmatic airway mucosa. J. Allergy Clin. Immunol. 115 : 280–286.

Funakoshi, Y., Ichiki, T., Ito, K., and Takeshita, A. (1999). Induction of interleukin-6 expression by angiotensin II in rat vascular smooth muscle cells. Hypertension 34: 118–125.

Gauvreau, G.M., Boulet, L.P., Schmid-Wirlitsch, C., Côté, J., Duong, M.L., Killian, K.J., et al. (2011). Roflumilast attenuates allergen-induced inflammation in mild asthmatic subjects. Respir. Res. 12 : 140.

Gentile, F., Raptis, A., Knipling, L.G., and Wolff, J. (1988). Extracellular cAMP formation from host cell ATP by Bordetella pertussis adenylate cyclase. BBA - Mol. Cell Res. 971 : 63–71.

George, P.M., Wells, A.U., and Jenkins, R.G. (2020). Pulmonary fibrosis and COVID-19: the potential role for antifibrotic therapy. Lancet Respir. Med.

Gérard, A., Romani, S., Fresse, A., Viard, D., Parassol, N., Granvuillemin, A., et al. (2020). "Off-label" use of hydroxychloroquine, azithromycin, lopinavir-ritonavir and chloroquine in COVID-19: A survey of cardiac adverse drug reactions by the French Network of Pharmacovigilance Centers. Therapies. Gernez, Y., Tirouvanziam, R., and Chanez, P. (2010). Neutrophils in chronic inflammatory airway diseases: Can we target them and how? Eur. Respir. J. 35 : 467–469.

Ghasemi, H. (2018). Roles of IL-6 in Ocular Inflammation: A Review. Ocul. Immunol. Inflamm. 26: 37-50.

Graf, K., Kunkel, K., Zhang, M., Gräfe, M., Schultz, K., Schudt, C., et al. (1995). Activation of adenylate cyclase and phosphodiesterase inhibition enhance neutral endopeptidase activity in human endothelial cells. Peptides 16 : 1273–1278.

Growcott, E.J., Spink, K.G., Ren, X., Afzal, S., Banner, K.H., and Wharton, J. (2006). Phosphodiesterase type 4 expression and anti-proliferative effects in human pulmonary artery smooth muscle cells. Respir. Res. 7:.

Gupta, P., and O'Mahony, M.S. (2008). Potential adverse effects of bronchodilators in the treatment of airways obstruction in older people: Recommendations for prescribing. Drugs and Aging 25 : 415–443.

Halpin, D.M.G. (2008). ABCD of the phosphodiesterase family: interaction and differential activity in COPD. Int. J. Chron. Obstruct. Pulmon. Dis. 3 : 543–561.

Hamming, I., Timens, W., Bulthuis, M.L.C., Lely, A.T., Navis, G.J., and Goor, H. van (2004). Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis. J. Pathol. 203 : 631–637.

Harding, S.D., Sharman, J.L., Faccenda, E., Southan, C., Pawson, A.J., Ireland, S., et al. (2018). The IUPHAR/BPS Guide to PHARMACOLOGY in 2018: updates and expansion to encompass the new guide to IMMUNOPHARMACOLOGY. Nucleic Acids Res. 46 : 1091–1106.

Hatzelmann, A., Morcillo, E.J., Lungarella, G., Adnot, S., Sanjar, S., Beume, R., et al. (2010). The preclinical pharmacology of roflumilast–a selective, oral phosphodiesterase 4 inhibitor in development for chronic obstructive pulmonary disease. Pulm. Pharmacol. Ther. 23 : 235–256.

He, J., Hu, L., Huang, X., Wang, C., Zhang, Z., Wang, Y., et al. (2020). Potential of coronavirus 3C-like protease inhibitors for the development of new anti-SARS-CoV-2 drugs: Insights from structures of protease and inhibitors. Int. J. Antimicrob. Agents 106055.

Henkin, R.I., Schmidt, L., and Velicu, I. (2013). Interleukin 6 in hyposmia. JAMA Otolaryngol. - Head Neck Surg. 139 : 728–734.

Hertz, A.L., Bender, A.T., Smith, K.C., Gilchrist, M., Amieux, P.S., Aderem, A., et al. (2009). Elevated cyclic AMP and PDE4 inhibition induce chemokine expression in human monocyte-derived macrophages. Proc. Natl. Acad. Sci. 106 : 21978–21983.

Hu, C., Lu, L., Wan, J.-P., and Wen, C. (2017). The Pharmacological Mechanisms and Therapeutic Activities of Hydroxychloroquine in Rheumatic and Related Diseases. Curr. Med. Chem. 24 : 2241–2249.

Hu, F., Jiang, J., and Yin, P. (2020). Prediction of potential commercially inhibitors against SARS-CoV-2 by multi-task deep model. ArXiv Prepr. 2003.00728 :

Huang, C., Wang, Y., Li, X., Ren, L., Zhao, J., Hu, Y., et al. (2020). Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet 395 : 497–506.

Hung, M.J., Cherng, W.J., Hung, M.Y., Wu, H.T., and Pang, J.H.S. (2010). Interleukin-6 inhibits endothelial nitric oxide synthase activation and increases endothelial nitric oxide synthase binding to stabilized caveolin-1 in human vascular endothelial cells. J. Hypertens. 28 : 940–951.

Hyun, Y., and Hong, C. (2017). Deep insight into neutrophil trafficking in various organs. J. Leukoc. Biol. 102: 617–629.

Insel, P.A., Murray, F., Yokoyama, U., Romano, S., Yun, H., Brown, L., et al. (2012). CAMP and Epac in the regulation of tissue fibrosis. Br. J. Pharmacol. 166 : 447–456.

Isoni, C.A., Borges, É.A., Veloso, C.A., Mattos, R.T., Chaves, M.M., and Nogueira-Machado, J.A. (2009). cAMP activates the generation of reactive oxygen species and inhibits the secretion of IL-6 in peripheral blood mononuclear cells from type 2 diabetic patients. Oxid. Med. Cell. Longev. 2 : 317–321.

Jacoby, D.B., Tamaoki, J., Borson, D.B., and Nadel, J.A. (1988). Influenza infection causes airway hyperresponsiveness by decreasing enkephalinase. J. Appl. Physiol. 64 : 2653–2658.

Jia, H. (2016). Pulmonary Angiotensin-Converting Enzyme 2 (ACE2) and Inflammatory Lung Disease. Shock 46: 239-248.

Jin, J.-M., Bai, P., He, W., Wu, F., Liu, X.-F., Han, D.-M., et al. (2020). Gender Differences in Patients With COVID-19: Focus on Severity and Mortality. Front. Public Heal. 8 : 152.

Jones, N.A., Boswell-Smith, V., Lever, R., and Page, C.P. (2005). The effect of selective phosphodiesterase isoenzyme inhibition on neutrophil function in vitro. Pulm. Pharmacol. Ther. 18 : 93–101.

Joyce, E., Fabre, A., and Mahon, N. (2013). Hydroxychloroquine cardiotoxicity presenting as a rapidly evolving biventricular cardiomyopathy: Key diagnostic features and literature review. Eur. Hear. J. Acute Cardiovasc. Care 2 : 77–83.

Kassi, E., and Moutsatsou, P. (2010). Estrogen Receptor Signaling and Its Relationship to Cytokines in Systemic Lupus Erythematosus. J. Biomed. Biotechnol.

Kawamatawong, T. (2017). Roles of roflumilast, a selective phosphodiesterase 4 inhibitor, in airway diseases. J. Thorac. Dis.9:1144.

Kell, D.B., Heyden, E.L., and Pretorius, E. (2020). The Biology of Lactoferrin, an Iron-Binding Protein That Can Help Defend Against Viruses and Bacteria. Front. Immunol. 11 : 1221.

Kimball, A.S., Obi, A.T., Diaz, J.A., and Henke, P.K. (2016). The Emerging Role of NETs in Venous Thrombosis and Immunothrombosis. Front. Immunol. 7 : 236.

Kletsas, D., Caselgrandi, E., Barbieri, D., Stathakos, D., Franceschi, C., and Ottaviani, E. (1998). Neutral endopeptidase-24.11 (NEP) activity in human fibroblasts during development and ageing. Mech. Ageing Dev. 102 : 15–23.

Klok, F.A., Kruip, M.J.H.A., Meer, N.J.M. van der, Arbous, M.S., Gommers, D.A.M.P.J., Kant, K.M., et al. (2020). Incidence of thrombotic complications in critically ill ICU patients with COVID-19. Thromb. Res.

Kowalczyk, A., Kleniewska, P., Kolodziejczyk, M., Skibska, B., and Goraca, A. (2015). The role of endothelin-1 and endothelin receptor antagonists in inflammatory response and sepsis. Arch. Immunol. Ther. Exp. (Warsz). 63 : 41–52.

Kuba, K., Imai, Y., Rao, S., Gao, H., Guo, F., Guan, B., et al. (2005). A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus-induced lung injury. Nat. Med. 11 : 875–879.

Kumar, N., Goldminz, A.M., Kim, N., and Gottlieb, A.B. (2013). Phosphodiesterase 4-targeted treatments for autoimmune diseases. BMC Med. 11 : 96.

Kumar, R.K., Herbert, C., Thomas, P.S., Wollin, L., Beume, R., Yang, M., et al. (2003). Inhibition of inflammation and remodeling by roflumilast and dexamethasone in murine chronic asthma. J. Pharmacol. Exp. Ther. *307* : 349–355.

Lai, C.C., Liu, Y.H., Wang, C.Y., Wang, Y.H., Hsueh, S.C., Yen, M.Y., et al. (2020). Asymptomatic carrier state, acute respiratory disease, and pneumonia due to severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2): Facts and myths. J. Microbiol. Immunol. Infect. 53 : 404.

Leonard-Lorant, I., Delabranche, X., Severac, F., Helms, J., Pauzet, C., Collange, O., et al. (2020). Acute Pulmonary Embolism in COVID-19 Patients on CT Angiography and Relationship to D-Dimer Levels. Radiology 201561. Levi, M., and Poll, T. van der (2017). Coagulation and sepsis. Thromb. Res. 149: 38–44.

Li, C., Booze, R.M., and Hersh, L.B. (1995). Tissue-specific expression of rat neutral endopeptidase (neprilysin) mRNAs. J. Biol. Chem. 270 : 5723–5728.

Li, H., Liu, L., Zhang, D., Xu, J., Dai, H., Tang, N., et al. (2020a). SARS-CoV-2 and viral sepsis: observations and hypotheses. Lancet 395 : 1517–1520.

Li, H., Liu, S.M., Yu, X.H., Tang, S.L., and Tang, C.K. (2020b). Coronavirus disease 2019 (COVID-19): current status and future perspectives. Int. J. Antimicrob. Agents 55 : 105951.

Li, H., Zuo, J., and Tang, W. (2018). Phosphodiesterase-4 inhibitors for the treatment of inflammatory diseases. Front. Pharmacol. 9 :.

Liu, H., Wang, L.L., Zhao, S.J., Kwak-Kim, J., Mor, G., and Liao, A.H. (2020a). Why are pregnant women susceptible to COVID-19? An immunological viewpoint. J. Reprod. Immunol. 139 : 103122.

Liu, T., Zhang, J., Yang, Y., Ma, H., Li, Z., Zhang, J., et al. (2020b). The potential role of IL-6 in monitoring severe case of coronavirus disease 2019. MedRxiv.

Liu, Y., Yang, Y., Zhang, C., Huang, F., Wang, F., Yuan, J., et al. (2020c). Clinical and biochemical indexes from 2019-nCoV infected patients linked to viral loads and lung injury. Sci. China Life Sci. 63 : 364–374.

Luërs, J.C., Klumann, J.P., and Guntinas-Lichius, O. (2020). The COVID-19 pandemic and otolaryngology: What it comes down to? Laryngorhinootologie. 99 : 287–291.

Lyu, J., Miao, T., Dong, J., Cao, R., Li, Y., and Chen, Q. (2020). Reflection on lower rates of COVID-19 in children: Does childhood immunizations offer unexpected protection? Med. Hypotheses 143 : 109842.

Malińska, D., Wieckowski, M.R., Michalska, B., Drabik, K., Prill, M., Patalas-Krawczyk, P., et al. (2019). Mitochondria as a possible target for nicotine action. J. Bioenerg. Biomembr. 51 : 259–276.

Mantzarlis, K., Tsolaki, V., and Zakynthinos, E. (2017). Role of Oxidative Stress and Mitochondrial Dysfunction in Sepsis and Potential Therapies. Oxid. Med. Cell. Longev.

Mao, L., Wang, M., Chen, S., He, Q., Chang, J., Hong, C., et al. (2020). Neurologic Manifestations of Hospitalized Patients With Coronavirus Disease 2019 in Wuhan, China. JAMA Neurol. 77 : 683–690.

Maria, G.U. Di, Bellofiore, S., and Geppetti, P. (1998). Regulation of airway neurogenic inflammation by neutral endopeptidase. Eur. Respir. J.12 : 1454–62.

Marinosci, A., Landis, B.N., and Calmy, A. (2020). Possible link between anosmia and COVID-19: sniffing out the truth. Eur. Arch. Oto-Rhino-Laryngology 277 : 2149–2150.

Martinez, F.J., Calverley, P.M.A., Goehring, U.-M., Brose, M., Fabbri, L.M., and Rabe, K.F. (2015). Effect of roflumilast on exacerbations in patients with severe chronic obstructive pulmonary disease uncontrolled by combination therapy (REACT): a multicentre randomised controlled trial. Lancet *385* : 857–866.

McGonagle, D., O'Donnell, J.S., Sharif, K., Emery, P., and Bridgewood, C. (2020). Immune mechanisms of pulmonary intravascular coagulopathy in COVID-19 pneumonia. Lancet Rheumatol.

Mehra, M.R., Desai, S.S., Ruschitzka, F., and Patel, A.N. (2020). Hydroxychloroquine or chloroquine with or without a macrolide for treatment of COVID-19: a multinational registry analysis. Lancet.

Meyer-Hoffert, U. (2009). Neutrophil-derived serine proteases modulate innate immune responses. Front. Biosci. 14 : 3409–3418.

Milara, J., Morcillo, E., Monleon, D., Tenor, H., and Cortijo, J. (2015a). Roflumilast prevents the metabolic effects of bleomycin-induced fibrosis in a murine model. PLoS One 10 : e0133453.

Milara, J., Morell, A., Ballester, B., Sanz, C., Freire, J., Qian, X., et al. (2015b). Roflumilast improves corticosteroid resistance COPD bronchial epithelial cells stimulated with toll like receptor 3 agonist. Respir. Res. 16 : 12.

Milara, J., Peiro, T., Serrano, A., Artigues, E., Aparicio, J., Tenor, H., et al. (2015c). Simvastatin increases the ability of roflumilast N-oxide to inhibit cigarette smoke-induced epithelial to mesenchymal transition in well-differentiated human bronchial epithelial cells in vitro. COPD J. Chronic Obstr. Pulm. Dis. 12: 327–338.

Molano Franco, D., Arevalo-Rodriguez, I., Roqué I Figuls, M., Montero Oleas, N.G., Nuvials, X., and Zamora, J. (2019). Plasma interleukin-6 concentration for the diagnosis of sepsis in critically ill adults. Cochrane Database Syst. Rev. 4 : CD011811.

Mudter, J., and Neurath, M.F. (2007). Il-6 signaling in inflammatory bowel disease: Pathophysiological role and clinical relevance. Inflamm. Bowel Dis. 13 : 1016–1023.

Muo, I.M., MacDonald, S.D., Madan, R., Park, S.-J., Gharib, A.M., Martinez, P.E., et al. (2019). Early effects of roflumilast on insulin sensitivity in adults with prediabetes and overweight/obesity involve age-associated fat mass loss-results of an exploratory study. Diabetes, Metab. Syndr. Obes. Targets Ther. 12: 743.

Mustafa, S.B., Gandhi, C.R., Harvey, S.A.K., and Olson, M.S. (1995). Endothelin stimulates plateletactivating factor synthesis by cultured rat kupffer cells. Hepatology 21 : 545–553.

Owa, A.B., and Owa, O.T. (2020). Lopinavir/ritonavir use in Covid-19 infection: is it completely non-beneficial? J. Microbiol. Immunol. Infect.

Pani, A., Lauriola, M., Romandini, A., and Scaglione, F. (2020). Macrolides and viral infections: focus on azithromycin in COVID-19 pathology. Int. J. Antimicrob. Agents 106053.

Patel, B.S., Rahman, M.M., Baehring, G., Xenaki, D., Tang, F.S.-M., Oliver, B.G., et al. (2017). Roflumilast N-oxide in combination with formoterol enhances the antiinflammatory effect of dexamethasone in airway smooth muscle cells. Am. J. Respir. Cell Mol. Biol. 56 : 532–538.

Pham, C.T.N. (2006). Neutrophil serine proteases: Specific regulators of inflammation. Nat. Rev. Immunol. 6:541-550.

Press, N.J., and Banner, K.H. (2009). 2 PDE4 Inhibitors - A Review of the Current Field. Prog. Med. Chem. 47: 37–74.

Rabe, K.F. (2011). Update on roflumilast, a phosphodiesterase 4 inhibitor for the treatment of chronic obstructive pulmonary disease. Br. J. Pharmacol. 163 : 53–67.

Rabe, K.F., Watz, H., Baraldo, S., Pedersen, F., Biondini, D., Bagul, N., et al. (2018). Anti-inflammatory effects of roflumilast in chronic obstructive pulmonary disease (ROBERT): a 16-week, randomised, placebo-controlled trial. Lancet Respir. Med. 6 : 827–836.

Raker, V.K., Becker, C., and Steinbrink, K. (2016). The cAMP pathway as the rapeutic target in autoimmune and inflammatory diseases. Front. Immunol. 7: 123.

Rao, G.H.R., and White, J.G. (1982). Platelet activating factor (paf) causes human platelet aggregation through the mechanism of membrane modulation. Prostaglandins, Leukot. Med. 9:459-472.

Rao, V.U., Pavlov, A., Klearman, M., Musselman, D., Giles, J.T., Bathon, J.M., et al. (2015). An evaluation of risk factors for major adverse cardiovascular events during tocilizumab therapy. Arthritis Rheumatol. 67 : 372–380.

Rasmo, D. De, Micelli, L., Santeramo, A., Signorile, A., Lattanzio, P., and Papa, S. (2016). CAMP regulates the functional activity, coupling efficiency and structural organization of mammalian FOF1 ATP synthase.

Biochim. Biophys. Acta - Bioenerg. 1857 : 350–358.

Reddy, A.T., Lakshmi, S.P., Banno, A., and Reddy, R.C. (2020). Glucocorticoid Receptor α Mediates Roflumilast's Ability to Restore Dexamethasone Sensitivity in COPD. Int. J. Chron. Obstruct. Pulmon. Dis. 15 : 125–134.

Redford, P.S., Murray, P.J., and O'garra, A. (2011). The role of IL-10 in immune regulation during M. tuberculosis infection. Mucosal Immunol.4 : 261–270.

Remy, K.E., Brakenridge, S.C., Francois, B., Daix, T., Deutschman, C.S., Monneret, G., et al. (2020). Immunotherapies for COVID-19: lessons learned from sepsis. Lancet. Respir. Med.

Rhee, C.K., and Kim, D.K. (2020). Role of phosphodiesterase-4 inhibitors in chronic obstructive pulmonary disease. Korean J. Intern. Med. 35 : 276–283.

Rosenberg, E.S., Dufort, E.M., Udo, T., Wilberschied, L.A., Kumar, J., Tesoriero, J., et al. (2020). Association of Treatment with Hydroxychloroquine or Azithromycin with In-Hospital Mortality in Patients with COVID-19 in New York State. JAMA - J. Am. Med. Assoc.

Sanz, M.J., Cortijo, J., Taha, M.A., Cerdá-Nicolás, M., Schatton, E., Burgbacher, B., et al. (2007). Roflumilast inhibits leukocyte-endothelial cell interactions, expression of adhesion molecules and microvascular permeability. Br. J. Pharmacol. 152 : 481–492.

Sato, T., Hongu, T., Sakamoto, M., Funakoshi, Y., and Kanaho, Y. (2013). Molecular Mechanisms of N-Formyl-Methionyl-Leucyl-Phenylalanine-Induced Superoxide Generation and Degranulation in Mouse Neutrophils: Phospholipase D Is Dispensable. Mol. Cell. Biol. 33 : 136–145.

Schalkwyk, E. van, Strydom, K., Williams, Z., Venter, L., Leichtl, S., Schmid-Wirlitsch, C., et al. (2005). Roflumilast, an oral, once-daily phosphodiesterase 4 inhibitor, attenuates allergen-induced asthmatic reactions. J. Allergy Clin. Immunol. *116* : 292–298.

Schouten, M., Wiersinga, W.J., Levi, M., and Poll, T. van der (2008). Inflammation, endothelium, and coagulation in sepsis. J. Leukoc. Biol. 83 : 536–545.

Shames, B.D., McIntyre, R.C., Bensard, D.D., Pulido, E.J., Selzman, C.H., Reznikov, L.L., et al. (2001). Suppression of tumor necrosis factor α production by cAMP in human monocytes: Dissociation with mRNA level and independent of interleukin-10. J. Surg. Res. 99: 187–193.

Shen, L.-F., Lv, X.-D., Chen, W.-Y., Yang, Q., Fang, Z.-X., and Lu, W.-F. (2018). Effect of roflumilast on chronic obstructive pulmonary disease: a systematic review and meta-analysis. Ir. J. Med. Sci. 187 : 731–738.

Shimamoto, K., Ura, N., and Iimura, O. (1994). The natriuretic mechanisms of neutral endopeptidase inhibitor in rats. Brazilian J. Med. Biol. Res. Rev. Bras. Pesqui. Medicas e Biol. 27 : 1965–1973.

Silva, S., Amarasena, R., Moorcroft, J., Rajakulenthiran, T., and Singh, R. (2020). Tocilizumab-induced pulmonary fibrosis in a patient with rheumatoid arthritis. Clin. Med. (Northfield. Il). 20 : s57–s57.

Singhal, T. (2020). A Review of Coronavirus Disease-2019 (COVID-19). Indian J. Pediatr. 87: 281–286.

Song, Y., Liu, P., Shi, X.L., Chu, Y.L., Zhang, J., Xia, J., et al. (2020). SARS-CoV-2 induced diarrhoea as onset symptom in patient with COVID-19. Gut 69 : 1143–1144.

Sont, J.K., Krieken, J.H.J.M. Van, Klink, H.C.J. Van, Roldaan, A.C., Apap, C.R., Willems, L.N.A., et al. (1997). Enhanced Expression of Neutral Endopeptidase (NEP) in Airway Epithelium in Biopsies from Steroid- versus Nonsteroid-treated Patients with Atopic Asthma. Am. J. Respir. Cell Mol. Biol. 16: 549–556.

Sprague, A.H., and Khalil, R.A. (2009). Inflammatory cytokines in vascular dysfunction and vascular disease. Biochem. Pharmacol. 78 : 539–552.

Srivastava, S. (2017). The mitochondrial basis of aging and age-related disorders. Genes (Basel). 8 : 398.

Tabaa, M.M. El, and Tabaa, M.M. El (2020). Targeting Neprilysin (NEP) pathways: A potential new hope to defeat COVID-19 ghost. Biochem. Pharmacol. 114057.

Taghizadeh-Hesary, F., and Akbari, H. (2020). The powerful immune system against powerful COVID-19: A hypothesis. Med. Hypotheses 140 : 109762.

Tamara, A., and Tahapary, D.L. (2020). Obesity as a predictor for a poor prognosis of COVID-19: A systematic review. Diabetes Metab. Syndr. Clin. Res. Rev. 14 : 655–659.

Tay, M.Z., Poh, C.M., Rénia, L., MacAry, P.A., and Ng, L.F.P. (2020). The trinity of COVID-19: immunity, inflammation and intervention. Nat. Rev. Immunol. 20 : 363–374.

Tenor, H., Hatzelmann, A., Church, M.K., Schudt, C., and Shute, J.K. (1996). Effects of theophylline and rolipram on leukotriene C4 (LTC4) synthesis and chemotaxis of human eosinophils from normal and atopic subjects. Br. J. Pharmacol. 118 : 1727–1735.

Togo, S., Liu, X., Wang, X., Sugiura, H., Kamio, K., Kawasaki, S., et al. (2009). PDE4 inhibitors roflumilast and rolipram augment PGE2 inhibition of TGF-β1-stimulated fibroblasts. Am. J. Physiol. - Lung Cell. Mol. Physiol. 296 : L959-69.

Totani, L., Amore, C., Santo, A. Di, Dell'Elba, G., Piccoli, A., Martelli, N., et al. (2016). Roflumilast inhibits leukocyte–platelet interactions and prevents the prothrombotic functions of polymorphonuclear leukocytes and monocytes. J. Thromb. Haemost. 14 : 191–204.

Varga, Z., Flammer, A.J., Steiger, P., Haberecker, M., Andermatt, R., Zinkernagel, A.S., et al. (2020). Endothelial cell infection and endotheliitis in COVID-19. Lancet 395 : 1417–1418.

Vishnevetsky, A., and Levy, M. (2020). Rethinking high-risk groups in COVID-19. Mult. Scler. Relat. Disord. 42 : 102139.

Vollert, S., Kaessner, N., Heuser, A., Hanauer, G., Dieckmann, A., Knaack, D., et al. (2012). The glucoselowering effects of the PDE4 inhibitors roflumilast and roflumilast-N-oxide in db/db mice. Diabetologia 55 : 2779–2788.

Wang, M., Gao, P., Wu, X., Chen, Y., Feng, Y., Yang, Q., et al. (2016). Impaired anti-inflammatory action of glucocorticoid in neutrophil from patients with steroid-resistant asthma. Respir. Res. 17 : 1–9.

Weiss, B., and Hait, W.N. (1977). Selective cyclic nucleotide phosphodiesterase inhibitors as potential therapeutic agents. Annu. Rev. Pharmacol. Toxicol. 17 : 441–477.

Wermuth, P.J., Li, Z., Mendoza, F.A., and Jimenez, S.A. (2016). Stimulation of Transforming Growth Factor- β 1-Induced Endothelial-To-Mesenchymal Transition and Tissue Fibrosis by Endothelin-1 (ET-1): A Novel Profibrotic Effect of ET-1. PLoS One 11 : e0161988.

Westphal, J.F. (2000). Macrolide - Induced clinically relevant drug interactions with cytochrome P-450A (CYP) 3A4: An update focused on clarithromycin, azithromycin and dirithromycin. Br. J. Clin. Pharma-col.50 : 285–295.

Wick, M.J., Buesing, E.J., Wehling, C.A., Loomis, Z.L., Cool, C.D., Zamora, M.R., et al. (2011). Decreased neprilysin and pulmonary vascular remodeling in chronic obstructive pulmonary disease. Am. J. Respir. Crit. Care Med. 183 : 330–340.

Wintroub, B.U., Schechter, N.B., Lazarus, G.S., Kaempfer, C.E., and Schwartz, L.B. (1984). Angiotensin I conversion by human and rat chymotryptic proteinases. J. Invest. Dermatol. *83* : 336–339.

Wouters, E.F.M., Bredenbröker, D., Teichmann, P., Brose, M., Rabe, K.F., Fabbri, L.M., et al. (2012). Effect of the phosphodiesterase 4 inhibitor roflumilast on glucose metabolism in patients with treatmentnaive, newly diagnosed type 2 diabetes mellitus. J. Clin. Endocrinol. Metab. 97 : E1720–E1725. Wu, P., Duan, F., Luo, C., Liu, Q., Qu, X., Liang, L., et al. (2020). Characteristics of Ocular Findings of Patients with Coronavirus Disease 2019 (COVID-19) in Hubei Province, China. JAMA Ophthalmol. 138 : 575–578.

Xiao, Z.M., Sun, L., Liu, Y.M., Zhang, J.J., and Huang, J. (2009). Estrogen regulation of the neprilysin gene through a hormone-responsive element. J. Mol. Neurosci. 39 : 22–26.

Xiong, Y., Liu, Y., Cao, L., Wang, D., Guo, M., Jiang, A., et al. (2020). Transcriptomic characteristics of bronchoalveolar lavage fluid and peripheral blood mononuclear cells in COVID-19 patients. Emerg. Microbes Infect. 9 : 761–770.

Yang, X., Yu, Y., Xu, J., Shu, H., Xia, J., Liu, H., et al. (2020). Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: a single-centered, retrospective, observational study. Lancet Respir. Med. 8 : 475–481.

Yao, M., Nguyen, T.V. V., Rosario, E.R., Ramsden, M., and Pike, C.J. (2008). And rogens regulate neprilysin expression: Role in reducing β -amyloid levels. J. Neurochem. 105 : 2477–2488.

Zhang, H., Penninger, J.M., Li, Y., Zhong, N., and Slutsky, A.S. (2020a). Angiotensin-converting enzyme 2 (ACE2) as a SARS-CoV-2 receptor: molecular mechanisms and potential therapeutic target. Intensive Care Med. 46 : 586–590.

Zhang, Y., Geng, X., Tan, Y., Li, Q., Xu, C., Xu, J., et al. (2020b). New understanding of the damage of SARS-CoV-2 infection outside the respiratory system. Biomed. Pharmacother. 127 : 110195.

Zhou, F., Yu, T., Du, R., Fan, G., Liu, Y., Liu, Z., et al. (2020). Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. Lancet 395 : 1054–1062.

Ziebuhr, J. (2005). The coronavirus replicase. Curr. Top. Microbiol. Immunol. 287: 57–94.

Zuo, Y., Zuo, M., Yalavarthi, S., Gockman, K., Madison, J.A., Shi, H., et al. Neutrophil extracellular traps and thrombosis in COVID-19. MedRxiv.

Zyl-Smit, R.N. van, Richards, G., and Leone, F.T. (2020). Tobacco smoking and COVID-19 infection. Lancet Respir. Med. 1451–1454.

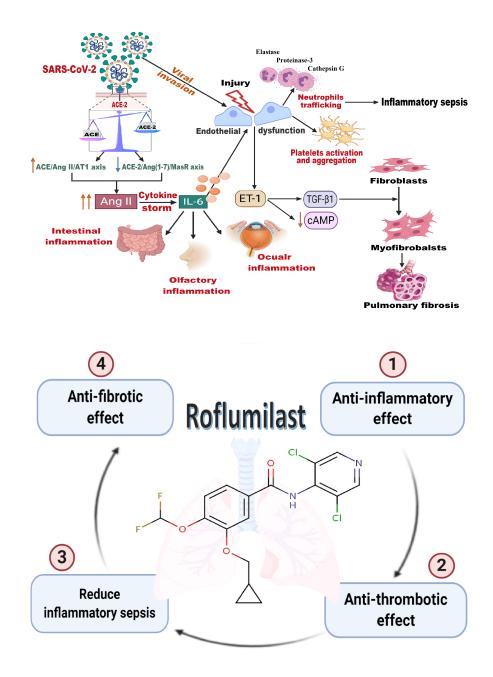
 Table 1: Multiple pharmacological properties of roflumilast.

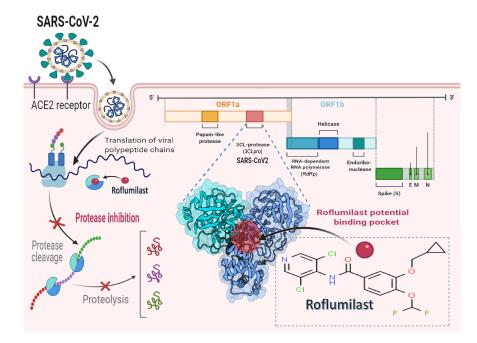
Dose	Effect	Main molecular mechanisms of action	Model	References
10 ⁻⁹ – 10 ⁻⁶ M	Inhibition of neutrophils function	Suppressed the release of MPO, NE and MMP-9	Neutrophils adhesion to HUVECs	(Jones et al., 2005)
1–1000 nM L ⁻¹	Inhibition of the prothrombotic functions of PMNs and MNs	Inhibited the release of NETs and suppressed tissue factor expression in MNs	Human PLTs and PMNs	(Totani et al., 2016)

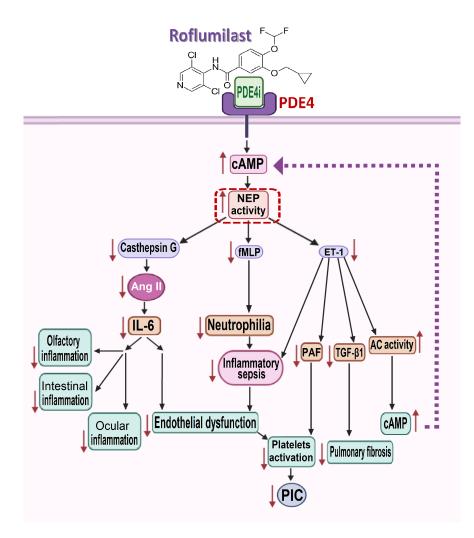
		Main molecular mechanisms of		
Dose	Effect	action	Model	References
500 ug/d	Anti- inflammatory effects	Inhibited phosphodiesterase- 4 enzyme that targets the systemic inflammation associated with COPD and decreased inflammatory mediators	COPD patients	(Martinez et al., 2015)
500 ug/d	Anti- inflammatory effects	Inhibited allergen-induced sputum eosinophils, neutrophils and ECP	Allergic asthmatic patients	(Gauvreau et al., 2011; Bateman et al., 2016)
0.3 – 1.0 mg/kg body	Prevention of polymicrobial sepsis	Reduced βαςτεριαλ λοαδ, ινηιβιτεδ εξπρεσσιον οφ προ- ινφλαμματορψ ςψτοχινες μαινλψ ΙΛ-6 ανδ ΤΝΦ-αλπηα ανδ συππρεσσεδ ΝΦ-χΒ, π38 ΜΑΠΚ ανδ ΣΤΑΤ3	Mice with cecal ligation and puncture-induced sepsis	(Feng et al., 2017)
1, 10, $\alpha\nu\delta$ 100 ν $\mu o\lambda/\Lambda$ $\alpha\nu\delta$ 1 μ $\mu o\lambda/\Lambda$ dissolved in DMSO	Inhibition of airway remodeling	Inhibited ECM protein deposition and thereby, airway remodeling	Human ASM cells	(Burgess et al., 2006)
5 mg/kg/d, suspended in 2.5% polyethylene glycol 4% methylcellulose solution	Inhibition of airway remodeling	Reduced the accumulation of chronic inflammatory cells, and thickening of airway epithelium	BALB/c mice model of chronic asthma	(Kumar et al., 2003)
10 ⁻⁹ – 10 ⁻⁶ M	Anti- proliferative effects	Attenuated cell proliferation and production of (MMP-2 and MMP-9)	Distal human PASMCs	(Growcott et al., 2006)

Dose	Effect	Main molecular mechanisms of action	Model	References
5 mg/kg/day	Anti- fibrotic effects	Antagonized metabolic effects related to pulmonary fibrosis (like alterations in the oxidative equilibrium, a strong inflammatory response and collagen synthesis activation)	Bleomycin- Induced Fibrosis in mice	(Milara et al., 2015a)
10 ⁻⁶ – 10 ⁻⁷ M	Anti-fibrotic effects	Antagonized the profibrotic activity of fibroblasts stimulated by TGF-beta1	Adult human lung fibroblast cell lines	(Togo et al., 2009)
500 ug/d	Anti- hyperglycemic effects	Enhanced secretion of intestinal GLP-1, a main incretin with potent insulinotropic effect	35–70 yr. patients with newly diagnosed DM type II	(Wouters et al., 2012)

ASM; Airway smooth muscle, COPD; Chronic obstructive pulmonary disease, CS; Cigarette smoke, DM; Diabetes mellitus, DMSO; Dimethyl sulfoxide, ECM; Extracellular matrix, ECP; Eosinophil cationic protein, GLP-1;glucagon like peptide-1, HUVECs; Human umbilical vein endothelial cells, MAPK; Mitogen-activated protein kinase, MMP-9; Matrix metalloproteinase-9, MN;Monocytes, MPO; Myeloperoxidase, NE; Neutrophil elastase, NETs; Neutrophil extracellular traps, NΦ-×B·Nuclear factor-kappa B, OVA; Ovalbumin, PASMCs;Pulmonary artery smooth muscle cells, PMNs;Polymorphonuclear leukocytes, PLTs; Platelets, STAT3;Signal transducer and activator of transcription 3







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Table.docx available at https://authorea.com/users/339038/articles/465335-new-putativeinsights-into-neprilysin-nep-dependent-pharmacotherapeutic-role-of-roflumilast-intreating-covid-19