

Phylogenetic significance of the characteristics of simple sequence repeats at the genus level based on the complete chloroplast genome sequences of Cyatheaceae

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Abstract

Recent taxonomic and molecular phylogenetic studies have shown that *Gymnosphaera* should be recognized as an independent taxonomic unit at the genus level under the family Cyatheaceae. In this study, the complete chloroplast genomes of the eight species of Cyatheaceae were sequenced, and their phylogenetic relationships were reconstructed using the maximum likelihood, Bayesian inference, maximum parsimony, and neighbor-joining methods, and the characteristics of their simple sequence repeats (SSRs) were compared and analyzed for the first time. The results showed that when Cyatheaceae was divided into three genera, the number, relative abundance, relative density, and GC content of all SSRs and of SSRs of certain unit lengths in the chloroplast genomes of the eight species of Cyatheaceae were genus specific in the whole chloroplast genomes and in their different regions (large single-copy, small single-copy, inverted repeat, intergenic spacer, intron, rRNA gene, and coding sequence regions). The SSRs overall and the single-nucleotide SSRs had significant differences in number, relative abundance, relative density, and GC content between the chloroplast genomes, their intergenic regions, and large single-copy regions. When Cyatheaceae was divided into two genera, only the difference in GC content was significant. Therefore, our results support the restoration of the hierarchical status of *Gymnosphaera*. This study provides an important basis for the identification of the phylogenetic relationship of Cyatheaceae plants.

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Abstract Recent taxonomic and molecular phylogenetic studies have shown that *Gymnosphaera* should be recognized as an independent taxonomic unit at the genus level under the family Cyatheaceae. In this

study, the complete chloroplast genomes of the eight species of Cyatheaceae were sequenced, and their phylogenetic relationships were reconstructed using the maximum likelihood, Bayesian inference, maximum parsimony, and neighbor-joining methods, and the characteristics of their simple sequence repeats (SSRs) were compared and analyzed for the first time. The results showed that when Cyatheaceae was divided into three genera, the number, relative abundance, relative density, and GC content of all SSRs and of SSRs of certain unit lengths in the chloroplast genomes of the eight species of Cyatheaceae were genus specific in the whole chloroplast genomes and in their different regions (large single-copy, small single-copy, inverted repeat, intergenic spacer, intron, rRNA gene, and coding sequence regions). The SSRs overall and the single-nucleotide SSRs had significant differences in number, relative abundance, relative density, and GC content between the chloroplast genomes, their intergenic regions, and large single-copy regions. When Cyatheaceae was divided into two genera, only the difference in GC content was significant. Therefore, our results support the restoration of the hierarchical status of *Gymnosphaera*. This study provides an important basis for the identification of the phylogenetic relationship of Cyatheaceae plants.

Keywords Cyatheaceae, Phylogeny, SSR, *Gymnosphaera*

Also known as microsatellites, simple sequence repeats (SSRs) are short tandem repeat sequences with a motif length of 1-6 bp that are characterized by high variability, high repeatability, codominant inheritance, and interspecies generality. SSRs have been widely used in species identification, genetic diversity, and phylogenetic relationships(Chmielewski et al., 2015; Dashnow et al., 2015). SSRs are caused by slipped strand mispairing and subsequent errors during DNA replication, repair, and recombination (Levinson & Gutman, 1987). They are widespread in the genomes of prokaryotes, eukaryotes, and some viruses(Mrazek et al., 2007; Toth et al., 2000; Zhao et al., 2011). SSRs are mainly found in intergenic and noncoding regions, with a few in introns (Toth et al., 2000; Li et al., 2004). Under selection, the number of genomic SSRs is significantly higher than that under random accumulation(Ellegren, 2004). The characteristics of genomic SSRs in different taxa (such as their distribution pattern) accurately reflect their phylogenetic relationships (Srivastava et al., 2019). SSRs are highly mutagenic and are an important source of genetic evolution, and they can play an important role in genome evolution by generating and maintaining quantitative genetic variation (Chaoui et al., 2012; Gemayel et al., 2010; Kashi & King, 2006). Therefore, the identification and characterization of SSRs can contribute to the study of population variation, germline identification, and genetic diversity.

Cyatheaceae belong to the phylum Pteridophyta, the class Polypodiopsida, and the order Cyatheales, with more than 600 species present today. Cyatheaceae account for the vast majority of known tree ferns and are mainly distributed in warm and humid tropical and subtropical regions (Kramer, 1990; Korall et al., 2006; Smith et al., 2006; PPG I, 2016). The classification of the genera in Cyatheaceae has always been a difficult problem. In the early classification system, Christensen (1906) classified Cyatheaceae into *Cyathea* Sm. (with a complete cup-shaped indusium), *Hemitelia* R. Br. (with a partial indusium), and *Alsophila* R. Br. (without an indusium) based on the characteristics of indusium. Holttum (1963) treated Cyatheaceae plants as a genus (*Cyathea*) and divided it into two subgenera, *Cyathea* subg. *Cyathea* (marginate scales) and *Cyathea* subg. *Sphaeropteris* (Bernh.) Holttum (conform scales), based on the structural characteristics of the scales at the base of the petiole. Based on the study of the American Cyatheaceae plants, Tryon (1970) delineated three evolutionary paths and six genera, namely, (1) *Sphaeropteris* Bernh (with conform scales); (2)*Alsophila* and *Nephelea* R. M. Tryon (with marginate scales and apical setae); and (3) *Trichipteris* C. Presl, *Cyathea* and *Cnemidaria* C. Presl (with marginate scales but without apical setae). Ching (1978) classified the Cyatheaceae plants that grow in China into three genera: *Sphaeropteris*, *Gymnosphaera* and *Alsophila*. On this basis, Xia (1989) reduced the genera *Alsophila* and *Alsophila* to subgenera, respectively, and combined them into *Alsophila*.

Recent molecular phylogenetic studies have shown that Cyatheaceae includes four monophyletic groups, namely, *Alsophila*, *Cyathea*, *Gymnosphaera*, and *Sphaeropteris* (Dong & Zuo, 2018; Janssen & Rakotozondrainibe, 2008; Korall et al., 2007; Korall & Pryer, 2014). The morphological characteristics of *Gymnosphaera*, such as marginate scales, a slightly black rachis, and the lack of an indusium, and molecu-

lar evidence suggest that it is different from other Cyatheaceae plants, and particularly obvious is its different sporogenesis from *Alsophila* (*Gymnosphaera* produces 64 spores per sac, *Alsophila* produces 16 spores per sac) (Dong & Zuo, 2018). Still, some scholars classify *Gymnosphaera* into the genus *Alsophila* (Xia 1989; Zhang and Nishida, 2013; PPG I, 2016).

In this study, the chloroplast genomes of eight species of Cyatheaceae were sequenced, and the chloroplast genome, the SSRs of different unit lengths in the chloroplast genome, and the number, relative abundance, relative density, and GC content of all SSRs and SSRs of specific unit lengths in the intergenic spacer regions (IGS), intron regions, and protein-coding sequence regions (CDS) of the chloroplast genome were compared and analyzed to reveal the phylogenetic significance of the SSR characteristics of the chloroplast genome of Cyatheaceae plants.

1 Materials and methods

1.1 Sampling

The leaves of *Alsophila denticulata* Baker and *Alsophila metteniana* Hance were collected from Nankunshan in Huizhou and the botanical garden of South China Agricultural University in Guangzhou, respectively. The leaves of *Alsophila podophylla* Hook and *Alsophila gigantean* Wall. ex Hook were collected from the South China Botanical Garden of the Chinese Academy of Sciences in Guangzhou (Liu et al., 2018; Wang et al., 2019b). The leaves of *Alsophila costularis* Baker, *Sphaeropteris brunonianus* (Hook.) R. M. Tryon, and *Sphaeropteris lepifera* (Hook.) R. M. Tryon were collected from the Fairy Lake Botanical Garden of the Chinese Academy of Sciences in Shenzhen (Wang et al., 2019a; Liu et al., 2020; Zhu et al., 2020). The leaves of *Alsophila spinulosa* (Wall. ex Hook.) R. M. Tryon were collected from the Wuhan Botanical Garden of the Chinese Academy of Sciences in Wuhan (Gao et al., 2009). Fresh young leaves from well-grown plants were collected, wrapped in tin paper, flash-frozen in liquid nitrogen, and then stored at -80 °C before use.

1.2 DNA extraction and sequencing

The plant genomic DNA extraction kit (TIANGEN) was used to extract the total DNA of the samples. After the quality of the total DNA samples was confirmed by Shanghai Hanyu Biotechnology Co., Ltd., the samples were subjected to bidirectional sequencing using an Illumina HiSeq 2500, and the raw data obtained were converted into raw reads by CASAVA base-calling analysis. The clean data obtained after removing the adaptor-containing, low-quality sequences were taken for subsequent analysis. Data processing was performed by Trimmomatic v0.32 (Bolger et al., 2014) with the following steps: (1) removal of sequences containing N bases; (2) removal of adaptor sequences in the reads; (3) removal of low-quality bases (Q value < 20) from the reads in the 3' to 5' direction; (4) removal of low-quality bases (Q value < 20) from the reads in the 5' to 3' direction; (5) removal of four bases with an average base quality of less than 20; and (6) removal of the reads and their pairs with a length less than 50 nt. Velvet v1.2.03 (Zerbino & Birney, 2008) was used to assemble the clean data.

1.3 Characterization of chloroplast genome

The chloroplast genome of *Alsophila* was used as the reference genome, and Dual Organellar GenoMe Annotator (DOGMA) (Milne et al., 2010) was used to predict the protein-coding genes, rRNA genes, and tRNA genes in other genomes. Geneious Prime (Kearse et al., 2012) was used for manual correction according to the reference genome. The Shuffle-Lagan mode in the online software mVISTA (Frazer et al., 2004) was used for genome-wide comparison. Organellar Genome DRAW (OGDRAW) (Lohse et al., 2007) was used to draw the physical chloroplast genome map, and Sequin software was used for the submission of the chloroplast genome of *Alsophila denticulata*. Microsatellite repeats were predicted using the software MISA (Beier et al., 2017). The threshold repeat number of single-nucleotide units was set to 10, the threshold repeat number of dinucleotide units was set to six, and the threshold repeat number of trinucleotides, tetranucleotides, pentanucleotides, and hexanucleotides was set to three. The minimum distance between two SSRs was set to 0 bp, that is, there was no statistical compound SSR. The distribution characteristics of SSRs of different species in the whole genome and its different regions were compared and analyzed. Among these character-

istics, the relative abundance refers to the number of SSRs in the unit sequence length (kb), and the relative density refers to the length of the SSR (bp) in the unit sequence length (kb).

1.4 Phylogenetic analysis

The maximum likelihood (ML), Bayesian inference (BI), maximum parsimony (MP) and neighbor-joining (NJ) methods were used for phylogenetic analysis. MAFFT software (Katoh & Standley, 2013) was used to align the complete chloroplast genome sequences of eight species of Cyatheaceae and one species of *Cibotium*, *Cibotium barometz* (Linn.) J. Sm. A phylogenetic tree was constructed using *Cibotium barometz* (Linn.) J. Sm. as an outgroup. When the ML, MP, and BI trees had been constructed, the whole chloroplast genome was screened in MrModelTest software to obtain the optimal nucleotide substitution model (GTR+I+G) selected based on the Akaike information criterion, and the relevant parameters were estimated. The ML tree was constructed by the software RAxML8.0.20 (Stamatakis, 2014), GTRGAMMA1 was selected as the nucleotide substitution model, and the confidence of the branch was completed using the bootstrap analysis in autoMR. The BI tree was constructed by MrBayes v3.2.0 software (Ronquist et al., 2012) and was estimated by running 2,000,000 generations (Nst = 6, rates = invgamma). The MP tree was constructed in PAUP 4.0 software (Swofford, 2002) with the bootstrap value set to 1000. The NJ tree was constructed in MEGA7.0 software (Kumar et al., 2016), and the maximum composite likelihood algorithm was selected with the bootstrap value set to 1000 times. The resulting phylogenetic tree was viewed and edited in Figtree v 1.4.3 software.

1.5 Statistical analysis

When *Gymnosphaera* is considered as an independent taxonomic unit at the genus level, the eight species of Cyatheaceae are divided into three genera. That is, *Alsophila denticulata*, *Alsophilapodophylla*, *Alsophila metteniana*, and *Alsophila gigantea* belong to the genus *Gymnosphaera*; *Alsophila spinulosa* and *Alsophila costularis* belong to the genus *Alsophila*; and *Sphaeropteris brunonianana* and *Sphaeropteris lepifera* belong to the genus *Sphaeropteris*. When *Gymnosphaera* is classified into the genus *Alsophila*, Cyatheaceae is divided into two genera. The Kruskal-Wallis H test and Mann-Whitney U test in IBM SPSS v22.0 software (Allen et al., 2014) were used to analyze the significance of the differences between taxa when three genera and two genera were assumed, respectively. The statistical results covered the whole chloroplast genome, the SSRs of different unit lengths in the chloroplast genome, and the number, relative abundance, relative density, and GC content of SSRs and SSRs of different unit lengths in the IGS, LSC, intronic, and CDS regions of the chloroplast genome of the eight species of Cyatheaceae. Photovoltaic (PV) cluster analysis using the ward linkage method in R v3.5.1 (R Core Team, 2013) was performed on the SSRs of the chloroplast genome and its IGS regions, LSC regions and on the number, relative abundance, relative density, and GC content of single-nucleotide SSRs of the chloroplast genome with Euclidean distance as the measurement. The number of repetitions is 10,000.

2 Results and analysis

2.1 Genome structures and characteristics

The chloroplast genomes of all eight species of Cyatheaceae are double-stranded, closed, circular molecules with a typical tetrad structure (with *Alsophila denticulata* as an example, as shown in Figure 1). The genome length ranges from 154,046 bp (*Alsophila denticulata*) to 166,151 bp (*Alsophila gigantea*). The structure includes a large single-copy region (LSC) (bp 85975–92315), a small single-copy region (SSC) (bp 23245–28874), and an inverted repeat region (IR) (bp 23245–28874), of which IRA and IRB are two inverted repeat regions. The GC content of each region of the chloroplast genome of different species varied little, the total GC content ranging from 40.3% to 41.9% (Table 1). Only LSC, SSC, and one IR were analyzed. The chloroplast genome of each species of Cyatheaceae contained 117 genes, which encoded 85 proteins, four rRNAs, and 28 tRNAs. Pseudogenes (*ycf66*, *trnT-UGU*) were also present in these genomes. Among these genes are 13 genes located in the IR region. The *ndhB* gene spans the LSC and IRA regions, and there is a duplicated exon 2 sequence of the *ndhB* gene near the boundary of the IRB. Twelve genes have one intron, and three genes (*ycf3*, *clpP*, and *rps12*) have two introns.

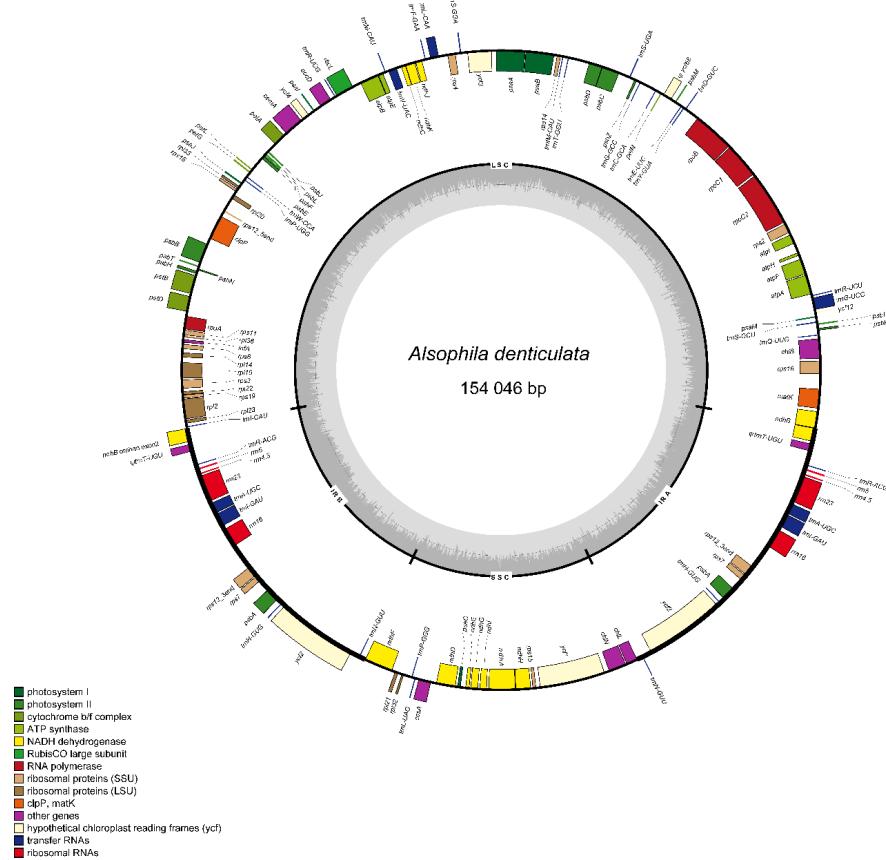


Figure 1 Gene map of the chloroplast genome of *Alsophila denticulata*. Genes located in outside of the outer circle are transcribed in the counterclockwise direction, whereas those inside are transcribed in the clockwise direction. Color codes represent different functional gene groups. In the middle circle, the GC and AT content variations are indicated by darker gray and lighter gray, respectively.

Table 1 List of eight Cyatheaceae species, GenBank accession numbers, and structural features of chloroplast genomes

Species	GenBank No.	LSC	LSC	LSC	IR	IR	IR	SSC	SSC
		Length (bp)	GC%	Length (%)	Length (bp)	GC%	Length (%)	Length (bp)	GC%
<i>A. denticulata</i>	MN795320	85975	40.0	55.8	23245	40.8	15.1	21581	33.3
<i>A. podophylla</i>	MG282389	86762	40.0	52.2	28874	46.2	17.4	21641	33.3
<i>A. gigantea</i>	MH603068	92315	41.1	57.1	23831	43.3	14.7	21702	33.3
<i>A. metteniana</i>	MT726940	92292	41.1	57.1	23822	43.3	14.7	21666	33.3
<i>A. costularis</i>	MH684489	86338	39.6	55.1	24356	43.0	15.5	21625	33.3
<i>A. spinulosa</i>	NC012818	86308	39.6	55.1	24365	43.0	15.6	21623	33.3
<i>S. brunonianana</i>	MT543220	86196	39.2	55.0	24011	43.2	15.3	22441	33.3
<i>S. lepifera</i>	MN623357	86349	39.3	53.3	24067	43.2	14.8	27733	40.0

LSC: Large Single Copy-Region; IR: Inverted Repeat Region; SSC: Small Single Copy-Region

2.2 Analysis of the characteristics of SSRs

The number, relative abundance, relative density, and GC content of SSRs in the chloroplast genomes of all eight species of Cyatheaceae were systematically compared (Table 2). The number (121–122), relative abundance (0.77–0.78/bp), relative density (9.81–9.82 bp/kb), and GC content (0.18–0.20) of SSRs in the chloroplast genomes of *Alsophila spinulosa* and *Alsophila costularis*; the number (102), relative abundance (0.63–0.65/bp), relative density (6.70–8.18 bp/kb), and GC content (0.08–0.10) of SSRs in the chloroplast genomes of *Sphaeropteris brunoniana* and *Sphaeropteris lepifera*; and the number (61–67), relative abundance (0.40/bp), relative density (4.11–5.06 bp/kb), and GC content (0.22–0.29) of SSRs of *Alsophila denticulata*, *Alsophila podophylla*, *Alsophila metteniana*, and *Alsophila gigantea* had similar values, which were not proportional to the size of the genome. The proportions of GC bases in the chloroplast genomes of the eight species of Cyatheaceae were much lower than the proportions of AT. SSRs accounted for 75.5–86.2%, 13.7–20.6%, 2.0–3.9%, and 2.0% of the IGS, Intron, CDS, and rRNA gene regions (pseudogenes were treated as IGS). Among them, SSR was only detected in the CDS of the chloroplast genomes of *Sphaeropteris brunoniana*, *Sphaeropteris lepifera*, *Alsophila spinulosa*, and *Alsophila costularis*, and SSRs were detected in the rRNA genes of the chloroplast genomes of *Sphaeropteris brunoniana* and *Sphaeropteris lepifera*. When *Gymnosphaera* was considered as an independent classification unit at the genus level, the eight species of Cyatheaceae were divided into three genera. That is, *Alsophila denticulata*, *Alsophila podophylla*, *Alsophila metteniana*, and *Alsophila gigantea* belonged to the genus *Gymnosphaera*; *Alsophila spinulosa* and *Alsophila costularis* belonged to the genus *Alsophila*; and *Sphaeropteris brunoniana* and *Sphaeropteris lepifera* belonged to the genus *Sphaeropteris*. The number, relative abundance, relative density, and GC content of SSRs in the chloroplast genomes of the three genera of Cyatheaceae also had similar values in different regions of the genome (LSC, SSC, and IR; IGS, intron, CDS, and rRNA gene regions), indicating that in the phylogenetic background of the three genera, the characteristics of SSRs are genus specific at the level of the genome and its different regions (Figure 3, Tables 2 and 3, and Supplementary Tables 9 and 10). The distribution of SSRs of different motif types in the three Cyatheaceae genera also had genus specificity (Figure 2). These results showed that on the phylogenetic background of dividing the eight species of Cyatheaceae into three genera, different taxa had different patterns of SSR characteristics in the chloroplast genome, namely, the SSR characteristics of the chloroplast genomes of the eight species of Cyatheaceae were consistent with their phylogenetic relationship.

Table 2 Overview of the eight Cyatheaceae chloroplast genomes and characteristics of their SSRs

	<i>A. denticulata</i>	<i>A. podophylla</i>	<i>A. gigantea</i>	<i>A. metteniana</i>	<i>A. costularis</i>	<i>A. spinulosa</i>
Sequence analyzed (kb)	154.05	166.15	161.68	161.60	156.68	156.66
No. of SSRs	61	67	64	65	121	122
Relative abundance (No./bp)	0.40	0.40	0.40	0.40	0.77	0.78
Total length of SSRs (bp)	634	802	818	801	1539	1538
Relative density (bp/kb)	4.11	4.82	5.06	4.96	9.82	9.81
GC content	0.24	0.25	0.29	0.22	0.20	0.18
Genome content	0.004	0.005	0.005	0.005	0.010	0.010

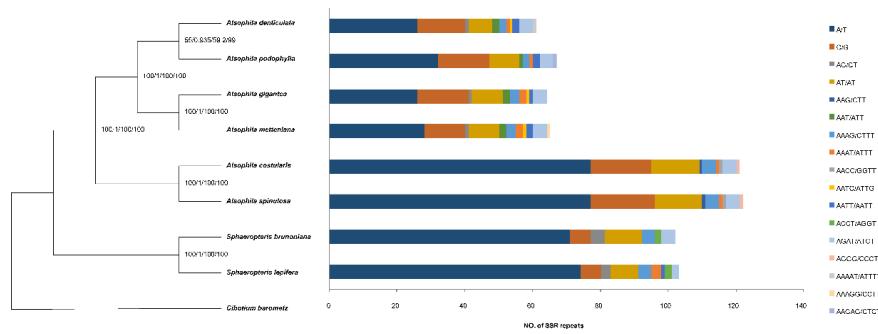


Figure 2 ML, BI, MP, and NJ phylogenetic trees based on eight complete chloroplast genome sequences in Cyatheaceae and the distribution of different types of SSR motifs. The outgroup is *Cibotium barometz*.

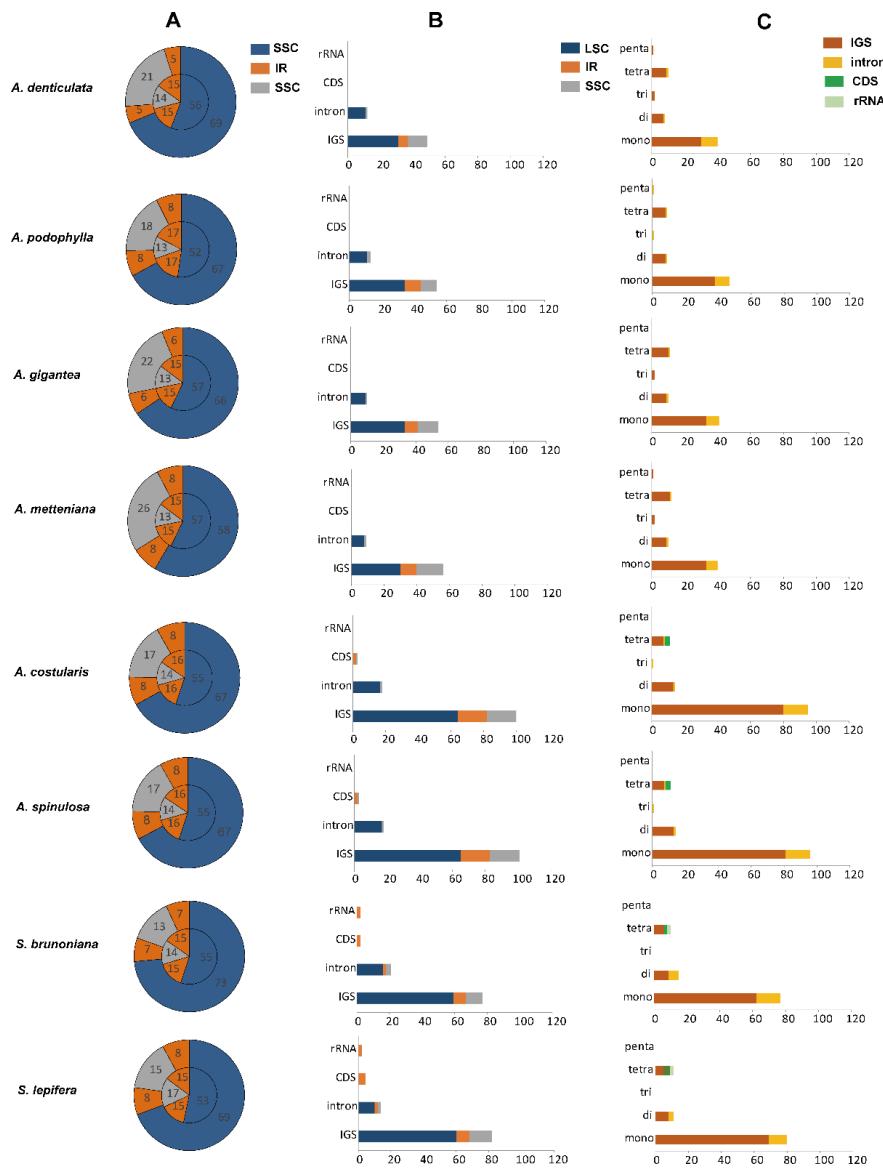


Figure 3 Comparison of microsatellite repeats among the eight Cyatheaceae chloroplast genomes.

(A) The inner circle is the distribution ratio of the four regions (LSC, SSC, and IRs) in the genome, and the outer circle is the distribution ratio of SSRs between the four regions. (B) Distribution ratio of SSRs in different regions (LSC, SSC, and IRs) of chloroplast the genome. (C) Ratio of mono- to pentanucleotide SSRs in different regions (IGS, intron, CDS, and rRNA gene) of the chloroplast genome. Numbers represent the distribution ratio of SSR numbers. LSC: large single-copy region; SSC: small single-copy region; IR: inverted repeat region; IGS: intergenic spacer region; CDS: coding sequence region.

2.3 Analysis of the types and characteristics of SSRs of different nucleotide numbers

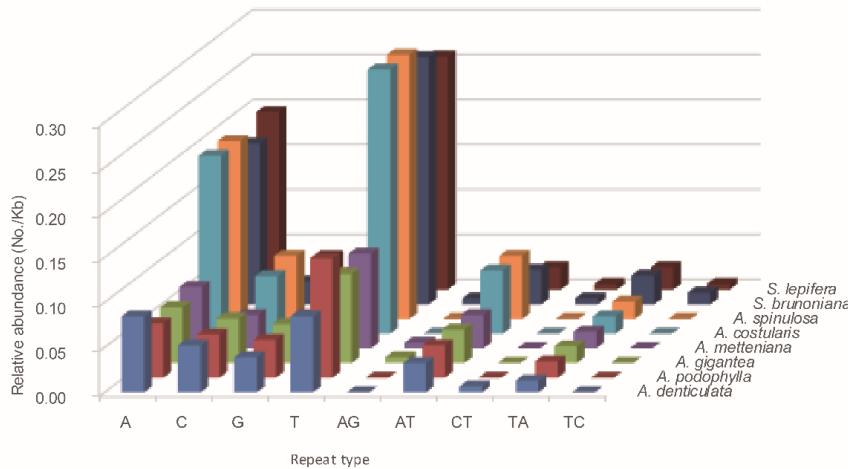
The proportions of single-nucleotide, dinucleotide, trinucleotide, tetranucleotide, and pentanucleotide SSRs

in each species were 62.5-78.0%, 10.6-15.6%, 0-3.3%, 9.0-18.5%, and 0-1.5%, respectively. No hexanucleotide SSR was detected. Among single-nucleotide repeats, there were more A/T motifs, and the dinucleotide repeats were dominated by AT/TA motifs. The single-nucleotide, dinucleotide, trinucleotide, tetranucleotide, and pentanucleotide SSRs of the chloroplast genomes of the three genera were similar in number, relative abundance, relative density, and GC content at the level of the genome and in the specific regions of the genome (LSC, SSC, and IRs; IGS, intron, CDS, and rRNA gene regions). This was especially true for single-nucleotide and dinucleotide SSRs (Table 3, Supplementary Tables 10 and 11). The number, relative abundance, relative density, and GC content of SSRs of different unit lengths and SSRs in different regions of the genome had genus specificity in the phylogenetic context of dividing the eight species of Cyatheaceae into three genera. In addition, the number, relative abundance, and relative density of SSRs of different base types in the chloroplast genomes of the three genera of plants also had genus specificity. This was especially true for single- and dinucleotide SSRs (Figure 4, Supplementary Table 9).

Table 3 The number, relative abundance, relative density, and GC content of mono- to pentanucleotide SSRs in the eight chloroplast genomes of Cyatheaceae

Repeat type	Characteristics of SSR	<i>A. denticulata</i>	<i>A. podophylla</i>	<i>A. gigantea</i>	<i>A. metteniana</i>	<i>A. costularis</i>
Mononucleotide	No. of SSRs	40	47	41	40	95
	Abundance (No./kb)	0.26	0.28	0.25	0.25	0.61
	Density (bp/kb)	2.93	3.16	3.13	2.88	7.67
	GC	0.36	0.35	0.40	0.3	0.22
Dinucleotide	No. of SSRs	8	9	10	10	14
	Abundance (No./kb)	0.05	0.05	0.06	0.06	0.09
	Density (bp/kb)	0.69	0.83	0.94	0.92	1.35
	GC	0.06	0	0.08	0.05	0
Trinucleotide	No. of SSRs	2	1	2	2	1
	Abundance (No./kb)	0.01	0.01	0.01	0.01	0.01
	Density (bp/kb)	0.16	0.07	0.15	0.15	0.08
	GC	0	0	0	0	0.33
Tetranucleotide	No. of SSRs	10	9	11	12	11
	Abundance (No./kb)	0.80	0.05	0.07	0.07	0.07
	Density (bp/kb)	0.14	0.67	0.84	0.92	0.87
	GC	0.18	0.14	0.18	0.16	0.29
Pentanucleotide	No. of SSRs	1	1	0	1	0
	Abundance (No./kb)	0.01	0.01	0	0.01	0
	Density (bp/kb)	0.10	0.09	0	0.09	0
	GC	0	0.4	0	0.4	0

A



B

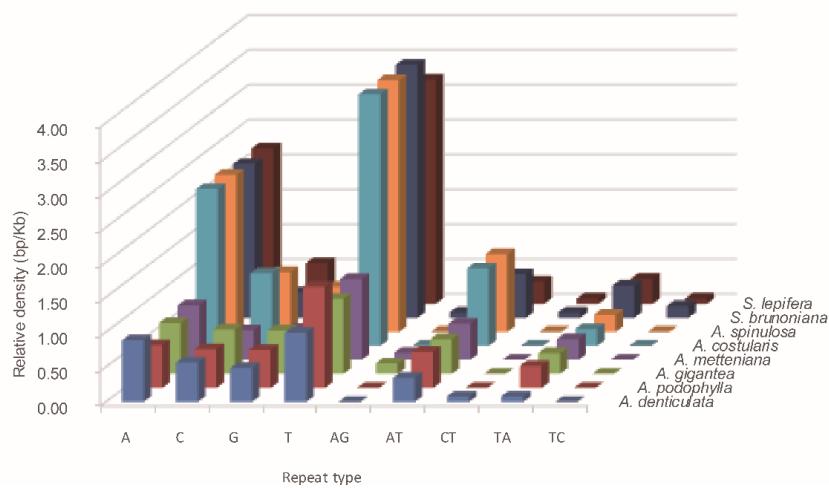


Figure 4 Relative abundance and relative density of single- and dinucleotide SSRs in the eight chloroplast genomes of Cyatheaceae.

2.4 Phylogenetic analysis

The chloroplast genomes of the eight species of Chinese Cyatheaceae were compared globally, and the phylogenetic trees were constructed with four methods (ML, BI, MP, and NJ) using *Cibotium barometz* (Linn.) J. Sm. as an outgroup, as shown in Figure 2. The topologies of the four trees were consistent, except that the support rate of the branches of the *Alsophila denticulata* and *Alsophila gigantea* was lower (the bootstrap values of ML, MP, and NJ were 55%, 59.2%, 99%, respectively, and the posterior probability of BI was 0.935). The support rate of the other branches was higher (the bootstrap values of ML, MP, and NJ were all 100%, and the posterior probability of BI was 1.00). Closely related *Sphaeropteris brunoniana* and *Sphaeropteris*

lepisifera were clustered into one branch, which was located at the base of the phylogenetic tree. That means they were an earlier-diverged group in this family. *Alsophila denticulata*, *Alsophila podophylla*, *Alsophila gigantea*, and *Alsophila metteniana* were clustered into one branch, which was located inside the branch of *Sphaeropteris brunonianana* and *Sphaeropteris lepisifera* and was a sister group of the branch formed by *Alsophila spinulosa* and *Alsophila costularis*.

2.5 Statistical analysis of the results

The Kruskal-Wallis H-test and Mann-Whitney U-test values are shown in Supplementary Table 11. In this study, only SSRs in the chloroplast genome in its IGS and LSC regions, as well as single-nucleotide SSRs in the whole chloroplast genome, were considered. There were significant differences in the number, relative abundance, relative density, and GC content of SSRs when the eight species of Cyatheaceae were divided into three genera (Kruskal-Wallis H, $P < 0.05$). When *Gymnosphaera* was included in the genus *Alsophila*, they were divided into two genera, and only the difference in the GC content was significant (Table 4). The number of SSRs in other regions of the chloroplast genome and the number of SSRs of other unit lengths were small, so they are not discussed in this study. The clustering results of the number, relative abundance, relative density, and GC content of the SSRs in the chloroplast genomes and their IGS regions and the single-nucleotide SSRs of the whole chloroplast genome of the eight species of Cyatheaceae (Figure 5) showed that the eight species were divided into two groups. That is, *Sphaeropteris brunonianana*, *Sphaeropteris lepisifera*, *Alsophila spinulosa* and *Alsophila costularis* were in a group, and *Alsophila denticulata*, *Alsophila podophylla*, *Alsophila metteniana*, and *Alsophila gigantea* were in a group.

Table 4 Significance test of the number, relative abundance, relative density, and GC content of SSRs in the whole chloroplast genome, IGS, and LSC and mononucleotide SSRs in the whole chloroplast genome of the eight Cyatheaceae species

Classification treatment	Genus	Species	Characteristics of SSR	<i>P</i>	<i>P</i>	<i>P</i>	<i>P</i>	Literature cited
Three genera	<i>Gymnosphaera</i>	<i>A. denticulata</i>	No. of SSRs	0.048*	0.048*	0.048*	0.048*	Smith et al., 2000
		<i>A. podophylla</i>						Korall et al., 2001
		<i>A. gigantea</i>	Relative abundance (No./kb)	0.033*	0.050 *	0.050 *	0.050 *	Janssen et al., 2000
		<i>A. metteniana</i>						Korall et al., 2001
	<i>Alsophila</i>	<i>A. costularis</i>	Relative density (bp/kb)	0.050 *	0.050 *	0.050 *	0.050 *	Pryer, 2014;
		<i>A. spinulosa</i>						Ching, 1978;
	<i>Sphaeropteris</i>	<i>S. brunonianana</i>	GC content	0.050 *	0.050 *	0.050 *	0.050 *	Dong, 2019.
		<i>S. lepisifera</i>						

Classification treatment	Genus	Species	Characteristics of SSR	P	P	P	P	Literature cited
Two genera	<i>Alsophila</i>	<i>A. denticulata</i> <i>A. podophylla</i>	No. of SSRs	0.502	0.502	0.502	0.502	PPG I, 2016; X 1989; Zhang & Nishida, 2013.
		<i>A. gigantea</i> <i>A. metteniana</i> <i>A. costularis</i> <i>A. spinulosa</i>	Relative abundance (No./kb)	0.478	0.505	0.505	0.505	
	<i>Sphaeropteris</i>	<i>S. brunonianana</i> <i>S. lepifera</i>	Relative density (bp/kb)	0.505	0.505	0.505	0.505	
			GC content	0.046*	0.046*	0.046*	0.046*	

When Cyatheaceae plants are treated as three genera, the Kruskal-Wallis H test is used; when treated as two genera, the Mann-Whitney U test is used; *: P [?] 0.05; IGS: intergenic spacer; LSC: large single-copy region.

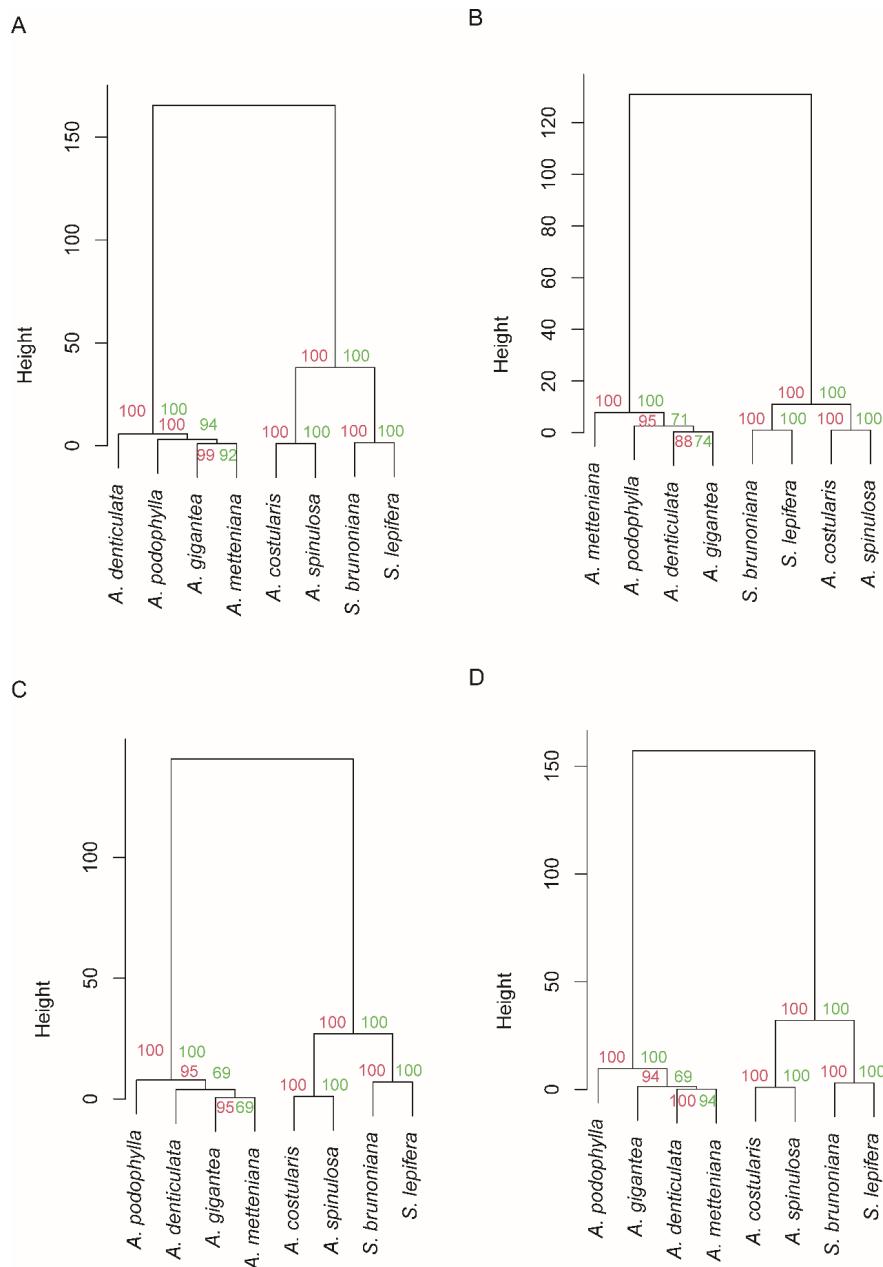


Figure 5 Clustering analysis of 8 Cyatheaceae species based on the number, relative abundance, relative density, and GC content of SSRs across the whole chloroplast genome (A),LSC (B), IGS (C) and mononucleotide (D) SSRs in the chloroplast genome.

3 Discussion

3.1 Characteristics of SSRs of chloroplast genomes of eight species of Cyatheaceae

The chloroplast genomes of all eight species of Cyatheaceae are similar in structure and gene content, and the types and order of genes are the same. On the phylogenetic background of dividing the eight species of Cyatheaceae into three genera, the characteristics of the chloroplast genome SSRs have genus specificity. The distribution of SSRs is not random, which has an effect on gene regulation, DNA recombination, DNA

replication, the cell cycle, and DNA mismatch repair, and the presence of repeat motifs and their repeat number can affect DNA recombination (Li et al., 2002). SSR copy number is an important source of genetic variation and can produce large phenotypic variation (Gemayel et al., 2010; Kashi & King, 2006). Single-nucleotide SSRs are the most abundant, which is a characteristic of eukaryotic genomes (Sharma et al., 2007), and A/T motifs are the most common. The number, relative abundance, relative density, and GC content of SSRs of different unit length are also genus specific. This is especially true for single-nucleotide and dinucleotide SSRs, which may be related to the lower content of SSRs of other unit lengths. The distribution of different repeat types (from single-nucleotide to hexanucleotide) of motifs in coding and noncoding regions, introns, and intergenic regions displays a high degree of taxon specificity, which can be partially explained by the interaction of mutation mechanisms and differential selection (Toth et al., 2000). The SSRs in eukaryotic genomes are mainly located in intergenic and noncoding regions, with a few in exons (Toth et al., 2000; Li et al., 2004), and the results of this study are consistent with this. This phenomenon is related to the higher variability (Nie et al., 2012; Wu et al., 2010) and faster evolutionary rates of the intergenic and noncoding regions of the chloroplast genome, so their sequences can be used to effectively classify low taxonomic and closely related groups and subspecies variant plants. The proportion of SSRs in the IR region was 2-3.3 times the proportion of IR sequences out of the whole genome sequence (Figure 3), indicating that there are fewer SSRs in the IR region. Mismatch repair is the key to the stability of SSRs. SSRs have a high mutation rate, which facilitates the study of the effect of environmental factors on the mutation rate of the genome. The lower distribution of SSRs in the IR region may be related to the lower mutation rate in the IR region (Ellegren, 2004; Li et al., 2016). The lower GC content may be associated with the fact that GC-rich regions are prone to mutations toward AT(Ren et al., 2007). The high GC content is significantly associated with the high recombination rate in meiosis (Tortereau et al., 2012), while AT-rich SSRs may be more conducive to maintaining the stability of the genome structure. The number, relative abundance, relative density, and GC content of SSRs are not proportional to the size of genome, indicating that the abundance of SSRs is related to the genetic characteristics of the species (Li et al., 2014).

3.2 Phylogenetic significance of SSR characteristics of chloroplast genomes of the eight species of Cyatheaceae

The focus of the debate between the Holttum and Edwards (1963) system and the Tryon (1970) system, which classify Cyatheaceae based on morphological characteristics, is the theoretical explanation of the morphological evolution of indusium in this family. In this study, we show that the genus *Sphaeropteris*, lacking the indusium, is a basal group, supporting Tryon's hypothesis that the indusium is derived from the tissues or scales on the abaxial side of the leaf, away from the leaf margin, and that the indusium is a derived trait, which is consistent with a phylogenetic analysis based on the chloroplast *trnL* intron sequence and the *trnL-F* intergenic region sequence (Wang et al., 2003). Dong (2018) pointed out that *Gymnosphaera* and *Alsophila* were significantly differentiated in morphological traits such as petiole color, the presence or absence of degenerated pinnae at the base, and the presence or absence of indusium and sporogenesis, and advocated the restoration of the hierarchical status of the genus *Gymnosphaera* to reflect the divergence mechanisms of this group of plants in molecular phylogeny, morphology, and sporogenesis. In this study, eight species of Cyatheaceae were divided into three genera or two genera, in which case SSRs were compared by the Kruskal-Wallis H test or the Mann-Whitney U test, respectively, and the results showed that *Gymnosphaera* was an independent genus-level taxon under the Cyatheaceae family. The results of the PV clustering analysis of SSRs also indicated that *Gymnosphaera* should be independent from the genus *Alsophila*. Phylogenetic trees were constructed using the chloroplast genomes of eight species of Cyatheaceae, and the topology of the phylogenetic trees obtained by the four methods was consistent. Except for the lower support rate of the branches of the *Alsophila denticulata* and *Alsophila gigantea*, the support rate for the branches was 100%. This result supports the monophyletic nature of *Gymnosphaera*, and its sister group is the genus *Alsophila*. *Gymnosphaera*, *Alsophila*, and *Cyathea* constitute a monophyletic group with a high support rate, while *Sphaeropteris* resolved as the basal group of the Cyatheaceae family. In this phylogenetic context, the SSR characteristics have genus specificity.

SSRs play a role in genome-wide regulation. Some definite distribution patterns exist in the genomes of

organisms, and the characteristic distribution of SSRs in the genomes of different taxonomic units has a significantly similar pattern (Qi et al., 2015; Wang et al., 2015; Liu et al., 2017; Manee et al., 2019; Srivastava et al., 2019). The SSRs of different groups of genomes have specific distribution patterns, which are related to their common ancestors. Evolutionary trends have been linked to the inclusion of SSRs, which may have been preserved because of their ability to adapt to novel regulatory mechanisms (Srivastava et al., 2019). Unique DNA replication, repair, and recombination mechanisms may play an important role in the evolution of SSRs (Katti et al., 2001). The molecular mechanism of the origin of SSRs is not yet fully understood. The most common mutation mechanism affecting SSRs is slipped replication. Other mechanisms, such as unequal crossing-over, nucleotide substitution, and duplication events, are also responsible for SSR variation (Schlotterer & Tautz, 1992; Hancock, 1999). Codon preference, DNA replication, and mismatch repair systems, as well as the unique structure and function of the genome, may be responsible for the unique SSR distribution pattern in plant genomes. In addition, the length, motif structure, and GC content of genomic SSRs are also factors that influence the evolution of the SSRs (Chakraborty et al., 1997; Anderson et al., 2000; Whittaker et al., 2003). The plants in the three genera of Cyatheaceae have similar phenotypes and specific characteristics, which may be the result of interactions of their common ancestors with similar habitats. The analysis of the characteristics of SSRs provides useful clues for the phylogenetic study of Cyatheaceae and helps to understand the evolution of SSRs in plant genomes.

The chloroplast genome has a simple structure, low molecular weight, and high copy number, and its genes are maternally inherited in ferns with few gene rearrangements, thus facilitating the study of plant phylogeny (Tonti-Filippini et al., 2017). Based on high-throughput sequencing technology, the highly conserved chloroplast genome sequence will make primer design easier. SSR markers in chloroplast genomes can be used for the analysis of multiple chloroplast regions, thereby improving the resolution of phylogenetic studies of target species (Melotto-Passarin et al., 2011). Since the software programs that identify SSRs are limited by their efficiency and parameter settings and may also be affected by the quality of the SSR dataset generated, their accuracy needs to be improved (Ellegren, 2004; Lim et al., 2013). In addition, chloroplast genomes play an important role in dissecting the higher hierarchical phylogenetic relationships of ferns. However, in the process of evolution, plants experience events such as hybridization, polyploidization, introgression, and incomplete lineage sorting, so the evolutionary relationship of plants is essentially reticular. Such reticular relationships are often manifested in the form of gene tree conflicts (Guo & Ge, 2005).

In this study, the chloroplast genomes of eight species of Cyatheaceae were used to construct phylogenetic trees. This analytical method has some limitations. There are still relatively few studies on phylogenetic relationships in ferns that have analyzed the SSRs of the chloroplast genome. This study provides a new basis for the classification of Cyatheaceae at the levels of species and genus, thus advancing the phylogenetic study of Cyatheaceae. In the future, more genomic and transcriptomic data are needed to validate these results.

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Conflict of interest

The authors declare no competing interests.

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helpful comments.

Author contributions

MZ conducted the statistics and analysis of the data, and wrote the manuscript. PPF, JYP and JYL analyzed part of the data and provided Suggestions. YJS and TW designed the study and wrote the manuscript.

Data availability statement

The chloroplast genomes of *Alsophila denticulata* Baker and *Alsophila metteniana* Hance have been submitted to the National Center for Biotechnology Information (NCBI) and have not yet been published. The other 6 chloroplast genomes of Cyatheaceae species can be downloaded from NCBI.

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Appendix table 1-8 List of simple sequence repeats(SSRs) in the chloroplast genomes of 8 species in Cyatheaceae.

Appendix table 9 The number, relative abundance, and relative density of SSR motifs in the chloroplast genomes of 8 species in Cyatheaceae.

Appendix table 10 The number, relative abundance, relative density and GC content of different types of motif SSRs in different regions in the chloroplast genomes of 8 species in Cyatheaceae.

Appendix table 11 Significant differences in the number, relative abundance, relative density, and GC content of the chloroplast genomes, Mono- to Pentanucleotide SSRs, IGS, LSC, intron and CDS regions SSRs of chloroplast genomes, and the mono- to pentanucleotide SSRs in the chloroplast genomes of 8 Cyatheaceae species

Appendix table 1 List of simple sequence repeats (SSRs) in the chloroplast genomes of *Alsophila denticulata*

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
1	p5	spacer	LSC	ATTTT	15	5331	5345	<i>rps16-chlB</i>
2	p1	spacer	LSC	G	14	7040	7053	<i>chlB-trnQ(UGG)</i>
3	p1	spacer	LSC	C	11	8373	8383	<i>trnS(GCU)-psaM</i>
4	p1	intron	LSC	A	12	9401	9412	<i>trnG(UCC)</i>
5	p4	intron	LSC	AATT	12	9648	9659	<i>trnG(UCC)</i>
6	p1	spacer	LSC	G	16	13299	13314	<i>atpF-atpH</i>
7	p1	spacer	LSC	C	12	14280	14291	<i>atpH-atpI</i>
8	p1	spacer	LSC	C	12	15231	15242	<i>atpI-rps2</i>
9	p1	intron	LSC	C	10	22475	22484	<i>rpoC1</i>
10	p1	spacer	LSC	A	11	23468	23478	<i>rpoC1-rpoB</i>
11	p1	spacer	LSC	C	10	26793	26802	<i>rpoB-trnD(GUC)</i>
12	p4	spacer	LSC	ATAG	12	27725	27736	<i>rpoB-trnD(GUC)</i>
13	p1	spacer	LSC	T	13	28419	28431	<i>trnE(UUC)-psbM</i>
14	p4	spacer	LSC	TGAT	12	28511	28522	<i>trnE(UUC)-psbM</i>
15	p1	spacer	LSC	A	10	29560	29569	<i>ycf66</i>
16	p1	spacer	LSC	G	14	29640	29653	<i>ycf66</i>
17	p1	spacer	LSC	G	10	31641	31650	<i>trnC(GCA)-trnG(GCC)</i>
18	p1	spacer	LSC	A	10	31715	31724	<i>trnC(GCA)-trnG(GCC)</i>
19	p1	spacer	LSC	C	10	32102	32111	<i>trnG(GCC)-psbZ</i>
20	p4	spacer	LSC	TATC	16	33543	33558	<i>trnS(UGA)-psbC</i>
21	p4	spacer	LSC	ATAG	12	33562	33573	<i>trnS(UGA)-psbC</i>
22	p1	spacer	LSC	A	10	37593	37602	<i>psbD-trnT(GGU)</i>
23	p1	intron	LSC	A	11	44104	44114	<i>ycf3</i>

SSR nr.	SSR type	Locus	Region	SSR motif	size	start	end	Location
24	p1	intron	LSC	T	13	44123	44135	<i>ycf3</i>
25	p1	intron	LSC	C	11	45216	45226	<i>ycf3</i>
26	p1	spacer	LSC	T	10	45601	45610	<i>ycf3-trnS(GGA)</i>
27	p1	spacer	LSC	G	10	47798	47807	<i>rps4-trnL(CAA)</i>
28	p1	spacer	LSC	A	10	51902	51911	<i>trnV(UAC)-trnM(CAU)</i>
29	p1	spacer	LSC	T	11	56710	56720	<i>trnR(UCG)-accD</i>
30	p4	spacer	LSC	TATT	12	57765	57776	<i>accD-psaI</i>
31	p2	spacer	LSC	AT	18	58231	58248	<i>accD-psaI</i>
32	p2	spacer	LSC	TA	12	62570	62581	<i>petA-psbJ</i>
33	p1	spacer	LSC	A	10	68052	68061	<i>rpl20-rps12</i>
34	p1	spacer	LSC	T	11	68120	68130	<i>rpl20-rps12</i>
35	p4	spacer	LSC	GATA	12	69048	69059	<i>rps12-clpP</i>
36	p1	intron	LSC	C	11	69574	69584	<i>chlP</i>
37	p1	intron	LSC	T	11	69615	69625	<i>chlP</i>
38	p1	intron	LSC	T	11	70413	70423	<i>chlP</i>
39	p2	intron	LSC	CT	12	70704	70715	<i>chlP</i>
40	p2	spacer	LSC	AT	12	79990	80001	<i>rps8-rpl14</i>
41	p1	intron	LSC	T	12	81564	81575	<i>rpl16</i>
42	p1	spacer	LSC	T	10	85769	85778	<i>rpl23-trnI(CAU)</i>
43	p2	spacer	IRA	AT	12	87653	87664	<i>trnT(UGU)</i>
44	p4	spacer	IRA	TCTT	12	96474	96485	<i>rrn16-rps12</i>
45	p1	spacer	IRA	A	12	97573	97584	<i>rrn16-rps12</i>
46	p1	spacer	SSC	A	10	111684	111693	<i>ndhF-rpl21</i>
47	p3	spacer	SSC	AAT	12	112846	112857	<i>rpl32-trnP(GGG)</i>
48	p2	spacer	SSC	TA	12	112860	112871	<i>rpl32-trnP(GGG)</i>
49	p3	spacer	SSC	TAT	12	112873	112884	<i>rpl32-trnP(GGG)</i>
50	p1	spacer	SSC	A	10	113225	113234	<i>trnP(GGG)-trnL(UAG)</i>
51	p1	spacer	SSC	T	11	113467	113477	<i>trnL(UAG)-ccsA</i>
52	p1	spacer	SSC	A	10	116602	116611	<i>ndhD-psaC</i>
53	p4	spacer	SSC	AATT	12	116610	116621	<i>ndhD-psaC</i>
54	p1	spacer	SSC	G	11	117154	117164	<i>psaC-ndhE</i>
55	p1	spacer	SSC	T	11	118927	118937	<i>ndhI-ndhA</i>
56	p2	spacer	SSC	AT	16	118966	118981	<i>ndhI-ndhA</i>
57	p1	intron	SSC	T	17	120503	120519	<i>ndhA</i>
58	p1	spacer	SSC	A	10	122795	122804	<i>rps15-ycf1</i>
59	p1	spacer	IRB	T	12	142438	142449	<i>rps12-rrn16</i>
60	p4	spacer	IRB	AAAG	12	143536	143547	<i>rps12-rrn16</i>
61	p2	spacer	IRB	AT	12	152358	152369	<i>trnT(UGU)</i>

Appendix table 2 List of simple sequence repeats (SSRs) in the chloroplast genomes of *Alsophila podophylla*

SSR nr.	SSR type	Locus	Region	SSR motif	size	start	end	Location
1	p1	spacer	LSC	G	13	2113	2125	<i>ndhB-matK</i>
2	p1	intron	LSC	T	10	5156	5165	<i>rps16</i>
3	p1	spacer	LSC	G	16	7667	7682	<i>chlB-trnQ(UUG)</i>
4	p2	spacer	LSC	AT	14	9108	9121	<i>trnS(GUC)-psaM</i>
5	p1	spacer	LSC	G	11	9713	9723	<i>ycf12-trnG(UCC)</i>
6	p4	intron	LSC	AATT	12	10328	10339	<i>trnG(UCC)</i>
7	p1	spacer	LSC	C	14	14948	14961	<i>atpH-atpI</i>

SSR nr.	SSR type	Locus	Region	SSR motif	size	start	end	Location
8	p1	spacer	LSC	C	11	15898	15908	<i>atpI-rps2</i>
9	p1	spacer	LSC	C	11	27484	27494	<i>rpoB-trnD(GUC)</i>
10	p4	spacer	LSC	ATAG	12	28409	28420	<i>rpoB-trnD(GUC)</i>
11	p1	spacer	LSC	T	12	29103	29114	<i>trnE(UUC)-psbM</i>
12	p4	spacer	LSC	TGAT	12	29197	29208	<i>trnE(UUC)-psbM</i>
13	p1	spacer	LSC	G	13	30325	30337	<i>ycf66</i>
14	p1	spacer	LSC	A	10	32400	32409	<i>trnC(GCA)-trnG(GCC)</i>
15	p1	spacer	LSC	C	12	32787	32798	<i>trnG(GCC)-psbZ</i>
16	p4	spacer	LSC	TATC	16	34223	34238	<i>trnS(UGA)-psbC</i>
17	p4	spacer	LSC	ATAG	12	34242	34253	<i>trnS(UGA)-psbC</i>
18	p1	spacer	LSC	T	10	37895	37904	<i>psbD-trnT(GGU)</i>
19	p5	intron	LSC	TTCTC	15	39317	39331	<i>rps14</i>
20	p3	intron	LSC	AAT	12	44793	44819	<i>ycf3</i>
21	p1	intron	LSC	T	10	44810	44819	<i>ycf3</i>
22	p1	intron	LSC	C	11	45899	45909	<i>ycf3</i>
23	p1	spacer	LSC	T	10	46284	46293	<i>ycf3-trnS(GGA)</i>
24	p1	spacer	LSC	T	10	46867	46876	<i>trnS(GGA)-rps4</i>
25	p1	spacer	LSC	G	12	48604	48615	<i>rps4-trnL(CAA)</i>
26	p1	spacer	LSC	T	10	57501	57510	<i>trnR(UCG)-accD</i>
27	p4	spacer	LSC	TATT	12	58555	58566	<i>accD-psaI</i>
28	p1	spacer	LSC	A	10	58615	58624	<i>accD-psaI</i>
29	p2	spacer	LSC	AT	18	59022	59039	<i>accD-psaI</i>
30	p2	spacer	LSC	TA	14	63376	63389	<i>petA-psbJ</i>
31	p1	spacer	LSC	T	10	66579	66588	<i>trnW(CCA)-trnP(UGG)</i>
32	p1	spacer	LSC	C	12	67291	67302	<i>psaJ-rpl33</i>
33	p1	spacer	LSC	A	10	68892	68901	<i>rpl20-rps12</i>
34	p1	spacer	LSC	T	11	68960	68970	<i>rpl20-rps12</i>
35	p4	spacer	LSC	GATA	12	69890	69901	<i>rps12-clpP</i>
36	p1	intron	LSC	T	10	70443	70452	<i>clpP</i>
37	p1	intron	LSC	T	11	71239	71249	<i>clpP</i>
38	p1	intron	LSC	T	11	75002	75012	<i>petB</i>
39	p1	spacer	LSC	T	10	77981	77990	<i>petD-rpoA</i>
40	p1	spacer	LSC	A	10	79921	79930	<i>rpl36-infa</i>
41	p1	spacer	LSC	T	11	80751	80761	<i>rps8-rpl14</i>
42	p1	spacer	LSC	A	10	80792	80801	<i>rps8-rpl14</i>
43	p2	spacer	LSC	AT	16	80801	80816	<i>rps8-rpl14</i>
44	p1	intron	LSC	T	13	82378	82390	<i>rpl16</i>
45	p1	intron	LSC	T	10	82733	82742	<i>rpl16</i>
46	p2	spacer	IRB	AT	12	87789	87800	<i>trnT(UGU)</i>
47	p1	spacer	IRB	A	10	94889	94898	<i>trnI(GAU)-rrn16</i>
48	p1	spacer	IRB	A	12	104005	104016	<i>rrn16-rps12</i>
49	p1	spacer	IRB	A	10	107991	108000	<i>trnH(GUG)-ycf2</i>
50	p1	spacer	IRB	C	10	115418	115427	<i>trnN(GUU)-ndhF</i>
51	p1	spacer	SSC	A	10	117925	117934	<i>ndhF-rpl21</i>
52	p2	spacer	SSC	TA	22	119288	119309	<i>rpl32-trnP(GGG)</i>
53	p4	spacer	SSC	AAAT	12	119665	119676	<i>trnP(GGG)-trnL(UAG)</i>
54	p1	spacer	SSC	T	13	119901	119913	<i>trnL(UAG)-ccsA</i>
55	p1	spacer	SSC	A	10	123033	123042	<i>ndhD-psaC</i>
56	p4	spacer	SSC	AATT	12	123041	123052	<i>ndhD-psaC</i>
57	p1	spacer	SSC	G	15	123587	123601	<i>psaC-ndhE</i>

SSR nr.	SSR type	Locus	Region	SSR motif	size	start	end	Location
58	p1	spacer	SSC	T	10	125374	125383	<i>ndhI-ndhA</i>
59	p2	spacer	SSC	AT	14	125418	125431	<i>ndhI-ndhA</i>
60	p2	intron	SSC	TA	16	126454	126469	<i>ndhA</i>
61	p1	intron	SSC	T	17	126960	126976	<i>ndhA</i>
62	p1	spacer	SSC	C	11	134606	134616	<i>ycf1-chlN</i>
63	p1	spacer	IRA	G	10	137487	137496	<i>chlL-trnN(GUU)</i>
64	p1	spacer	IRA	T	10	144914	144923	<i>ycf2-trnH(GUG)</i>
65	p1	spacer	IRA	T	12	148898	148909	<i>rps12-rrn16</i>
66	p1	spacer	IRA	T	10	158016	158025	<i>rrn16-trnI(GAU)</i>
67	p2	spacer	IRA	AT	12	165114	165125	<i>trnT(UGU)</i>

Appendix table 3 List of simple sequence repeats (SSRs) in the chloroplast genomes of *Alsophila gigantea*

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
1	p1	spacer	LSC	G	10	3677	3686	<i>matK-rps16</i>
2	p1	spacer	LSC	G	11	7015	7025	<i>chlB-trnQ(UUG)</i>
3	p2	spacer	LSC	AT	14	8451	8464	<i>trnS(GUC)-psaM</i>
4	p4	intron	LSC	AATT	12	9622	9633	<i>trnG(UCC)</i>
5	p1	spacer	LSC	C	10	14251	14260	<i>atpH-atpI</i>
6	p1	spacer	LSC	C	10	15197	15206	<i>atpI-rps2</i>
7	p1	intron	LSC	C	10	22460	22469	<i>rpoC1</i>
8	p1	spacer	LSC	T	10	27192	27201	<i>rpoB-trnD(GUC)</i>
9	p4	spacer	LSC	ATAG	12	27704	27715	<i>rpoB-trnD(GUC)</i>
10	p4	spacer	LSC	TGAT	12	28490	28501	<i>trnE(UUC)-psbM</i>
11	p1	spacer	LSC	G	17	29622	29638	<i>ycf66</i>
12	p1	spacer	LSC	G	11	31628	31638	<i>trnC(GCA)-trnG(GCC)</i>
13	p1	spacer	LSC	C	10	32088	32097	<i>trnG(GCC)-psbZ</i>
14	p4	spacer	LSC	TATC	12	33522	33533	<i>trnS(UGA)-psbC</i>
15	p4	spacer	LSC	ATAG	12	33537	33548	<i>trnS(UGA)-psbC</i>
16	p1	spacer	LSC	T	10	36642	36651	<i>psbD-trnT(GGU)</i>
17	p2	spacer	LSC	AG	24	36747	36770	<i>psbD-trnT(GGU)</i>
18	p1	intron	LSC	A	10	44100	44109	<i>ycf3</i>
19	p1	intron	LSC	T	12	44122	44133	<i>ycf3</i>
20	p1	spacer	LSC	T	10	45598	45607	<i>ycf3-trnS(GGA)</i>
21	p1	spacer	LSC	G	10	47441	47450	<i>rps4-trnL(CAA)</i>
22	p1	spacer	LSC	G	10	47936	47945	<i>rps4-trnL(CAA)</i>
23	p1	spacer	LSC	A	14	48913	48926	<i>trnL(CAA)-trnF(GAA)</i>
24	p1	spacer	LSC	T	10	49833	49842	<i>ndhJ-ndhK</i>
25	p1	spacer	LSC	A	11	52073	52083	<i>trnV(UAC)-trnM(CAU)</i>
26	p1	spacer	LSC	T	11	54541	54551	<i>atpB-rbcL</i>
27	p1	spacer	LSC	T	10	56891	56900	<i>trnR(UCG)-accD</i>
28	p4	spacer	LSC	TATT	12	57947	57958	<i>accD-psaI</i>
29	p2	spacer	LSC	AT	14	58413	58426	<i>accD-psaI</i>
30	p2	spacer	LSC	TA	16	62759	62774	<i>petA-psbJ</i>
31	p1	spacer	LSC	C	11	72815	72825	<i>psaJ-rpl33</i>
32	p1	spacer	LSC	A	11	74414	74424	<i>rpl20-rps12</i>
33	p1	spacer	LSC	T	10	74483	74492	<i>rpl20-rps12</i>
34	p4	spacer	LSC	GATA	12	75418	75429	<i>rps12-clpP</i>
35	p1	intron	LSC	C	11	75940	75950	<i>clpP</i>

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
36	p1	intron	LSC	T	10	75981	75990	<i>clpP</i>
37	p1	intron	LSC	T	11	81166	81176	<i>petB</i>
38	p1	spacer	LSC	A	11	86310	86320	<i>rps8-rpl14</i>
39	p2	spacer	LSC	AT	16	86320	86335	<i>rps8-rpl14</i>
40	p1	spacer	LSC	T	11	86335	86345	<i>rps8-rpl14</i>
41	p1	intron	LSC	T	12	88251	88262	<i>rpl16</i>
42	p1	intron	LSC	C	10	91441	91450	<i>rpl2</i>
43	p2	spacer	IRB	AT	12	93990	94001	<i>trnT(UGU)</i>
44	p1	spacer	IRB	A	12	101110	101121	<i>trnI(GAU)-rrn16</i>
45	p4	spacer	IRB	TCTT	12	103414	103425	<i>rrn16-rps12</i>
46	p1	spacer	IRB	A	12	104529	104540	<i>rrn16-rps12</i>
47	p3	spacer	SSC	AAT	12	119784	119795	<i>rpl32-trnP(GGG)</i>
48	p2	spacer	SSC	TA	18	119798	119815	<i>rpl32-trnP(GGG)</i>
49	p3	spacer	SSC	TAT	12	119817	119828	<i>rpl32-trnP(GGG)</i>
50	p4	spacer	SSC	GAAA	12	120167	120178	<i>trnP(GGG)-trnL(UAG)</i>
51	p1	spacer	SSC	T	13	120409	120421	<i>trnL(UAG)-ccsA</i>
52	p1	spacer	SSC	A	11	121680	121690	<i>ccsA-ndhD</i>
53	p1	spacer	SSC	A	13	123545	123557	<i>ndhD-psaC</i>
54	p1	spacer	SSC	T	10	123566	123575	<i>ndhD-psaC</i>
55	p1	spacer	SSC	G	31	124104	124134	<i>psaC-ndhE</i>
56	p1	spacer	SSC	A	24	125949	125961	<i>ndhI-ndhA</i>
57	p2	spacer	SSC	AT	12	125961	125972	<i>ndhA</i>
58	p2	intron	SSC	TA	14	126999	127012	<i>ndhA</i>
59	p1	spacer	SSC	C	31	135145	135175	<i>ycf1-chlN</i>
60	p4	spacer	SSC	AAAT	16	135394	135409	<i>ycf1-chlN</i>
61	p1	spacer	IRA	T	12	149455	149466	<i>rps12-rrn16</i>
62	p4	spacer	IRA	AAAG	12	150569	150580	<i>rps12-rrn16</i>
63	p1	spacer	IRA	T	12	152874	152885	<i>rrn16-trnI(GAU)</i>
64	p2	spacer	IRA	AT	12	159994	160005	<i>trnT(UGU)</i>

Appendix table 4 List of simple sequence repeats (SSRs) in the chloroplast genomes of *Alsophila gigantea*

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
1	p1	spacer	LSC	G	12	7094	7105	<i>chlB-trnQ(UUG)</i>
2	p1	spacer	LSC	T	10	8416	8425	<i>trnS(GUC)-psaM</i>
3	p2	spacer	LSC	AT	14	8532	8545	<i>trnS(GUC)-psaM</i>
4	p4	intron	LSC	AATT	12	9703	9714	<i>trnG(UCC)</i>
5	p1	intron	LSC	C	10	22538	22547	<i>trnG(UCC)</i>
6	p1	spacer	LSC	T	10	27270	27279	<i>rpoB-trnD(GUC)</i>
7	p4	spacer	LSC	ATAG	12	27782	27793	<i>rpoB-trnD(GUC)</i>
8	p4	spacer	LSC	TGAT	12	28568	28579	<i>trnE(UUC)-psbM</i>
9	p1	spacer	LSC	G	14	29700	29713	<i>ycf66</i>
10	p1	spacer	LSC	G	11	31714	31724	<i>trnC(GCA)-trnG(GCC)</i>
11	p1	spacer	LSC	C	12	32174	32185	<i>trnG(GCC)-psbZ</i>
12	p4	spacer	LSC	TATC	12	33610	33621	<i>trnS(UGA)-psbC</i>
13	p4	spacer	LSC	ATAG	12	33625	33636	<i>trnS(UGA)-psbC</i>
14	p1	spacer	LSC	T	10	36730	36739	<i>psbD-trnT(GGU)</i>
15	p2	spacer	LSC	AG	16	36835	36850	<i>psbD-trnT(GGU)</i>
16	p1	intron	LSC	T	13	44201	44213	<i>ycf3</i>

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
17	p1	spacer	LSC	T	10	45678	45687	<i>ycf3-trnS(GGA)</i>
18	p1	spacer	LSC	G	10	47521	47530	<i>rps4-trnL(CAA)</i>
19	p1	spacer	LSC	G	10	48027	48036	<i>rps4-trnL(CAA)</i>
20	p1	spacer	LSC	A	13	48988	49000	<i>trnL(CAA)-trnF(GAA)</i>
21	p1	spacer	LSC	T	10	49907	49916	<i>ndhJ-ndhK</i>
22	p1	spacer	LSC	A	10	52147	52156	<i>trnV(UAC)-trnM(CAU)</i>
23	p1	spacer	LSC	T	11	54614	54624	<i>atpB-rbcL</i>
24	p1	spacer	LSC	T	10	56964	56973	<i>trnR(UCG)-accD</i>
25	p4	spacer	LSC	TATT	12	58020	58031	<i>accD-psaI</i>
26	p2	spacer	LSC	AT	14	58486	58499	<i>accD-psaI</i>
27	p2	spacer	LSC	TA	16	62833	62848	<i>petA-psbJ</i>
28	p5	spacer	LSC	CTTTC	15	71752	71766	<i>petG-trnW(CCA)</i>
29	p1	spacer	LSC	C	12	72879	72890	<i>psaJ-rpl33</i>
30	p1	spacer	LSC	A	10	74479	74488	<i>rpl20-rps12</i>
31	p1	spacer	LSC	T	10	74547	74556	<i>rpl20-rps12</i>
32	p4	spacer	LSC	GATA	12	75482	75493	<i>rps12-clpP</i>
33	p1	intron	LSC	C	10	76004	76013	<i>clpP</i>
34	p1	intron	LSC	T	14	76044	76057	<i>clpP</i>
35	p1	intron	LSC	T	10	81233	81242	<i>petB</i>
36	p2	spacer	LSC	AT	18	86384	86401	<i>rps8-rpl14</i>
37	p1	intron	LSC	T	11	88315	88325	<i>rpl16</i>
38	p1	intron	LSC	C	10	91506	91515	<i>rpl2</i>
39	p2	spacer	IRB	AT	12	94055	94066	<i>trnT(UGU)</i>
40	p1	spacer	IRB	A	12	101175	101186	<i>trnI(GAU)-rrn16</i>
41	p4	spacer	IRB	TCTT	12	103479	103490	<i>rrn16-rps12</i>
42	p1	spacer	IRB	A	14	104578	104591	<i>rrn16-rps12</i>
43	p1	spacer	IRB	T	10	106780	106789	<i>rps7-psbA</i>
44	p3	spacer	SSC	AAT	12	119836	119847	<i>rpl32-trnP(GGG)</i>
45	p2	spacer	SSC	TA	18	119850	119867	<i>rpl32-trnP(GGG)</i>
46	p3	spacer	SSC	TAT	12	119869	119880	<i>rpl32-trnP(GGG)</i>
47	p4	spacer	SSC	GAAA	12	120219	120230	<i>trnP(GGG)-trnL(UAG)</i>
48	p1	spacer	SSC	T	12	120461	120472	<i>trnL(UAG)-ccsA</i>
49	p1	spacer	SSC	A	11	121731	121741	<i>ccsA-ndhD</i>
50	p1	spacer	SSC	A	11	123596	123606	<i>ndhD-psaC</i>
51	p4	spacer	SSC	AATT	12	123605	123616	<i>ndhD-psaC</i>
52	p1	spacer	SSC	T	10	123615	123624	<i>ndhD-psaC</i>
53	p1	spacer	SSC	G	15	124153	124167	<i>psaC-ndhE</i>
54	p1	spacer	SSC	A	24	125982	125992	<i>ndhI-ndhA</i>
55	p2	spacer	SSC	AT	14	125992	126005	<i>ndhI-ndhA</i>
56	p1	spacer	SSC	A	13	126041	126053	<i>ndhI-ndhA</i>
57	p2	intron	SSC	TA	14	127030	127043	<i>ndhA</i>
58	p1	spacer	SSC	A	11	129846	129856	<i>rps15-ycf1</i>
59	p1	spacer	SSC	C	14	135178	135191	<i>ycf1-chlN</i>
60	p4	spacer	SSC	AAAT	16	135410	135425	<i>ycf1-chlN</i>
61	p1	spacer	IRA	A	10	147274	147283	<i>psbA-rps7</i>
62	p1	spacer	IRA	T	14	149472	149485	<i>rps12-rrn16</i>
63	p4	spacer	IRA	AAAG	12	150572	150583	<i>rps12-rrn16</i>
64	p1	spacer	IRA	T	12	152877	152888	<i>rrn16-trnI(GAU)</i>
65	p2	spacer	IRA	AT	12	159997	160008	<i>trnT(UGU)</i>

Appendix table 5 List of simple sequence repeats (SSRs) in the chloroplast genomes of *Alsophila costularis*

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
1	p1	intron	LSC	T	11	367	374	<i>ndhB</i>
2	p1	spacer	LSC	A	12	3443	3454	<i>matK-rps16</i>
3	p1	spacer	LSC	T	10	3538	3547	<i>matK-rps16</i>
4	p1	intron	LSC	T	11	4288	4298	<i>rps16</i>
5	p1	spacer	LSC	G	15	6798	6812	<i>chlB-trnQ(UUG)</i>
6	p1	spacer	LSC	A	11	7517	7527	<i>psbK-psbI</i>
7	p2	spacer	LSC	TA	12	7529	7540	<i>psbK-psbI</i>
8	p1	spacer	LSC	C	13	8126	8138	<i>trnS(GCU)-psaM</i>
9	p2	spacer	LSC	AT	16	8229	8244	<i>trnS(GCU)-psaM</i>
10	p1	spacer	LSC	T	10	8244	8253	<i>trnS(GCU)-psaM</i>
11	p1	spacer	LSC	G	11	8797	8807	<i>ycf12-trnG(UCC)</i>
12	p1	intron	LSC	C	14	9681	9694	<i>trnG(UCC)</i>
13	p1	spacer	LSC	C	19	14047	14065	<i>atpH-atpI</i>
14	p1	spacer	LSC	A	11	14078	14088	<i>atpH-atpI</i>
15	p1	spacer	LSC	T	13	15006	15018	<i>atpI-rps2</i>
16	p1	intron	LSC	T	13	22288	22300	<i>rpoC1</i>
17	p3	intron	LSC	AAG	12	22479	22490	<i>rpoC1</i>
18	p1	spacer	LSC	A	11	23235	23245	<i>rpoC1-rpoB</i>
19	p1	spacer	LSC	T	10	27599	27608	<i>rpoB-trnD(GUC)</i>
20	p1	spacer	LSC	A	12	28205	28216	<i>trnE(UUC)-psbM</i>
21	p1	spacer	LSC	A	12	29369	29380	<i>ycf66</i>
22	p2	spacer	LSC	AT	18	30361	30378	<i>petN-trnC(GCA)</i>
23	p1	spacer	LSC	G	12	31701	31712	<i>trnC(GCA)-trnG(GCC)</i>
24	p1	spacer	LSC	A	13	31782	31794	<i>trnC(GCA)-trnG(GCC)</i>
25	p1	spacer	LSC	A	12	32996	33007	<i>trnS(UGA)-psbC</i>
26	p1	spacer	LSC	T	13	33067	33079	<i>trnS(UGA)-psbC</i>
27	p4	spacer	LSC	ATCT	16	33600	33615	<i>trnS(UGA)-psbC</i>
28	p4	spacer	LSC	ATAG	12	33618	33629	<i>trnS(UGA)-psbC</i>
29	p1	spacer	LSC	T	11	36726	36736	<i>psbD-trnT(GGU)</i>
30	p1	spacer	LSC	A	10	36854	36863	<i>psbD-trnT(GGU)</i>
31	p1	spacer	LSC	T	19	37272	37290	<i>psbD-trnT(GGU)</i>
32	p1	spacer	LSC	A	12	37670	37681	<i>psbD-trnT(GGU)</i>
33	p1	intron	LSC	T	11	44231	44241	<i>ycf3</i>
34	p1	intron	LSC	C	11	45328	45338	<i>ycf3</i>
35	p1	spacer	LSC	T	15	45725	45739	<i>ycf3-trnS(GGA)</i>
36	p1	spacer	LSC	A	10	45781	45790	<i>ycf3-trnS(GGA)</i>
37	p1	spacer	LSC	T	12	45851	45862	<i>ycf3-trnS(GGA)</i>
38	p1	spacer	LSC	T	11	46321	46331	<i>trnS(GGA)-rps4</i>
39	p1	spacer	LSC	G	10	48024	48033	<i>rps4-trnL(CAA)</i>
40	p1	intron	LSC	A	14	48342	48355	<i>trnL(CAA)</i>
41	p1	spacer	LSC	T	14	48944	48957	<i>trnL(CAA)-trnF(GAA)</i>
42	p1	spacer	LSC	T	10	51286	51295	<i>ndhC-trnV(UAC)</i>
43	p1	spacer	LSC	A	11	51302	51312	<i>ndhC-trnV(UAC)</i>
44	p1	spacer	LSC	A	10	52147	52156	<i>trnV(UAC)-trnM(CAU)</i>
45	p2	spacer	LSC	AT	14	52242	52255	<i>trnV(UAC)-trnM(CAU)</i>
46	p1	spacer	LSC	T	13	54712	54724	<i>atpB-rbcL</i>
47	p1	spacer	LSC	T	12	56557	56568	<i>rbcL-trnR(UCG)</i>
48	p1	spacer	LSC	T	19	56978	56996	<i>trnR(UCG)-accD</i>

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
49	p4	spacer	LSC	TATT	12	58025	58036	<i>accD-psaI</i>
50	p1	spacer	LSC	A	10	58085	58094	<i>accD-psaI</i>
51	p1	spacer	LSC	T	10	58508	58517	<i>accD-psaI</i>
52	p1	spacer	LSC	C	11	62532	62542	<i>petA-psbJ</i>
53	p1	spacer	LSC	T	13	63161	63173	<i>petA-psbJ</i>
54	p1	spacer	LSC	T	11	64725	64735	<i>psbE-petL</i>
55	p1	spacer	LSC	T	10	65541	65550	<i>petG-trnW(CCA)</i>
56	p1	spacer	LSC	T	10	65940	65949	<i>trnW(CCA)-trnP(UGG)</i>
57	p1	spacer	LSC	T	18	67335	67352	<i>rps18-rpl20</i>
58	p1	spacer	LSC	T	12	68310	68321	<i>rpl20-rps12</i>
59	p1	spacer	LSC	T	11	68513	68523	<i>rpl20-rps12</i>
60	p1	spacer	LSC	T	11	68770	68780	<i>rpl20-rps12</i>
61	p4	spacer	LSC	TATC	12	69226	69237	<i>rps12-clpP</i>
62	p4	spacer	LSC	GATA	12	69244	69255	<i>rps12-clpP</i>
63	p1	intron	LSC	C	42	69761	69802	<i>clpP</i>
64	p1	intron	LSC	C	10	70520	70529	<i>clpP</i>
65	p2	intron	LSC	AT	24	70655	70678	<i>clpP</i>
66	p1	spacer	LSC	A	11	71659	71669	<i>clpP-psbB</i>
67	p1	intron	LSC	T	16	74446	74461	<i>petB</i>
68	p1	intron	LSC	T	10	76585	76594	<i>petD</i>
69	p1	spacer	LSC	T	13	80192	80204	<i>rps8-rpl14</i>
70	p2	spacer	LSC	AT	18	80224	80241	<i>rps8-rpl14</i>
71	p1	spacer	LSC	A	14	80308	80321	<i>rps8-rpl14</i>
72	p2	spacer	LSC	TA	12	80322	80333	<i>rps8-rpl14</i>
73	p1	spacer	LSC	T	12	80342	80353	<i>rps8-rpl14</i>
74	p1	intron	LSC	T	12	81928	81939	<i>rpl16</i>
75	p4	intron	LSC	CCTC	12	82094	82105	<i>rpl16</i>
76	p1	intron	LSC	T	13	82267	82279	<i>rpl16</i>
77	p1	spacer	LSC	T	10	82493	82502	<i>rpl16-rps3</i>
78	p1	intron	LSC	A	14	84696	84709	<i>rpl2</i>
79	p1	spacer	LSC	A	12	85684	85695	<i>rpl2-rpl23</i>
80	p1	spacer	LSC	T	13	85997	86009	<i>rpl23-trnI(CAU)</i>
81	p1	spacer	LSC	T	11	86139	86149	<i>rpl23-trnI(CAU)</i>
82	p2	spacer	IRB	AT	12	88021	88032	<i>trnT(UGU)</i>
83	p2	spacer	IRB	AT	12	88476	88487	<i>trnT(UGU)-trnR(ACG)</i>
84	p1	spacer	IRB	T	13	89923	89935	<i>trnR(ACG)-rrn5</i>
85	p1	spacer	IRB	C	13	97451	97463	<i>rrn16-rps12</i>
86	p4	spacer	IRB	TCTT	12	97984	97995	<i>rrn16-rps12</i>
87	p1	spacer	IRB	T	11	99097	99107	<i>rrn16-rps12</i>
88	p1	spacer	IRB	A	12	103055	103066	<i>trnH(GUG)-ycf2</i>
89	p4	CDS	IRB	AGAA	12	108231	108242	<i>ycf2</i>
90	p1	spacer	IRB	A	17	109641	109657	<i>ycf2-trnN(GUU)</i>
91	p1	spacer	IRB	G	17	110427	110443	<i>trnN(GUU)-ndhF</i>
92	p1	spacer	SSC	T	10	113600	113609	<i>rpl21-rpl32</i>
93	p1	spacer	SSC	A	13	113713	113725	<i>rpl21-rpl32</i>
94	p2	spacer	SSC	AT	16	113725	113740	<i>rpl21-rpl32</i>
95	p1	spacer	SSC	A	12	114078	114089	<i>rpl32-trnP(GGG)</i>
96	p1	spacer	SSC	G	10	114333	114342	<i>rpl32-trnP(GGG)</i>
97	p1	spacer	SSC	T	14	114400	114413	<i>rpl32-trnP(GGG)</i>
98	p1	spacer	SSC	A	13	114740	114752	<i>trnP(GGG)-trnL(UAG)</i>

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
99	p1	spacer	SSC	A	11	114900	114910	<i>trnL(UAG)-ccsA</i>
100	p1	spacer	SSC	T	12	114979	114990	<i>trnL(UAG)-ccsA</i>
101	p1	spacer	SSC	A	11	118132	118142	<i>ndhD-psaC</i>
102	p1	spacer	SSC	T	12	118147	118158	<i>ndhD-psaC</i>
103	p1	spacer	SSC	G	13	118694	118706	<i>psaC-ndhE</i>
104	p2	spacer	SSC	TA	14	120509	120522	<i>ndhI-ndhA</i>
105	p1	spacer	SSC	A	13	120526	120538	<i>ndhI-ndhA</i>
106	p2	spacer	SSC	AT	20	120538	120557	<i>ndhI-ndhA</i>
107	p1	spacer	SSC	A	10	120602	120611	<i>ndhI-ndhA</i>
108	p1	intron	SSC	T	10	121522	121531	<i>ndhA</i>
109	p1	spacer	SSC	A	14	124314	124327	<i>rps15-ycf1</i>
110	p4	CDS	SSC	ACCA	12	129098	129109	<i>ycf1</i>
111	p1	spacer	SSC	C	13	129639	129651	<i>ycf1-chlN</i>
112	p1	spacer	IRA	C	17	132571	132587	<i>chlL-trnN(GUU)</i>
113	p1	spacer	IRA	T	17	133357	133373	<i>trnN(GUU)-ycf2</i>
114	p4	CDS	IRA	TTTC	12	134771	134782	<i>ycf2</i>
115	p1	spacer	IRA	T	12	139948	139959	<i>ycf2-trnH(GUG)</i>
116	p1	spacer	IRA	A	11	143907	143917	<i>rps12-rrn16</i>
117	p4	spacer	IRA	AAAG	12	145018	145029	<i>rps12-rrn16</i>
118	p1	spacer	IRA	G	13	145551	145563	<i>rps12-rrn16</i>
119	p1	spacer	IRA	A	13	153079	153091	<i>rrn5-trnR(ACG)</i>
120	p2	spacer	IRA	AT	12	154527	154538	<i>trnR(ACG)-trnT(UGU)</i>
121	p2	spacer	IRA	AT	12	154982	154993	<i>trnT(UGU)</i>

Appendix table 6 List of simple sequence repeats (SSRs) in the chloroplast genomes of *Alsophila spinulosa*

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
1	p1	intron	LSC	T	11	364	374	<i>ndhB</i>
2	p1	spacer	LSC	A	13	3443	3455	<i>matK-rps16</i>
3	p1	spacer	LSC	T	10	3539	3548	<i>matK-rps16</i>
4	p1	intron	LSC	T	11	4289	4299	<i>rps16</i>
5	p1	spacer	LSC	G	14	6799	6812	<i>chlB-trnQ(UUG)</i>
6	p1	spacer	LSC	A	24	7517	7527	<i>psbK-psbI</i>
7	p2	spacer	LSC	TA	12	7529	7540	<i>psbK-psbI</i>
8	p1	spacer	LSC	C	12	8126	8137	<i>trnS(GCU)-psaM</i>
9	p2	spacer	LSC	AT	16	8228	8243	<i>trnS(GCU)-psaM</i>
10	p1	spacer	LSC	T	10	8243	8252	<i>trnS(GCU)-psaM</i>
11	p1	spacer	LSC	G	11	8796	8806	<i>ycf12-trnG(UCC)</i>
12	p1	intron	LSC	C	13	9680	9692	<i>trnG(UCC)</i>
13	p1	spacer	LSC	C	10	14045	14054	<i>atpH-atpI</i>
14	p1	spacer	LSC	A	11	14072	14082	<i>atpH-atpI</i>
15	p1	spacer	LSC	T	12	15000	15011	<i>atpI-rps2</i>
16	p1	intron	LSC	T	13	22281	22293	<i>rpoC1</i>
17	p3	intron	LSC	AAG	12	22472	22483	<i>rpoC1</i>
18	p1	spacer	LSC	A	11	23228	23238	<i>rpoC1-rpoB</i>
19	p1	spacer	LSC	T	10	27592	27601	<i>rpoB-trnD(GUC)</i>
20	p1	spacer	LSC	A	12	28198	28209	<i>trnE(UUC)-psbM</i>
21	p1	spacer	LSC	A	12	29362	29373	<i>ycf66</i>
22	p2	spacer	LSC	AT	18	30354	30371	<i>petN-trnC(GCA)</i>

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
23	p1	spacer	LSC	G	12	31694	31705	<i>trnC(GCA)-trnG(GCC)</i>
24	p1	spacer	LSC	A	13	31775	31787	<i>trnC(GCA)-trnG(GCC)</i>
25	p1	spacer	LSC	A	12	32989	33000	<i>trnS(UGA)-psbC</i>
26	p1	spacer	LSC	T	13	33060	33072	<i>trnS(UGA)-psbC</i>
27	p4	spacer	LSC	ATCT	16	33593	33608	<i>trnS(UGA)-psbC</i>
28	p4	spacer	LSC	ATAG	12	33611	33622	<i>trnS(UGA)-psbC</i>
29	p1	spacer	LSC	T	11	36719	36729	<i>psbD-trnT(GGU)</i>
30	p1	spacer	LSC	A	10	36847	36856	<i>psbD-trnT(GGU)</i>
31	p1	spacer	LSC	T	19	37265	37283	<i>psbD-trnT(GGU)</i>
32	p1	spacer	LSC	A	11	37663	37673	<i>psbD-trnT(GGU)</i>
33	p1	intron	LSC	T	12	44223	44234	<i>ycf3</i>
34	p1	intron	LSC	C	11	45321	45331	<i>ycf3</i>
35	p1	spacer	LSC	T	15	45718	45732	<i>ycf3-trnS(GGA)</i>
36	p1	spacer	LSC	A	10	45774	45783	<i>ycf3-trnS(GGA)</i>
37	p1	spacer	LSC	T	12	45844	45855	<i>ycf3-trnS(GGA)</i>
38	p1	spacer	LSC	T	11	46314	46324	<i>trnS(GGA)-rps4</i>
39	p1	spacer	LSC	G	10	48017	48026	<i>rps4-trnL(CAA)</i>
40	p1	intron	LSC	A	14	48335	48348	<i>trnL(CAA)</i>
41	p1	spacer	LSC	T	13	48937	48949	<i>trnL(CAA)-trnF(GAA)</i>
42	p1	spacer	LSC	T	10	51278	51287	<i>ndhC-trnV(UAC)</i>
43	p1	spacer	LSC	A	11	51294	51304	<i>ndhC-trnV(UAC)</i>
44	p1	spacer	LSC	A	10	52139	52148	<i>trnV(UAC)-trnM(CAU)</i>
45	p2	spacer	LSC	AT	14	52234	52247	<i>trnV(UAC)-trnM(CAU)</i>
46	p1	spacer	LSC	T	13	54704	54716	<i>atpB-rbcL</i>
47	p1	spacer	LSC	T	12	56549	56560	<i>rbcL-trnR(UCG)</i>
48	p1	spacer	LSC	T	19	56970	56988	<i>trnR(UCG)-accD</i>
49	p4	spacer	LSC	TATT	12	58017	58028	<i>accD-psaI</i>
50	p1	spacer	LSC	A	10	58077	58086	<i>accD-psaI</i>
51	p1	spacer	LSC	T	11	58500	58510	<i>accD-psaI</i>
52	p1	spacer	LSC	C	11	62525	62535	<i>petA-psbJ</i>
53	p1	spacer	LSC	T	13	63154	63166	<i>petA-psbJ</i>
54	p1	spacer	LSC	T	11	64718	64728	<i>psbE-petL</i>
55	p1	spacer	LSC	T	10	65534	65543	<i>petG-trnW(CCA)</i>
56	p1	spacer	LSC	T	10	65933	65942	<i>trnW(CCA)-trnP(UGG)</i>
57	p1	spacer	LSC	C	10	66640	66649	<i>psaJ-rpl33</i>
58	p1	spacer	LSC	T	17	67329	67345	<i>rps18-rpl20</i>
59	p1	spacer	LSC	T	12	68303	68314	<i>rpl20-rps12</i>
60	p1	spacer	LSC	T	11	68506	68516	<i>rpl20-rps12</i>
61	p1	spacer	LSC	T	11	68763	68773	<i>rpl20-rps12</i>
62	p4	spacer	LSC	TATC	12	69219	69230	<i>rps12-clpP</i>
63	p4	spacer	LSC	GATA	12	69237	69248	<i>rps12-clpP</i>
64	p1	intron	LSC	C	17	69754	69770	<i>clpP</i>
65	p1	intron	LSC	C	11	70488	70498	<i>clpP</i>
66	p2	intron	LSC	AT	24	70624	70647	<i>clpP</i>
67	p1	spacer	LSC	A	11	71628	71638	<i>clpP-psbB</i>
68	p1	intron	LSC	T	16	74415	74430	<i>petB</i>
69	p1	intron	LSC	T	10	76554	76563	<i>petD</i>
70	p1	spacer	LSC	T	13	80161	80173	<i>rps8-rpl14</i>
71	p2	spacer	LSC	AT	18	80193	80210	<i>rps8-rpl14</i>
72	p1	spacer	LSC	A	14	80277	80290	<i>rps8-rpl14</i>

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
73	p2	spacer	LSC	TA	12	80291	80302	<i>rps8-rpl14</i>
74	p1	spacer	LSC	T	12	80311	80322	<i>rps8-rpl14</i>
75	p1	intron	LSC	T	12	81897	81908	<i>rpl16</i>
76	p4	intron	LSC	CCTC	12	82063	82074	<i>rpl16</i>
77	p1	intron	LSC	T	13	82236	82248	<i>rpl16</i>
78	p1	spacer	LSC	T	10	82462	82471	<i>rpl16-rps3</i>
79	p1	intron	LSC	A	15	84665	84679	<i>rpl2</i>
80	p1	spacer	LSC	A	12	85654	85665	<i>rpl2-rpl23</i>
81	p1	spacer	LSC	T	13	85967	85979	<i>rpl23-trnI(CAU)</i>
82	p1	spacer	LSC	T	11	86109	86119	<i>rpl23-trnI(CAU)</i>
83	p2	spacer	IRB	AT	12	87991	88002	<i>trnT(UGU)</i>
84	p2	spacer	IRB	AT	12	88446	88457	<i>trnT(UGU)-trnR(ACG)</i>
85	p1	spacer	IRB	T	13	89893	89905	<i>trnR(ACG)-rrn5</i>
86	p1	spacer	IRB	C	11	97421	97431	<i>rrn16-rps12</i>
87	p4	spacer	IRB	TCTT	12	97952	97963	<i>rrn16-rps12</i>
88	p1	spacer	IRB	T	11	99065	99075	<i>rrn16-rps12</i>
89	p1	spacer	IRB	A	12	103023	103034	<i>trnH(GUG)-ycf2</i>
90	p4	CDS	IRB	AGAA	12	108199	108210	<i>ycf2</i>
91	p1	spacer	IRB	A	23	109609	109631	<i>ycf2-trnN(GUU)</i>
92	p1	spacer	IRB	G	15	110408	110422	<i>trnN(GUU)-ndhF</i>
93	p1	spacer	SSC	T	10	113579	113588	<i>rpl21-rpl32</i>
94	p1	spacer	SSC	A	13	113692	113704	<i>rpl21-rpl32</i>
95	p2	spacer	SSC	AT	16	113704	113719	<i>rpl21-rpl32</i>
96	p1	spacer	SSC	A	11	114057	114067	<i>rpl32-trnP(GGG)</i>
97	p1	spacer	SSC	G	10	114311	114320	<i>rpl32-trnP(GGG)</i>
98	p1	spacer	SSC	T	13	114378	114390	<i>rpl32-trnP(GGG)</i>
99	p1	spacer	SSC	A	13	114717	114729	<i>trnP(GGG)-trnL(UAG)</i>
100	p1	spacer	SSC	A	11	114877	114887	<i>trnL(UAG)-ccsA</i>
101	p1	spacer	SSC	T	12	114956	114967	<i>trnL(UAG)-ccsA</i>
102	p1	spacer	SSC	A	11	118109	118119	<i>ndhD-psaC</i>
103	p1	spacer	SSC	T	12	118124	118135	<i>ndhD-psaC</i>
104	p1	spacer	SSC	G	13	118671	118683	<i>psaC-ndhE</i>
105	p2	spacer	SSC	TA	14	120486	120499	<i>ndhI-ndhA</i>
106	p1	spacer	SSC	A	13	120503	120515	<i>ndhI-ndhA</i>
107	p2	spacer	SSC	AT	20	120515	120534	<i>ndhI-ndhA</i>
108	p1	spacer	SSC	A	10	120579	120588	<i>ndhI-ndhA</i>
109	p1	intron	SSC	T	10	121499	121508	<i>ndhA</i>
110	p1	spacer	SSC	A	14	124291	124304	<i>rps15-ycf1</i>
111	p4	CDS	SSC	ACCA	12	129075	129086	<i>ycf1</i>
112	p1	spacer	SSC	C	13	129616	129628	<i>ycf1-chlN</i>
113	p1	spacer	IRA	C	15	132548	132562	<i>chlL-trnN(GUU)</i>
114	p1	spacer	IRA	T	23	133339	133361	<i>trnN(GUU)-ycf2</i>
115	p4	CDS	IRA	TTTC	12	134759	134770	<i>ycf2</i>
116	p1	spacer	IRA	T	12	139936	139947	<i>ycf2-trnH(GUG)</i>
117	p1	spacer	IRA	A	11	143895	143905	<i>rps12-rrn16</i>
118	p4	spacer	IRA	AAAG	12	145006	145017	<i>rps12-rrn16</i>
119	p1	spacer	IRA	G	11	145539	145549	<i>rps12-rrn16</i>
120	p1	spacer	IRA	A	13	153065	153077	<i>rrn5-trnR(ACG)</i>
121	p2	spacer	IRA	AT	12	154513	154524	<i>trnR(ACG)-trnT(UGU)</i>

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
122	p2	spacer	IRA	AT	12	154968	154979	<i>trnT(UGU)</i>

Appendix table 7 List of simple sequence repeats (SSRs) in the chloroplast genomes of *Sphaeropteris brunonianiana*

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
1	p1	spacer	LSC	A	15	3517	3531	<i>matK-rps16</i>
2	p1	spacer	LSC	T	10	3614	3623	<i>matK-rps16</i>
3	p1	intron	LSC	T	10	4364	4373	<i>rps16</i>
4	p1	intron	LSC	A	11	4861	4871	<i>rps16</i>
5	p1	spacer	LSC	G	10	6893	6902	<i>chlB-trnQ(UUG)</i>
6	p1	spacer	LSC	A	12	7068	7619	<i>psbK-psbI</i>
7	p2	spacer	LSC	TA	12	7621	7632	<i>psbK-psbI</i>
8	p1	spacer	LSC	C	17	8230	8246	<i>trnS(GCU)-psaM</i>
9	p1	spacer	LSC	A	11	8332	8342	<i>trnS(GCU)-psaM</i>
10	p1	spacer	LSC	T	12	8359	8370	<i>trnS(GCU)-psaM</i>
11	p1	intron	LSC	T	12	9637	9648	<i>trnG(UCC)</i>
12	p2	intron	LSC	TC	14	12639	12652	<i>atpF</i>
13	p1	spacer	LSC	C	16	13302	13317	<i>atpF-atpH</i>
14	p1	spacer	LSC	T	13	15171	15183	<i>atpI-rps2</i>
15	p1	spacer	LSC	A	10	23386	23395	<i>rpoC1-rpoB</i>
16	p1	spacer	LSC	T	12	27644	27655	<i>rpoB-trnD(GUC)</i>
17	p1	spacer	LSC	A	11	28256	28266	<i>trnE(UUC)-psbM</i>
18	p1	spacer	LSC	T	22	28291	28312	<i>trnE(UUC)-psbM</i>
19	p1	spacer	LSC	A	14	29431	29444	<i>ycf66</i>
20	p1	spacer	LSC	G	14	29522	29535	<i>ycf66</i>
21	p1	spacer	LSC	T	13	30206	30218	<i>petN-trnC(GCA)</i>
22	p1	spacer	LSC	T	11	30804	30814	<i>trnC(GCA)-trnG(GCC)</i>
23	p4	spacer	LSC	TATC	16	33672	33687	<i>trnS(UGA)-psbC</i>
24	p4	spacer	LSC	ATAG	16	33691	33706	<i>trnS(UGA)-psbC</i>
25	p1	spacer	LSC	T	11	36882	36892	<i>psbD-trnT(GGU)</i>
26	p1	spacer	LSC	A	10	36920	36929	<i>psbD-trnT(GGU)</i>
27	p1	spacer	LSC	T	19	37334	37352	<i>psbD-trnT(GGU)</i>
28	p1	spacer	LSC	T	21	37588	37608	<i>psbD-trnT(GGU)</i>
29	p4	spacer	LSC	ATAG	12	38356	38367	<i>trnfM(CAU)-rps14</i>
30	p1	spacer	LSC	T	10	38371	38380	<i>trnfM(CAU)-rps14</i>
31	p1	intron	LSC	T	21	44159	44179	<i>ycf3</i>
32	p1	spacer	LSC	T	23	45660	45682	<i>ycf3-trnS(GGA)</i>
33	p1	spacer	LSC	T	18	45968	45985	<i>ycf3-trnS(GGA)</i>
34	p1	spacer	LSC	T	13	46269	46281	<i>trnS(GGA)-rps4</i>
35	p1	intron	LSC	A	11	48282	48292	<i>trnL(CAA)</i>
36	p2	spacer	LSC	TA	12	48647	48658	<i>trnL(CAA)-trnF(GAA)</i>
37	p2	spacer	LSC	TA	24	48661	48684	<i>trnL(CAA)-trnF(GAA)</i>
38	p1	spacer	LSC	T	12	51179	51190	<i>ndhC-trnV(UAC)</i>
39	p1	intron	LSC	T	10	51514	51523	<i>trnV(UAC)</i>
40	p1	spacer	LSC	A	16	52031	52046	<i>trnV(UAC)-trnM(CAU)</i>
41	p1	spacer	LSC	T	10	54498	54507	<i>atpB-rbcL</i>
42	p1	spacer	LSC	T	12	56441	56452	<i>rbcL-trnR(UCG)</i>
43	p1	spacer	LSC	T	14	56859	56872	<i>trnR(UCG)-accD</i>

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
44	p1	spacer	LSC	T	10	58386	58395	<i>accD-psaI</i>
45	p2	spacer	LSC	AT	16	62683	62698	<i>petA-psbJ</i>
46	p1	spacer	LSC	T	13	63021	63033	<i>petA-psbJ</i>
47	p1	spacer	LSC	T	16	64669	64684	<i>psbE-petL</i>
48	p1	spacer	LSC	A	12	65041	65052	<i>petL-petG</i>
49	p1	spacer	LSC	C	10	66553	66562	<i>psaJ-rpl33</i>
50	p1	spacer	LSC	A	13	67731	67743	<i>rpl20-rps12</i>
51	p1	spacer	LSC	T	13	67974	67986	<i>rpl20-rps12</i>
52	p1	spacer	LSC	A	15	68134	68148	<i>rpl20-rps12</i>
53	p1	spacer	LSC	T	11	68406	68416	<i>rpl20-rps12</i>
54	p4	spacer	LSC	TATC	12	69106	69117	<i>rps12-clpP</i>
55	p2	intron	LSC	TA	12	70495	70506	<i>clpP</i>
56	p1	intron	LSC	C	14	70536	70549	<i>clpP</i>
57	p1	spacer	LSC	A	12	71504	71515	<i>clpP-psbB</i>
58	p1	spacer	LSC	T	11	71597	71607	<i>clpP-psbB</i>
59	p1	spacer	LSC	A	18	73387	73404	<i>psbB-psbT</i>
60	p1	intron	LSC	T	10	74735	74744	<i>petB</i>
61	p1	spacer	LSC	T	10	77271	77280	<i>petD-ropA</i>
62	p1	spacer	LSC	T	11	77328	77338	<i>petD-ropA</i>
63	p1	spacer	LSC	T	14	80050	80063	<i>rps8-rpl14</i>
64	p2	spacer	LSC	AT	14	80083	80096	<i>rps8-rpl14</i>
65	p2	spacer	LSC	AT	18	80177	80194	<i>rps8-rpl14</i>
66	p1	spacer	LSC	A	13	81030	81042	<i>rpl14-rpl16</i>
67	p1	intron	LSC	T	11	81765	81775	<i>rpl16</i>
68	p1	intron	LSC	T	11	82107	82117	<i>rpl16</i>
69	p2	spacer	LSC	AT	14	82324	82337	<i>rpl16-rps3</i>
70	p1	intron	LSC	A	10	84458	84467	<i>rpl2</i>
71	p1	intron	LSC	A	10	84570	84549	<i>rpl2</i>
72	p2	intron	LSC	AT	12	84561	84572	<i>rpl2</i>
73	p1	intron	LSC	T	14	84852	84865	<i>rpl2</i>
74	p1	spacer	LSC	T	13	85845	85857	<i>rpl23-trnI(CAU)</i>
75	p1	spacer	LSC	T	13	85991	86003	<i>rpl23-trnI(CAU)</i>
76	p4	gene	IRA	CTAC	12	91186	91197	<i>rrn23</i>
77	p2	intron	IRA	AG	12	94186	94197	<i>trnI(CAU)</i>
78	p4	spacer	IRA	TCTT	12	97474	97485	<i>rrn16-rps12</i>
79	p1	spacer	IRA	T	14	98567	98580	<i>rrn16-rps12</i>
80	p1	spacer	IRA	A	12	102544	102555	<i>trnH(GUG)-ycf2</i>
81	p4	CDS	IRA	AGAA	12	107744	107755	<i>ycf2</i>
82	p1	spacer	IRA	A	15	109158	109172	<i>ycf2-trnN(GUU)</i>
83	p1	spacer	SSC	T	11	110207	110217	<i>trnN(GUU)-ndhF</i>
84	p1	spacer	SSC	A	14	113545	113558	<i>rpl32-trnP(GGG)</i>
85	p1	spacer	SSC	A	11	113825	113835	<i>rpl32-trnP(GGG)</i>
86	p1	spacer	SSC	T	15	113869	113883	<i>rpl32-trnP(GGG)</i>
87	p1	spacer	SSC	A	13	114374	114386	<i>trnL(UAG)-ccsA</i>
88	p2	spacer	SSC	AT	20	114464	114483	<i>trnL(UAG)-ccsA</i>
89	p1	spacer	SSC	A	13	115715	115727	<i>ccsA-ndhD</i>
90	p1	spacer	SSC	A	14	115972	115985	<i>ccsA-ndhD</i>
91	p1	spacer	SSC	A	14	117633	117646	<i>ndhD-psaC</i>
92	p2	spacer	SSC	CT	12	119317	119328	<i>ndhG-ndhI</i>
93	p2	intron	SSC	AT	16	121036	121051	<i>ndhA</i>

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
94	p1	intron	SSC	T	11	121544	121554	<i>ndhA</i>
95	p1	intron	SSC	A	11	123840	123850	<i>rps15-ycf1</i>
96	p1	spacer	IRB	T	15	133684	133698	<i>trnN(GUU)-ycf2</i>
97	p4	CDS	IRB	TTTC	12	135100	135111	<i>ycf2</i>
98	p1	spacer	IRB	T	12	140301	140312	<i>ycf2-trnH(GUG)</i>
99	p1	spacer	IRB	A	14	144276	144289	<i>rps12-rrn16</i>
100	p4	spacer	IRB	AAAG	12	145370	145381	<i>rps12-rrn16</i>
101	p2	intron	IRB	TC	12	148658	148669	<i>trnI(GUA)</i>
102	p4	gene	IRB	AGGT	12	151657	151668	<i>rrn23</i>

Appendix table 8 List of simple sequence repeats (SSRs) in the chloroplast genomes of *Sphaeropteris lepifera*

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
1	p1	spacer	LSC	A	13	3514	3526	<i>matK-rps16</i>
2	p1	spacer	LSC	T	10	3613	3622	<i>matK-rps16</i>
3	p2	spacer	LSC	TA	16	7615	7630	<i>psbK-psbI</i>
4	p1	spacer	LSC	C	14	8229	8242	<i>trnS(GCU)-psaM</i>
5	p1	spacer	LSC	A	22	8332	8353	<i>trnS(GCU)-psaM</i>
6	p1	spacer	LSC	T	11	8370	8380	<i>trnS(GCU)-psaM</i>
7	p1	intron	LSC	A	10	9507	9516	<i>ycf12</i>
8	p1	intron	LSC	T	12	9648	9659	<i>ycf12</i>
9	p1	spacer	LSC	C	38	13308	13345	<i>atpF-atpH</i>
10	p1	spacer	LSC	T	12	15193	15204	<i>atpI-rps2</i>
11	p1	intron	LSC	C	10	22421	22430	<i>rpoC1</i>
12	p1	spacer	LSC	A	12	23409	23420	<i>rpoC1-rpoB</i>
13	p1	spacer	LSC	T	14	27774	27787	<i>rpoB-trnD(GUC)</i>
14	p1	spacer	LSC	A	14	28387	28400	<i>trnE(UUC)-psbM</i>
15	p1	spacer	LSC	T	21	28425	28445	<i>trnE(UUC)-psbM</i>
16	p1	spacer	LSC	A	12	29887	29898	<i>ycf66</i>
17	p1	spacer	LSC	T	10	30334	30343	<i>petN-trnC(GCA)</i>
18	p1	spacer	LSC	A	15	30525	30539	<i>petN-trnC(GCA)</i>
19	p1	spacer	LSC	T	10	30938	30947	<i>trnC(GCA)-trnG(GCC)</i>
20	p1	spacer	LSC	T	12	32361	32372	<i>trnG(GCC)-psbZ</i>
21	p1	spacer	LSC	T	10	33284	33293	<i>trnS(UGA)-psbC</i>
22	p4	spacer	LSC	TATC	16	33813	33828	<i>trnS(UGA)-psbC</i>
23	p4	spacer	LSC	ATAG	12	33832	33843	<i>trnS(UGA)-psbC</i>
24	p1	spacer	LSC	T	16	37027	37042	<i>psbD-trnT(GGU)</i>
25	p1	spacer	LSC	A	13	37070	37082	<i>psbD-trnT(GGU)</i>
26	p1	spacer	LSC	T	19	37491	37509	<i>psbD-trnT(GGU)</i>
27	p1	spacer	LSC	T	12	37753	37764	<i>psbD-trnT(GGU)</i>
28	p2	spacer	LSC	TA	16	38517	38532	<i>trnfM-rps14</i>
29	p1	spacer	LSC	T	10	38541	38550	<i>trnfM-rps14</i>
30	p1	intron	LSC	T	13	44335	44347	<i>ycf3</i>
31	p1	intron	LSC	C	10	45444	45453	<i>ycf3</i>
32	p1	spacer	LSC	T	12	45838	45849	<i>ycf3-trnS(GGA)</i>
33	p1	spacer	LSC	T	12	46134	46145	<i>ycf3-trnS(GGA)</i>
34	p1	spacer	LSC	T	11	46434	46444	<i>trnS(GGA)-rps4</i>
35	p1	intron	LSC	A	10	48446	48455	<i>trnL(CAA)</i>
36	p2	spacer	LSC	TA	14	48820	48833	<i>trnL(CAA)-trnF(GAA)</i>

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
37	p1	spacer	LSC	T	10	51310	51319	<i>ndhC-trnV(UAC)</i>
38	p1	spacer	LSC	A	14	52158	52171	<i>trnV(UAC)-trnM(CAU)</i>
39	p1	spacer	LSC	T	11	54626	54636	<i>atpB-rbcL</i>
40	p1	spacer	LSC	T	13	56570	56582	<i>rbcL-trnR(UCG)</i>
41	p1	spacer	LSC	T	13	56989	57001	<i>trnR(UCG)-accD</i>
42	p1	spacer	LSC	A	10	58098	58107	<i>accD-psaI</i>
43	p1	spacer	LSC	T	12	58522	58533	<i>accD-psaI</i>
44	p1	spacer	LSC	A	11	59707	59717	<i>ycf4-cemA</i>
45	p1	spacer	LSC	A	10	62694	62703	<i>petA-psbJ</i>
46	p1	spacer	LSC	T	14	63180	63193	<i>petA-psbJ</i>
47	p1	spacer	LSC	T	10	64840	64849	<i>psbE-petL</i>
48	p1	spacer	LSC	A	10	65211	65220	<i>petL-petG</i>
49	p1	spacer	LSC	C	12	66700	66711	<i>psaJ-rpl33</i>
50	p1	spacer	LSC	A	12	67888	67899	<i>rpl20-rps12</i>
51	p1	spacer	LSC	T	13	68130	68142	<i>rpl20-rps12</i>
52	p1	spacer	LSC	A	10	68290	68299	<i>rpl20-rps12</i>
53	p1	spacer	LSC	T	11	68557	68567	<i>rpl20-rps12</i>
54	p4	spacer	LSC	TTAT	12	69379	69390	<i>rps12-clpP</i>
55	p1	intron	LSC	C	10	70697	70706	<i>clpP</i>
56	p1	spacer	LSC	A	13	71660	71672	<i>clpP-psbB</i>
57	p1	spacer	LSC	T	10	71754	71763	<i>clpP-psbB</i>
58	p1	spacer	LSC	A	13	73543	73555	<i>psbB-psbT</i>
59	p1	spacer	LSC	A	10	74268	74277	<i>psbH-petB</i>
60	p1	spacer	LSC	T	13	77480	77492	<i>petD-rpoA</i>
61	p2	spacer	LSC	AT	20	80238	80257	<i>rps8-rpl14</i>
62	p2	spacer	LSC	AT	16	80338	80353	<i>rps8-rpl14</i>
63	p1	spacer	LSC	A	13	81186	81198	<i>rpl14-rpl16</i>
64	p1	intron	LSC	T	12	81919	81930	<i>rpl16</i>
65	p1	intron	LSC	T	12	82262	82273	<i>rpl16</i>
66	p1	spacer	LSC	T	11	82483	82493	<i>rpl16-rps3</i>
67	p1	intron	LSC	T	15	84995	85009	<i>rpl2</i>
68	p1	spacer	LSC	A	11	85683	85693	<i>rpl2-rpl23</i>
69	p1	spacer	LSC	T	12	85991	86002	<i>rpl23-trnI(CAU)</i>
70	p1	spacer	LSC	T	16	86136	86151	<i>rpl23-trnI(CAU)</i>
71	p4	gene	IRB	CTAC	12	91392	91403	<i>rrn23</i>
72	p2	intron	IRB	AG	14	94391	94404	<i>trnI(GAU)</i>
73	p4	spacer	IRB	TCTT	12	97681	97692	<i>rrn16-rps12</i>
74	p1	spacer	IRB	T	13	98774	98786	<i>rrn16-rps12</i>
75	p1	spacer	IRB	A	12	102750	102761	<i>trnH(GUG)-ycf2</i>
76	p4	CDS	IRB	TTTA	12	104706	104717	<i>ycf2</i>
77	p4	CDS	IRB	AGAA	12	107950	107961	<i>ycf2</i>
78	p1	spacer	IRB	A	15	109364	109378	<i>ycf2-trnN(GUU)</i>
79	p1	spacer	SSC	T	13	110416	110428	<i>ycf2-trnN(GUU)</i>
80	p1	spacer	SSC	A	11	112695	112705	<i>ndhF-rpl21</i>
81	p2	spacer	SSC	AT	18	113419	113436	<i>rpl21-rpl32</i>
82	p1	spacer	SSC	A	11	113761	113771	<i>rpl32-trnP(GGG)</i>
83	p1	spacer	SSC	T	13	114080	114092	<i>rpl32-trnP(GGG)</i>
84	p1	spacer	SSC	T	12	119615	119626	<i>trnP(GGG)-trnL(UAG)</i>
85	p1	spacer	SSC	A	10	119831	119840	<i>trnL(UAG)-ccsA</i>
86	p2	spacer	SSC	AT	16	119920	119935	<i>trnL(UAG)-ccsA</i>

SSR nr.	SSR type	Locus	Region	SSR motif	size	star	end	Location
87	p1	spacer	SSC	A	11	121168	121178	<i>ccsA-ndhD</i>
88	p1	spacer	SSC	A	14	121427	121440	<i>ccsA-ndhD</i>
89	p1	spacer	SSC	A	28	123088	123115	<i>ndhD-psaC</i>
90	p2	spacer	SSC	CT	12	124783	124794	<i>ndhG-ndhI</i>
91	p2	intron	SSC	TA	12	126492	126503	<i>ndhA</i>
92	p1	intron	SSC	T	13	127011	127023	<i>ndhA</i>
93	p1	spacer	SSC	A	10	129309	129318	<i>rps15-ycf1</i>
94	p1	spacer	SSC	A	10	129333	129342	<i>rps15-ycf1</i>
95	p1	spacer	IRA	T	15	139188	139202	<i>trnN(GUU)-ycf2</i>
96	p4	CDS	IRA	TTTC	12	140604	140615	<i>ycf2</i>
97	p4	CDS	IRA	TAAA	12	143849	143860	<i>ycf2</i>
98	p1	spacer	IRA	T	12	145805	145816	<i>ycf2-trnH(GUG)</i>
99	p1	spacer	IRA	A	13	149780	149792	<i>rps12-rrn16</i>
100	p4	spacer	IRA	AAAG	12	150873	150884	<i>rps12-rrn16</i>
101	p2	intron	IRA	TC	14	154161	154174	<i>trnI(GAU)</i>
102	p4	gene	IRA	AGGT	12	157161	157172	<i>rrn23</i>

Appendix table 9 The number, relative abundance, relative density, GC content of SSR motifs in the chloroplast genomes of 8 Cyatheaceae species

motif type	Characteristics of SSR	<i>A. denticulata</i>	<i>A. podophylla</i>	<i>A. gigantea</i>	<i>A. metteniana</i>	<i>A. costularis</i>
A	No. of SSRs	13	10	10	11	31
	Relative abundance (No./Kb)	0.084	0.060	0.062	0.068	0.198
	Relative density (bp/Kb)	0.883	0.614	0.730	0.780	2.247
C	No. of SSRs	8	8	8	6	10
	Relative abundance (No./Kb)	0.052	0.048	0.049	0.037	0.064
	Relative density (bp/Kb)	0.565	0.554	0.637	0.421	1.040
G	No. of SSRs	6	7	7	6	8
	Relative abundance (No./Kb)	0.039	0.042	0.043	0.037	0.051
	Relative density (bp/Kb)	0.487	0.542	0.619	0.446	0.645
T	No. of SSRs	13	22	16	17	46
	Relative abundance (No./Kb)	0.084	0.132	0.099	0.105	0.294
	Relative density (bp/Kb)	0.993	1.450	1.076	1.157	3.606
AG	No. of SSRs	0	0	1	1	0
	Relative abundance (No./Kb)	0	0	0.006	0.006	0
	Relative density (bp/Kb)	0	0	0.148	0.099	0
AT	No. of SSRs	5	6	6	6	11
	Relative abundance (No./Kb)	0.032	0.036	0.037	0.037	0.070
	Relative density (bp/Kb)	0.344	0.518	0.495	0.520	1.111
CT	No. of SSRs	1	0	0	0	0
	Relative abundance (No./Kb)	0.006	0	0	0	0
	Relative density (bp/Kb)	0.078	0	0	0	0
TA	No. of SSRs	2	3	3	3	3
	Relative abundance (No./Kb)	0.013	0.018	0.019	0.019	0.019
	Relative density (bp/Kb)	0.078	0.313	0.297	0.297	0.243
TC	No. of SSRs	0	0	0	0	0
	Relative abundance (No./Kb)	0	0	0	0	0
	Relative density (bp/Kb)	0	0	0	0	0
AAG	No. of SSRs	0	0	0	0	1

motif type	Characteristics of SSR	<i>A. denticulata</i>	<i>A. podophylla</i>	<i>A. gigantea</i>	<i>A. metteniana</i>	<i>A. costularis</i>
AAT	Relative abundance (No./Kb)	0	0	0	0	0.006
	Relative density (bp/Kb)	0	0	0	0	0.077
TAT	No. of SSRs	1	1	1	1	0
	Relative abundance (No./Kb)	0.006	0.006	0.006	0.006	0
AAAG	Relative density (bp/Kb)	0.078	0.072	0.074	0.074	0
	No. of SSRs	1	0	1	1	0
AAAT	Relative abundance (No./Kb)	0.006	0	0.006	0.006	0
	Relative density (bp/Kb)	0.078	0	0.074	0.074	0.077
AATT	No. of SSRs	0.000	1	1	1	0
	Relative abundance (No./Kb)	0.000	0.006	0.006	0.006	0
ACCA	Relative density (bp/Kb)	0.000	0.072	0.099	0.099	0
	No. of SSRs	2	2	1	2	0
AGAA	Relative abundance (No./Kb)	0.013	0.012	0.006	0.012	0
	Relative density (bp/Kb)	0.156	0.144	0.074	0.149	0
AGGT	No. of SSRs	0	0	0	0	1
	Relative abundance (No./Kb)	0	0	0	0	0.006
ATAG	Relative density (bp/Kb)	0	0	0	0	0.077
	No. of SSRs	2	2	2	2	1
ATCT	Relative abundance (No./Kb)	0.013	0.012	0.012	0.012	0.006
	Relative density (bp/Kb)	0.156	0.144	0.148	0.149	0.077
CCTC	No. of SSRs	0	0	0	0	1
	Relative abundance (No./Kb)	0	0	0	0	0.006
CTAC	Relative density (bp/Kb)	0	0	0	0	0.102
	No. of SSRs	0	0	0	0	0
GAAA	Relative abundance (No./Kb)	0	0	0	0	0
	Relative density (bp/Kb)	0	0	0	0	0
GATA	No. of SSRs	0	0	1	1	0
	Relative abundance (No./Kb)	0	0	0.006	0.006	0
TAAA	Relative density (bp/Kb)	0	0	0.074	0.074	0
	No. of SSRs	1	1	1	1	1
TATC	Relative abundance (No./Kb)	0.006	0.006	0.006	0.006	0.006
	Relative density (bp/Kb)	0.078	0.072	0.074	0.074	0.077
	No. of SSRs	0	0	0	0	0
	Relative abundance (No./Kb)	0	0	0	0	0
	Relative density (bp/Kb)	0	0	0	0	0
	No. of SSRs	1	1	1	1	1
	Relative abundance (No./Kb)	0.006	0.006	0.006	0.006	0.006
	Relative density (bp/Kb)	0.104	0.096	0.074	0.074	0.077

motif type	Characteristics of SSR	<i>A. denticulata</i>	<i>A. podophylla</i>	<i>A. gigantea</i>	<i>A. metteniana</i>	<i>A. costularis</i>
TATT	No. of SSRs	1	1	1	1	1
	Relative abundance (No./Kb)	0.006	0.006	0.006	0.006	0.006
	Relative density (bp/Kb)	0.078	0.072	0.074	0.074	0.077
TCTT	No. of SSRs	1	0	1	1	1
	Relative abundance (No./Kb)	0.006	0	0.006	0.006	0.006
	Relative density (bp/Kb)	0.078	0	0.074	0.074	0.077
TGAT	No. of SSRs	1	1	1	1	0
	Relative abundance (No./Kb)	0.006	0.006	0.006	0.006	0
	Relative density (bp/Kb)	0.078	0.072	0.074	0.074	0
TTAA	No. of SSRs	0	0	0	0	0
	Relative abundance (No./Kb)	0	0	0	0	0
	Relative density (bp/Kb)	0	0	0	0	0
TTAT	No. of SSRs	0	0	0	0	0
	Relative abundance (No./Kb)	0	0	0	0	0
	Relative density (bp/Kb)	0	0	0	0	0
TTTA	No. of SSRs	0	0	0	0	0
	Relative abundance (No./Kb)	0	0	0	0	0
	Relative density (bp/Kb)	0	0	0	0	0
TTTC	No. of SSRs	0	0	0	0	1
	Relative abundance (No./Kb)	0	0	0	0	0.006
	Relative density (bp/Kb)	0	0	0	0	0.077
ATTTT	No. of SSRs	1	0	0	0	0
	Relative abundance (No./Kb)	0.006	0	0	0	0
	Relative density (bp/Kb)	0.097	0	0	0	0
CTTTC	No. of SSRs	0	0	0	1	0
	Relative abundance (No./Kb)	0	0	0	0.006	0
	Relative density (bp/Kb)	0	0	0	0.093	0
TTCTC	No. of SSRs	0	1	0	0	0
	Relative abundance (No./Kb)	0	0.006	0	0	0
	Relative density (bp/Kb)	0	0.090	0	0	0

Appendix table 10 The number, relative abundance, relative density and GC content of different types of motif SSRs among different regions in the chloroplast genomes of 8 Cyatheaceae species

Species	motif type	Characteristics of SSR	IGS	intron	CDS	gene	LSC	SSC	IRA	IRE
<i>A. denticulata</i>	mononucleotide	No.of SSRs	30	10	0	0	30	8	1	1
		Relative abundance (No./Kb)	0.63	0.66	0	0	0.35	0.37	0.04	0.04
		Relative density (bp/Kb)	6.81	7.82	0	0	3.92	4.17	0.52	0.52
		GC content	0.40	0.27	0	0	0.45	0.12	0	0
	Dinucleotide	No.of SSRs	7	1	0	0	4	2	1	1
		Relative abundance (No./Kb)	0.15	0.07	0	0	0.05	0.09	0.04	0.04
		Relative density (bp/Kb)	1.35	0.79	0	0	0.63	1.30	0.52	0.52
		GC content	0	0.50	0	0	0.11	0	0	0
	Trinucleotide	No.of SSRs	2	0	0	0	0	2	0	0
		Relative abundance (No./Kb)	0.04	0	0	0	0	0.09	0	0
		Relative density (bp/Kb)	0.51	0	0	0	0	1.11	0	0
		GC content	0	0	0	0	0	0	0	0
	Tetranucleotide	No.of SSRs	9	1	0	0	7	1	1	1
		Relative abundance (No./Kb)	0.19	0.07	0	0	0.08	0.05	0.04	0.04

Species	motif type	Characteristics of SSR	IGS	intron	CDS	gene	LSC	SSC	IRA	IRE
<i>A. podophylla</i>	Pentanucleotide	Relative density (bp/Kb)	2.37	0.79	0	0	1.02	0.56	0.52	0.52
		GC content	0.20	0	0	0	0.18	0	0.25	0.25
		No.of SSRs	1	0	0	0	1	0	0	0
		Relative abundance (No./Kb)	0.02	0	0	0	0.01	0	0	0
		Relative density (bp/Kb)	0.32	0	0	0	0.17	0	0	0
	mononucleotide	GC content	0	0	0	0	0	0	0	0
		No.of SSRs	38	9	0	0	32	7	4	4
		Relative abundance (No./Kb)	0.66	0.59	0	0	0.37	0.32	0.14	0.14
		Relative density (bp/Kb)	7.33	6.78	0	0	4.09	3.97	1.45	1.45
		GC content	0.41	0.11	0	0	0.38	0.30	0.24	0.24
<i>A. gigantea</i>	Dinucleotide	No.of SSRs	8	1	0	0	4	3	1	1
		Relative abundance (No./Kb)	0.14	0.07	0	0	0.05	0.14	0.03	0.03
		Relative density (bp/Kb)	2.12	1.05	0	0	0.71	2.40	0.42	0.42
		GC content	0	0	0	0	0	0	0	0
		No.of SSRs	0	1	0	0	1	0	0	0
	Trinucleotide	Relative abundance (No./Kb)	0	0.07	0	0	0.01	0	0	0
		Relative density (bp/Kb)	0	0.79	0	0	0.14	0	0	0
		GC content	0	0	0	0	0	0	0	0
		No.of SSRs	8	1	0	0	7	2	0	0
		Relative abundance (No./Kb)	0.14	0.07	0	0	0.08	0.09	0	0
<i>A. metteniana</i>	Tetranucleotide	Relative density (bp/Kb)	1.74	0.79	0	0	1.01	1.11	0	0
		GC content	0.16	0	0	0	0.18	0	0	0
		No.of SSRs	0	1	0	0	1	0	0	0
		Relative abundance (No./Kb)	0	0.07	0	0	0.01	0	0	0
		Relative density (bp/Kb)	0	0.99	0	0	0.17	0	0	0
	Pentanucleotide	GC content	0	0.40	0	0	0.40	0	0	0
		No.of SSRs	33	8	0	0	30	7	2	2
		Relative abundance (No./Kb)	0.62	0.53	0	0	0.32	0.32	0.08	0.08
		Relative density (bp/Kb)	7.89	5.66	0	0	3.52	6.13	1.01	1.01
		GC content	0.41	0.36	0	0	0.43	0.47	0	0
<i>A. metteniana</i>	Dinucleotide	No.of SSRs	9	1	0	0	5	3	1	1
		Relative abundance (No./Kb)	0.17	0.07	0	0	0.05	0.14	0.04	0.04
		Relative density (bp/Kb)	2.59	0.92	0	0	0.91	2.03	0.50	0.50
		GC content	0.09	0	0	0	0.14	0	0	0
		No.of SSRs	2	0	0	0	0	2	0	0
	Trinucleotide	Relative abundance (No./Kb)	0.04	0	0	0	0	0.09	0	0
		Relative density (bp/Kb)	0.45	0	0	0	0	1.11	0	0
		GC content	0	0	0	0	0	0	0	0
		No.of SSRs	10	1	0	0	7	2	1	1
		Relative abundance (No./Kb)	0.19	0.07	0	0	0.08	0.09	0.04	0.04
<i>A. metteniana</i>	Tetranucleotide	Relative density (bp/Kb)	2.33	0.79	0	0	0.91	1.29	0.50	0.50
		GC content	0.19	0	0	0	0.18	0.11	0.25	0.25
		No.of SSRs	0	0	0	0	0	0	0	0
		Relative abundance (No./Kb)	0	0	0	0	0	0	0	0
		Relative density (bp/Kb)	0	0	0	0	0	0	0	0
	Pentanucleotide	GC content	0	0	0	0	0	0	0	0
		No.of SSRs	33	7	0	0	25	9	3	3
		Relative abundance (No./Kb)	0.62	0.46	0	0	0.27	0.42	0.13	0.13
		Relative density (bp/Kb)	7.29	5.14	0	0	2.96	5.58	1.51	1.51
		GC content	0.28	0.38	0	0	0.41	0.24	0	0

Species	motif type	Characteristics of SSR	IGS	intron	CDS	gene	LSC	SSC	IRA	IRE
<i>A. costularis</i>	Dinucleotide	No.of SSRs	9	1	0	0	5	3	1	1
		Relative abundance (No./Kb)	0.17	0.07	0	0	0.05	0.14	0.04	0.04
		Relative density (bp/Kb)	2.52	0.92	0	0	0.85	2.12	0.50	0.50
		GC content	0.06	0	0	0	0.10	0	0	0
<i>A. spinulosa</i>	Trinucleotide	No.of SSRs	2	0	0	0	0	2	0	0
		Relative abundance (No./Kb)	0.04	0	0	0	0	0.09	0	0
		Relative density (bp/Kb)	0.45	0	0	0	0	1.11	0	0
		GC content	0	0	0	0	0	0	0	0
<i>A. costularis</i>	Tetranucleotide	No.of SSRs	11	1	0	0	7	3	1	1
		Relative abundance (No./Kb)	0.21	0.07	0	0	0.08	0.14	0.04	0.04
		Relative density (bp/Kb)	2.56	0.79	0	0	0.91	1.85	0.50	0.50
		GC content	0.15	0	0	0	0.18	0.08	0.25	0.25
<i>A. spinulosa</i>	Pentanucleotide	No.of SSRs	1	0	0	0	1	0	0	0
		Relative abundance (No./Kb)	0.02	0	0	0	0.01	0	0	0
		Relative density (bp/Kb)	0.28	0	0	0	0.16	0	0	0
		GC content	0.4	0	0	0	0.4	0	0	0
<i>A. costularis</i>	mononucleotide	No.of SSRs	80	15	0	0	67	16	6	6
		Relative abundance (No./Kb)	1.66	0.94	0	0	0.78	0.74	0.25	0.25
		Relative density (bp/Kb)	20.49	13.27	0	0	9.78	8.83	3.41	3.41
		GC content	0.19	0.36	0	0	0.20	0.19	0.36	0.36
<i>A. spinulosa</i>	Dinucleotide	No.of SSRs	13	1	0	0	7	3	2	2
		Relative abundance (No./Kb)	0.27	0.06	0	0	0.08	0.14	0.08	0.08
		Relative density (bp/Kb)	3.89	1.50	0	0	1.32	2.31	0.99	0.99
		GC content	0	0	0	0	0	0	0	0
<i>A. costularis</i>	Trinucleotide	No.of SSRs	0	1	0	0	1	0	0	0
		Relative abundance (No./Kb)	0	0.06	0	0	0.01	0	0	0
		Relative density (bp/Kb)	0	0.75	0	0	0.14	0	0	0
		GC content	0	0.33	0	0	0.33	0	0	0
<i>A. spinulosa</i>	Tetranucleotide	No.of SSRs	7	1	3	0	6	1	2	2
		Relative abundance (No./Kb)	0.14	0.06	0.04	0	0.07	0.05	0.08	0.08
		Relative density (bp/Kb)	1.82	0.75	0.45	0	0.88	0.55	0.99	0.99
		GC content	0.22	0.75	0.33	0	0.29	0.50	0.25	0.25
<i>A. costularis</i>	Pentanucleotide	No.of SSRs	0	0	0	0	0	0	0	0
		Relative abundance (No./Kb)	0	0	0	0	0	0	0	0
		Relative density (bp/Kb)	0	0	0	0	0	0	0	0
		GC content	0	0	0	0	0	0	0	0
<i>A. spinulosa</i>	mononucleotide	No.of SSRs	81	15	0	0	68	16	6	6
		Relative abundance (No./Kb)	1.68	0.99	0	0	0.79	0.74	0.25	0.25
		Relative density (bp/Kb)	20.73	12.45	0	0	9.63	8.74	3.49	3.49
		GC content	0.18	0.28	0	0	0.17	0.19	0.31	0.31
<i>A. costularis</i>	Dinucleotide	No.of SSRs	13	1	0	0	7	3	2	2
		Relative abundance (No./Kb)	0.27	0.07	0	0	0.08	0.14	0.08	0.08
		Relative density (bp/Kb)	3.89	1.58	0	0	1.32	2.31	0.99	0.99
		GC content	0	0	0	0	0	0	0	0
<i>A. spinulosa</i>	Trinucleotide	No.of SSRs	0	1	0	0	1	0	0	0
		Relative abundance (No./Kb)	0	0.07	0	0	0.01	0	0	0
		Relative density (bp/Kb)	0	0.79	0	0	0.14	0	0	0
		GC content	0	0.33	0	0	0.33	0	0	0
<i>A. costularis</i>	Tetranucleotide	No.of SSRs	7	1	3	0	6	1	2	2
		Relative abundance (No./Kb)	0.14	0.07	0.04	0	0.07	0.05	0.08	0.08

Species	motif type	Characteristics of SSR	IGS	intron	CDS	gene	LSC	SSC	IRA	IRE
<i>S. brunoniana</i>	Pentanucleotide	Relative density (bp/Kb)	1.57	0.79	0.45	0	0.74	0.55	0.99	0.99
		GC content	0.25	0.75	0.33	0	0.34	0.50	0.25	0.25
		No.of SSRs	0	0	0	0	0	0	0	0
		Relative abundance (No./Kb)	0	0	0	0	0	0	0	0
		Relative density (bp/Kb)	0	0	0	0	0	0	0	0
	mononucleotide	GC content	0	0	0	0	0	0	0	0
		No.of SSRs	62	15	0	0	61	10	3	3
		Relative abundance (No./Kb)	1.29	1.04	0	0	0.71	0.45	0.12	0.12
		Relative density (bp/Kb)	17.20	12.25	0	0	9.22	5.66	1.71	1.71
		GC content	0.08	0.08	0	0	0.10	0	0	0
<i>S. lepifera</i>	Dinucleotide	No.of SSRs	9	6	0	0	10	3	1	1
		Relative abundance (No./Kb)	0.19	0.42	0	0	0.12	0.13	0.04	0.04
		Relative density (bp/Kb)	2.95	5.40	0	0	1.72	2.14	0.50	0.50
		GC content	0.04	0.24	0	0	0.05	0.13	0.50	0.50
		No.of SSRs	0	0	0	0	0	0	0	0
	Trinucleotide	Relative abundance (No./Kb)	0	0	0	0	0	0	0	0
		Relative density (bp/Kb)	0	0	0	0	0	0	0	0
		GC content	0	0	0	0	0	0	0	0
		No.of SSRs	6	0	2	2	4	0	3	3
		Relative abundance (No./Kb)	0.12	0	0.02	0.22	0.05	0	0.12	0.12
<i>S. lepifera</i>	Tetranucleotide	Relative density (bp/Kb)	1.66	0	0.30	2.64	0.65	0	1.50	1.50
		GC content	0.25	0	0.25	0.5	0.25	0	0.33	0.33
		No.of SSRs	0	0	0	0	0	0	0	0
		Relative abundance (No./Kb)	0	0	0	0	0	0	0	0
		Relative density (bp/Kb)	0	0	0	0	0	0	0	0
	Pentanucleotide	GC content	0	0	0	0	0	0	0	0
		No.of SSRs	0	0	0	0	0	0	0	0
		Relative abundance (No./Kb)	0	0	0	0	0	0	0	0
		Relative density (bp/Kb)	0	0	0	0	0	0	0	0
		GC content	0	0	0	0	0	0	0	0

Appendix table 11 Significant differences in the number, relative abundance, relative density, and GC content of the chloroplast genomes, Mono- to Pentanucleotide SSRs, IGS, LSC, intron and CDS regionsSSRs of

chloroplast genomes, and the mono- to pentanucleotide SSRs in the chloroplast genomes of 8 Cyatheaceae species

Classification treatment	Characteristics of SSR	P	P	P	P	P	P
3 genera	No.of SSRs	0.048*	0.048*	0.065	0.057	0.70	
	Relative abundance (No./Kb)	0.033*	0.050*	0.069	0.103	0.40	
	Relative density	0.050*	0.050*	0.069	0.103	0.40	
	GC content	0.050*	0.050*	0.068	0.030*	0.00	
2 genera	No.of SSRs	0.502	0.502	0.177	0.034*	0.72	
	Relative abundance (No./Kb)	0.478	0.505	0.182	0.044*	0.18	
	Relative density	0.505	0.505	0.182	0.044*	0.50	
	GC content	0.046*	0.046*	0.040*	0.378	0.17	
P	P	P	P	P	P	P	P
IGS-Mono	intron-Mono	CDS-Mono	rRNA gene-Mono	LSC-Mono	SSC-Mono	IRAM	
	0.048	0.056	1	1	0.048	0.046*	0.11
	0.05	0.069	1	1	0.050*	0.050*	0.13
	0.05	0.050*	1	1	0.050*	0.105	0.05
	0.05	0.223	1	1	0.050*	0.103	0.00
	0.502	0.306	1	1	0.502	0.500	0.73
	0.505	0.182	1	1	0.505	0.505	0.50
	0.505	0.505	1	1	0.505	1	0.50
P	P	P	P	P	P	P	P
IGS-Tri	intron-Tri	CDS-Tri	rRNA gene-Tri	LSC-Tri	SSC-Tri	IRAT	
	0.122	0.122	1	1	0.122	0.122	1
	0.14	0.140	1	1	0.067	0.14	1
	0.14	0.140	1	1	0.067	0.14	1
	1	0.030*	1	1	0.030*	1	1
	0.237	0.237	1	1	0.237	0.237	1
	0.252	0.252	1	1	0.252	0.252	1
	0.252	0.252	1	1	0.252	0.252	1
P	P	P	P	P	P	P	P
IGS-Penta	intron-Penta	CDS-Penta	rRNA gene-Penta	LSC-Penta	SSC-Penta	IRAP	
	0.311	0.607	1	1	0.122	1	1
	0.319	0.607	1	1	0.140	1	1
	0.319	0.607	1	1	0.140	1	1
	0.607	0.607	1	1	0.311	1	1
	0.378	0.564	1	1	0.237	1	1
	0.383	0.564	1	1	0.252	1	1
	0.383	0.564	1	1	0.252	1	1
P	P	P	P	P	P	P	P

When 8 Cyatheaceae plants are treated as three genera, Kruskal-Wallis H test is used; when treated as two genera, Mann-Whitney U test is used; *: $P \leq 0.05$; IGS: Intergenic Spacer-Region; LSC: Large Single Copy-Region; CDS: Coding Sequence-Region