

1 **Sedimentary ancient DNA from Kronotsky Peninsula: how sea ice, salinity and**
2 **insolation dynamics have shaped diatom composition and richness over the past**
3 **20,000 years**

4 **H. H. Zimmermann¹, K. R. Stoof-Leichsenring¹, S. Kruse¹, D. Nürnberg², R. Tiedemann³,**
5 **and U. Herzschuh^{1,4,5}**

6 ¹Polar Terrestrial Environmental Systems, Alfred Wegener Institute Helmholtz Centre for Polar
7 and Marine Research, Potsdam, 14473, Germany

8 ²Ocean circulation and climate dynamics, GEOMAR Helmholtz Centre for Ocean Research Kiel,
9 Kiel, 24148, Germany

10 ³Marine Geology, Alfred Wegener Institute Helmholtz Centre for Polar and Marine Research,
11 Bremerhaven, 27568, Germany

12 ⁴Institute of Biochemistry and Biology, University of Potsdam, 14476 Potsdam, Germany

13 ⁵Institute of Environmental Sciences and Geography, University of Potsdam, 14476 Potsdam,
14 Germany

15
16 Corresponding author: Heike H. Zimmermann (heike.zimmermann@awi.de), Ulrike Herzschuh
17 (ulrike.herzschuh@awi.de)

18
19 **Key Points:**

- 20 • diatom *seda*DNA composition is concordant with reconstructed sea-ice dynamics, SSTs
21 and subsurface salinities over the past millenia
- 22 • loss of diatom richness at ~11.1 cal kyr BP is possibly a consequence of increased
23 freshwater input from Kamchatka
- 24 • shifts of potential ecotypes of species from the genus *Chaetoceros* with changing
25 environmental conditions
26

27 **Abstract**

28 We traced diatom composition and diversity through time using diatom derived sedimentary
29 ancient DNA (*sedaDNA*) from eastern continental slope sediments off Kamchatka (North
30 Pacific) by applying a short, diatom-specific marker on 63 samples in a DNA metabarcoding
31 approach. The sequences were assigned to diatoms that are common in the area and
32 characteristic of cold water. *SedaDNA* allowed us to observe shifts of potential lineages from
33 species of the genus *Chaetoceros* that can be related to different climatic phases, suggesting that
34 pre-adapted ecotypes might have played a role in the long-term success of species in areas of
35 changing environmental conditions. These *sedaDNA* results complement our understanding of
36 the long-term history of diatom assemblages and their general relationship to environmental
37 conditions of the past. Sea-ice diatoms (*Pauliella taeniata* (Grunow) Round & Basson, *Attheya*
38 *septentrionalis* (Østrup) R.M.Crawford and *Nitzschia frigida* (Grunow)) detected during the late
39 glacial and Younger Dryas are in agreement with previous sea-ice reconstructions. A positive
40 correlation between pennate diatom richness and the sea-ice proxy IP₂₅ suggests that sea ice
41 fosters pennate diatom richness, whereas a negative correlation with June insolation and
42 temperature points to unfavorable conditions during the Holocene. A sharp increase in
43 proportions of freshwater diatoms at ~11.1 cal kyr BP implies the influence of terrestrial runoff
44 and coincides with the loss of 42% of diatom sequence variants. We assume that reduced salinity
45 at this time stabilized vertical stratification which limited the replenishment of nutrients in the
46 euphotic zone.

47

48 **1 Introduction**

49 Climate warming is transforming arctic and subarctic ecosystems by a reduction in the
50 duration, extent and thickness of sea ice (Parkinson et al., 1999; Walsh et al., 2017) – a key variable
51 of the global climate system whose disappearance is resulting in a positive temperature feedback
52 from a reduction of the surface albedo (Pistone et al., 2014). Rapidly melting glaciers and increased
53 riverine runoff resulting from thawing permafrost and augmented precipitation over the adjacent
54 continents are expected to decrease sea-surface salinities, for example by about 1.5 ± 1.1 psu on
55 average in the Arctic Ocean (Shu et al., 2018). As a consequence, the fresher, less dense water
56 masses are expected to enhance vertical water-column stratification, which could result in a
57 reduced supply of nutrients to the euphotic zone (Tremblay & Gagnon, 2009). On top of regionally
58 expected loss of sea ice-associated organisms, such a scenario could potentially affect the
59 distribution, composition and diversity of primary producers which are limited by the availability
60 of nutrients, amongst others, with unknown consequences for food-web structure, biochemical
61 cycles and the biological carbon pump (Coupel et al., 2015; Li et al., 2009).

62 The subarctic northwest (NW) Pacific and its adjacent seas have experienced pronounced
63 environmental changes since the Last Glacial Maximum (LGM). The variability of previously
64 reconstructed sea-surface temperatures and sea-ice coverage have been connected to the millennial
65 scale climatic changes recorded in sediment cores from the North Atlantic and in Greenland ice
66 cores (Max et al., 2012; Méheust et al., 2016). Several meltwater pulses have been detected in
67 sediment cores of this region (Gorbarenko et al., 2019), which make it attractive for analyzing the
68 effects of past environmental changes on primary producers.

69 A dominant group of primary producers in high-latitude sea-ice and coastal ecosystems
70 are diatoms (Poulin et al., 2011), which are single-celled eukaryotes that form a cell wall of
71 biomineralized silicium dioxide (biogenic silica) and fix carbon dioxide as a product of their
72 photosynthetic activity (Rotatore et al., 1995). They are sensitive to environmental change and can
73 be used as indicators of past sea-surface conditions. For example, morphological data from diatom
74 assemblages of the subarctic NW Pacific and the Bering and Okhotsk Seas have revealed
75 variations in ocean circulation patterns, past distribution of sea ice and past sea-surface
76 temperatures (Caissie et al., 2010; Ren et al., 2009; Sancetta, 1979; Sancetta & Silvestri, 1986;
77 Smirnova et al., 2015). While the investigation of microfossils can distinguish size distributions,
78 resting stages and, in many instances, taxonomic resolution to species level for millions of years
79 back in time, some questions cannot be addressed with morphological data. Amongst others, this
80 includes intra-specific diversity which may be linked to different environmental conditions.

81 The past decade has brought forward genetic surveys which have revealed substantially
82 concealed diversity in diatoms, showing that widely distributed species can have ecotypes
83 associated with, for example, different growth optima (Hamsher et al., 2013) that may not be
84 detected morphologically (cryptic diversity) or only by exhaustive analysis (pseudo-cryptic)
85 (Degerlund et al., 2012). In this regard, the analysis of *sedaDNA* is an advantage. Recently,
86 *sedaDNA* was used as a proxy for sea-ice reconstructions by targeting diatom *sedaDNA*
87 composition (Zimmermann et al., 2019), for tracing a sea-ice dinoflagellate east of Greenland (De
88 Schepper et al., 2019) and to identify changes of ocean circulation patterns by targeting diatoms
89 and non-fossilized foraminifers east of Svalbard (Pawłowska et al., 2020). Ancient DNA has
90 profound advantages. First, it can detect species that are absent from microfossil records either due
91 to dissolution or because they lack a biomineralized cell wall. Second, it allows for the detection
92 of (pseudo-)cryptic diversity in morphological species complexes, which might be adapted to

93 different ecological conditions. Third, it is amongst the most applied barcoding gene and thus has
94 a substantial reference database (Rimet et al., 2019; Zimmermann et al., 2014). However, ancient
95 DNA is usually highly degraded (Corinaldesi et al., 2008; Pääbo, 1989). Hence, for DNA
96 metabarcoding of diatoms, the marker should ideally target a short sequence stretch, and
97 additionally avoid co-amplification of other taxonomic groups or co-amplification of highly
98 similar paralogs. Targeting a part of the gene encoding for the large subunit of the RuBisCO (*rbcL*)
99 on the chloroplast genome of diatoms is of advantage. The variability of the *rbcL* gene allows
100 detection of diatoms to species level (Rimet et al., 2019).

101 Our aim is to trace temporal changes of diatom *seada*DNA composition and diversity
102 over the past 19.9 cal kyr BP, and to relate changes to environmental conditions to answer the
103 following questions: (1) How has diatom composition changed since the LGM? And how are such
104 changes related to past sea-ice conditions, subsurface salinities and incoming solar radiation
105 (insolation)? (2) How did diatom *seada*DNA richness change over time?

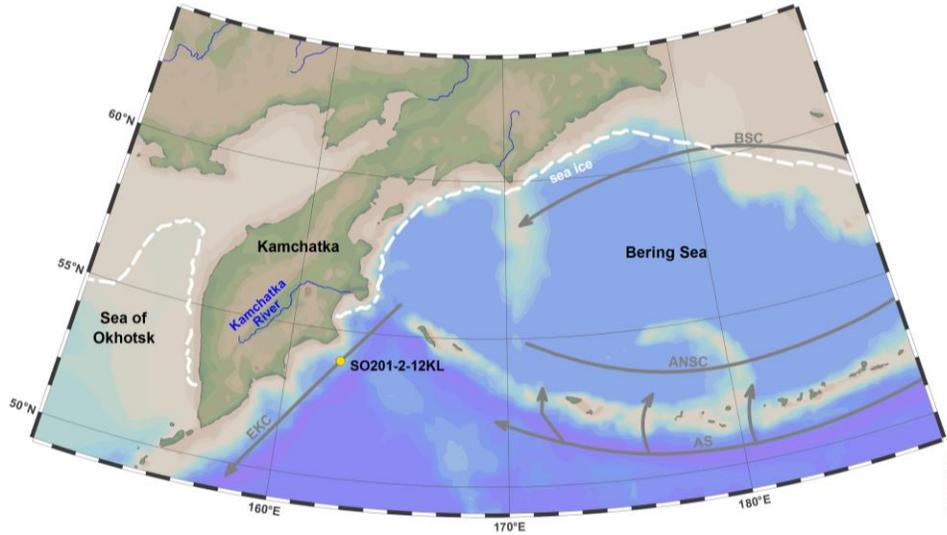
106

107 **2 Materials and Methods**

108 **2.1 Study site**

109 The sediment core SO201-2-12KL was collected from the Kamchatka Strait near
110 Kronotskii Peninsula at the eastern continental slope of Kamchatka where the shelf area is very
111 narrow (Fig. 1). The study area is influenced by water masses transported via the East Kamchatka
112 Current, which brings nutrient-rich waters from the Bering Sea to the subarctic North Pacific
113 Ocean (Stabeno et al., 1999) and by the Alaskan Stream, which transports water masses from the
114 Gulf of Alaska along the Aleutian Arc and water from the wind-driven, cyclonic western subarctic
115 gyre (Nagano et al., 2016; Stabeno & Reed, 1994). As evaporation is lower than precipitation and
116 runoff from Kamchatka, the water column is characterized by a stable halocline (Gebhardt et al.,
117 2008). Modern sea-surface temperatures in the area are about 1 °C in winter and range between 6
118 and 10 °C in summer (Riethdorf et al., 2013a) with winter sea-ice formation only along the coast
119 of Kamchatka (Matul' et al., 2015), but is influenced by drifting sea ice (Polyakova, 2007).

120



121
 122 **Figure 1.** Map showing the study area with surface current patterns (EKC = East Kamchatka
 123 Current, AS = Alaskan Stream, ANSC = Aleutian North Slope Current; BSC = Bering Sea Current)
 124 indicated by gray arrows and the coring site SO201-2-12KL marked by a yellow dot. The white
 125 dashed line represents median sea-ice extent in March between 1981 and 2010 (Fetterer et al.,
 126 2017). The map was produced with Ocean Data View (Schlitzer, 2002).

127

128 2.2 Sample material

129 Sediment material was retrieved with a piston corer (SO201-2-12KL) during RV Sonne
 130 cruise SO-201 (KALMAR) in (Max et al., 2012). The sediment core is 9.05 m long and mostly
 131 composed of sandy-silty-clayey mud. We here rely on the chronostratigraphy established by (Max
 132 et al., 2012). Core material was stored at 4 °C since retrieval. Samples for ancient DNA analyses
 133 were collected in 2018 at GEOMAR, Kiel, in a laboratory devoid of any molecular biology work.
 134 To avoid contamination of the samples with modern DNA, sampling was performed from the
 135 oldest to the youngest samples while wearing a plastic coverall, face mask, hair net. Gloves were
 136 changed between each sample. The treatment of the equipment and the sampling procedure for
 137 ancient DNA analyses followed the protocol for non-frozen sediment cores explained in Epp et al.
 138 (2019).

139

140 2.3. DNA extraction and concentration

141 The DNA extractions, PCR setups and reaction conditions were carried out in a dedicated
142 laboratory for ancient DNA at AWI Potsdam, while PCRs and downstream preparations for
143 sequencing were carried out in the genetics laboratories located in another building. Total DNA
144 was extracted using the DNeasy PowerMax Soil kit (Qiagen, Hilden, Germany) as described in
145 Zimmermann et al. (2017) from 63 samples (2–4 ml sediment). Each extraction batch contained
146 up to 9 samples and one negative control (in total 7 negative controls). Subsequently, we measured
147 the total DNA concentration on a Qubit 4.0 fluorometer (Invitrogen, Carlsbad, CA, USA) using
148 the Qubit dsDNA BR Assay Kit (Invitrogen, Carlsbad, CA, USA). We concentrated for each
149 sample 600 µl of the extracted DNA using the GeneJET PCR Purification KIT (Thermo Scientific,
150 Carlsbad, CA, USA) and eluted twice with 15 µl elution buffer to retain a final volume of ~30 µl.
151 The DNA concentration was measured again and the DNA was diluted to 3 ng/µl. The DNA
152 extracts and aliquots were stored at -20 °C.

153

154 2.4. *SedaDNA* metabarcoding

155 *SedaDNA* metabarcoding was carried out using the *rbcl_76* marker which was designed
156 for both marine and freshwater species (Stoof-Leichsenring et al., 2012). Amplifications were
157 carried out using the primers *Diat_rbcL_705F* (AACAGGTGAAGTTAAAGGTTTCATAYTT)
158 and *Diat_rbcL_808R* (TGTAACCCATAACTAAATCGATCAT), which were tagged for parallel
159 sequencing (Binladen et al., 2007) as described in Dulias et al. (2017) and Huang et al. (2020).
160 The PCR reaction mixes and conditions were prepared according to the adjusted protocol for
161 tagged *Diat_rbcL_705F* and *Diat_rbcL_808R* primers as described in Dulias et al. (2017) with the
162 exception that 3 µl DNA (DNA concentration 3 ng/µl) was used as a template. PCRs were carried
163 out under the following conditions: 5 minutes at 94 °C (initial denaturation), then 50 cycles at 94
164 °C (denaturation), 49 °C (annealing) and 68 °C (elongation) and a final elongation step at 72 °C
165 for 5 minutes. Each PCR batch was composed of one extraction batch (9 samples and the
166 corresponding extraction negative control) and a PCR negative control. Furthermore, each PCR
167 batch was subjected to three independent PCRs, with distinct primer-tag combinations on different

168 days. PCR success was checked with gel-electrophoresis. Subsequently, the PCR products
169 including all negative controls were purified using the MinElute purification Kit (Qiagen, Hilden,
170 Germany). The DNA concentrations of the PCR products were measured again with the Qubit
171 dsDNA BR Assay Kit and mixed in equal concentrations. As negative controls were below
172 detection limit, 10 µl for negative controls were added to the sample pool. The sample pool was
173 sent to FASTERIS SA sequencing service (Switzerland) who carried out library preparation with the
174 Mid Output kit v 2 according to the FASTERIS Metafast protocol and sequencing (2 x 150 bp, paired-
175 end) on the Illumina NextSeq 500 platform (Illumina Inc., San Diego, CA, USA). The sequences
176 are deposited at Dryad (Zimmermann et al., 2020a, 2020b).

177

178 2.5 Bioinformatic processing

179 The sequence reads were processed, filtered and assigned a taxonomic name according to
180 the NCBI taxonomy using the OBITools package (Boyer et al., 2016) with the same bioinformatics
181 parameter settings as described in (Dulias et al., 2017). Taxonomic assignment was applied via the
182 EMBL nucleotide reference database (EMBL release 138 from November 2018 (Kanz et al.,
183 2005)) using a least common ancestor approach implemented in OBITools (Table S1). The
184 resulting table was combined with samples sequenced on a previous run (Zimmermann et al.,
185 2020). Then, further denoising was carried out on the combined dataset using R v. 3.6.0 (R Core
186 Team, 2018): PCR and sequencing errors are known to inflate diversity estimates, especially with
187 ancient DNA, hence we kept only those amplicon sequence variants (ASVs) that (1) were assigned
188 a taxonomic name based on 90-100% similarity to an entry in the reference database, (2) were
189 represented with at least 100 read counts in total and (3) at least 10 read counts per PCR-product,
190 (4) were present at least 3 times among all PCR-products, (5) showed taxonomic resolution at least
191 to phylum level “Bacillariophyta”, and (6) were tagged as “internal” by obiclean in less than 50%
192 of the different replicates per sample. After filtering, the PCR replicates of a sample were
193 combined and subjected to rarefaction. We resampled the data based on the minimum number of
194 sequences (8,882 counts) using a custom R script (https://github.com/StefanKruse/R_Rarefaction;
195 Kruse, 2019).

196 The negative controls were mostly clean (Table S2). They contained, on average, 38
197 sequence variants of which 90.4% occurred with only one or two read counts, which can most
198 likely be attributed to tag jumps (Schnell et al., 2015). Only one PCR-product of an extraction
199 negative control contained more reads, which was probably a pipetting mistake during PCR-setup,
200 because the corresponding PCR negative control was clean and the other 2 PCR replicates of this
201 negative control were clean as well.

202

203 2.6 Statistical analysis

204 2.6.1 Composition

205 Constrained hierarchical clustering (CONISS; Grimm, 1987) was applied in which clusters
206 were constrained stratigraphically by sample depth with the *chclust*-function from the R package
207 “rioja” v. 0.9-21 (Juggins, 2012). Subsequently we used the *bstick*-function from the “vegan”
208 package v. 2.5-6 (Oksanen et al., 2011) to compare the dispersion of the computed classification
209 against the dispersion of a broken stick model in order to assess the number of significant zones.
210 The stratigraphic diagram was plotted using *strat.plot* from “rioja” and based only on ASVs having
211 proportions of at least 1% in the dataset.

212

213 2.6.2 Environmental variables

214 Environmental variables were retrieved from PANGAEA (Max et al., 2012b, 2012a;
215 Meyer et al., 2016a; Riethdorf et al., 2013b), NGRIP 20-year means of $\delta^{18}\text{O}$
216 (http://www.iceandclimate.nbi.ku.dk/data/2010-11-19_GICC05modelext_for_NGRIP.txt) and
217 June insolation was calculated after Laskar et al. (2004). Environmental variables were
218 interpolated using the methods described in Reschke et al. (2019). First, the environmental data
219 were transformed using the function *zoo* from the “zoo” package v. 1.8-7 and used in the function
220 *CorIrregTimser* using the package “corit” v. 0.0.0.9000
221 (<https://github.com/EarthSystemDiagnostics/corit>). As environmental data were limited, we
222 focused the multivariate analyses (package “vegan”) on the temporal interval from 7.56–16.48 cal
223 kyr BP. ASVs with proportions of at least 1.5% were selected for multivariate statistical analysis.

224 To avoid double-zeros being regarded as similar between samples we used Hellinger
225 transformation on the reduced proportion-based community matrix with decostand from “vegan”.

226 To test for linear dependencies of the environmental variables we ran a constrained
227 correspondence analysis (CCA) and calculated the variance inflation factor (VIF). Furthermore,
228 we applied forward selection using ordistep for stepwise extension of the regression model and
229 performed a permutation test to keep variables that significantly explain some of the variance of
230 the community matrix. Subsequently, we tested the chosen variables with conditional variables for
231 their unique explained variance as forward selection depends on the order by which the variables
232 were put into the model.

233

234 2.6.3 Taxonomic richness and beta-diversity

235 Richness, the number of ASVs, was calculated as a measure of taxonomic alpha-diversity
236 using the *estimateD* function from the iNEXT package v. 2.0.20 (Hsieh et al., 2016), which
237 allowed rarefaction analysis for both centric and pennate diatom ASVs in parallel. Rarefied
238 richness was tested for correlations with interpolated environmental variables using *rcorr* from the
239 package “Hmisc” v. 4.4-0 (Harrell Jr, 2020) with the method “Pearson”. Taxonomic beta-diversity
240 was calculated using the *betapart.core* and *betapart.pair* functions of the “betapart” v. 1.5.1 package
241 (Baselga & Orme, 2012), which allowed the split of pair-wise Jaccard dissimilarities into
242 nestedness (sample composition is a subset of the previous sample composition) and turnover
243 fractions (taxonomic replacement from one sample to the next). All analyses were carried out using
244 R v. 3.6.0 (R Core Team, 2018).

245

246 **3 Results**

247 A total of 13,584,296 reads were assigned to 3,038 amplicon sequence variants (ASVs) at
248 90–100% similarity to reference sequences. Of these, 2,191 ASVs were specific for diatoms and
249 comprised 11,746,154 reads. After further filtering, 11,114,776 reads remained assigned to 232
250 ASVs, which amounts to 94.6% of all reads assigned to diatoms. Of these, 47% are resolved to
251 species level and 38% to genus level. The lowest number of diatom read counts (8,882 counts)

252 was detected in the sample at 7.885 m depth (18.0 cal kyr BP) while the highest number (914,117
253 counts) was detected in the sample at 3.0 m (11.1 cal kyr BP). The majority of diatom-derived
254 reads were assigned to centric diatoms (92.7%), while 7.3% of reads were assigned to pennate
255 diatoms. The sequence variants were assigned to 25 different families, of which Bacillariaceae (68
256 ASVs), Chaetocerotaceae (40 ASVs), Naviculaceae (18 ASVs) and Thalassiosiraceae (54 ASVs)
257 encompassed the majority of the 231 ASVs that remained after filtering and rarefaction.

258 3.1 Diatom *seda*DNA composition

259 Throughout the past 19.9 cal kyr BP the *seda*DNA composition of the 63 samples is
260 dominated by ASVs assigned to cold-water and sea ice-associated species, such as *Bacterosira*
261 sp., *Nitzschia* (cf.) *frigida*, *Porosira* sp., *Thalassiosira antarctica* and *T. nordenskioeldii*.
262 Dominant ASVs are assigned to centric diatoms of the genera *Chaetoceros*, *Thalassiosira*,
263 *Porosira* and *Skeletonema*.

264 The proportions of ASVs assigned to pennate diatoms per sample ranges between a
265 minimum of 1.4% (2.36 cal kyr BP) and a maximum of 15.1% (19.9 cal kyr BP). Pennate diatoms,
266 predominantly pennate, raphid diatoms, have highest proportions during the end of the LGM
267 (10.9%) and early deglacial (Heinrich Stadial 1 (HS1) (9.8%) and Bølling/Allerød (9%)). Their
268 mean proportions decrease continuously until the Mid Holocene (4.7%), but increase again in the
269 Late Holocene (5.5%) (Fig. 2).

270 Constrained hierarchical clustering combined with the broken stick model suggested
271 division into three stratigraphic zones that are consistent with typical climatic phases: the late
272 glacial phase comprising samples dated to 19.91–15.26 cal kyr BP, the deglacial transition phase
273 comprising samples dated to 14.95–10.73 cal kyr BP and the Holocene phase comprising samples
274 dated to 10.43–1.08 cal kyr BP. We therefore used these phases for the description of the *seda*DNA
275 record.

276 The glacial phase comprises several ASVs assigned to *Thalassiosira* which account for
277 about 30–70% of the composition, *Porosira* (9.6–29.2%), *Chaetoceros* cf. *contortus* 1 SEH 2013
278 ASV 980 (8.7–9.6%), *Attheya* ASV 58 (0.4–6.8%), *Actinocyclus* 1 MPA-2013 (0.9–5.1 %), and
279 *Chaetoceros socialis* ASV 1280 (0.8–4.7%). Among pennate diatoms, ASVs assigned to

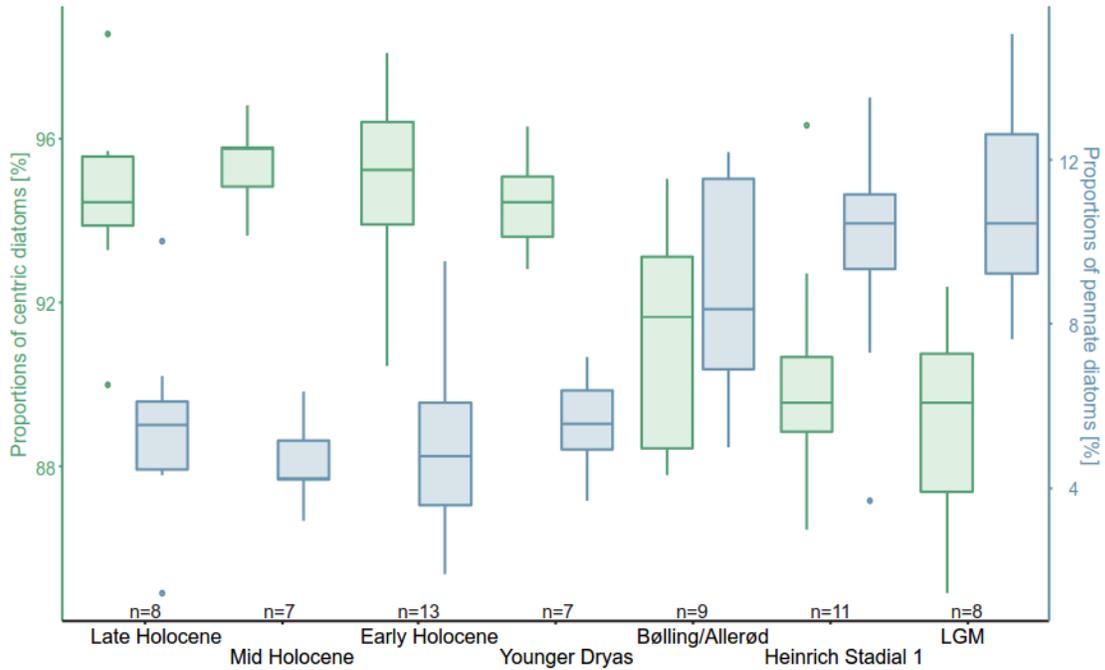
280 *Bacillaria* sp. ASV 139 (0.2–2.7%), *Navicula ramosissima* ASV 1651 (0–2.5%), and several
281 *Nitzschia* ASVs have notable proportions.

282 Although the deglacial phase encompasses climatically very different phases,
283 Cymatosiraceae (0.02–0.7%) and *Leptocylindrus minimus* (0–1.4%) ASVs are mostly restricted to
284 this zone. With the onset of this zone the proportions of most pennate diatoms as well as
285 *Thalassiosira nordenskiöldii* ASV 2697 (mostly <0.1%) decrease. The Bølling/Allerød phase
286 contains ASVs that have their peak proportions during this phase such as *Paralia* (0.6–16.8%) and
287 *Thalassiosira* sp. 15BOF (0–1.2%). While *Thalassiosira angulata* (0.3–5.2%) and *Nitzschia* cf.
288 *frigida* ASVs 1763 and 1764 (1.3–4.8%) increase, *Chaetoceros socialis* ASV 1280 (0–1.4%) and
289 *Minidiscus trioculatus* (0–0.8%) show a marked drop during this phase. During the Younger
290 Dryas, some ASVs that decreased during the Bølling/Allerød, increase again, such as *Chaetoceros*
291 *socialis* ASV 1280 (3–5%), *Thalassiosira* sp. ASV 2241 (2.6–7.4%) and *Attheya* ASV 58 (0–
292 5.7%).

293 The Holocene phase is marked by the reduction and sporadic presence of several ASVs
294 assigned to pennate diatoms, yet increased but highly variable proportions of *Haslea avium* (0.02–
295 4%). Predominantly, the Holocene is composed of sequence types assigned to the genera

296 *Chaetoceros* and *Thalassiosira* as well as *Porosira* (4.4–33.9%) and *Skeletonema* ASV 2155 (0.3-
 297 10.2%).

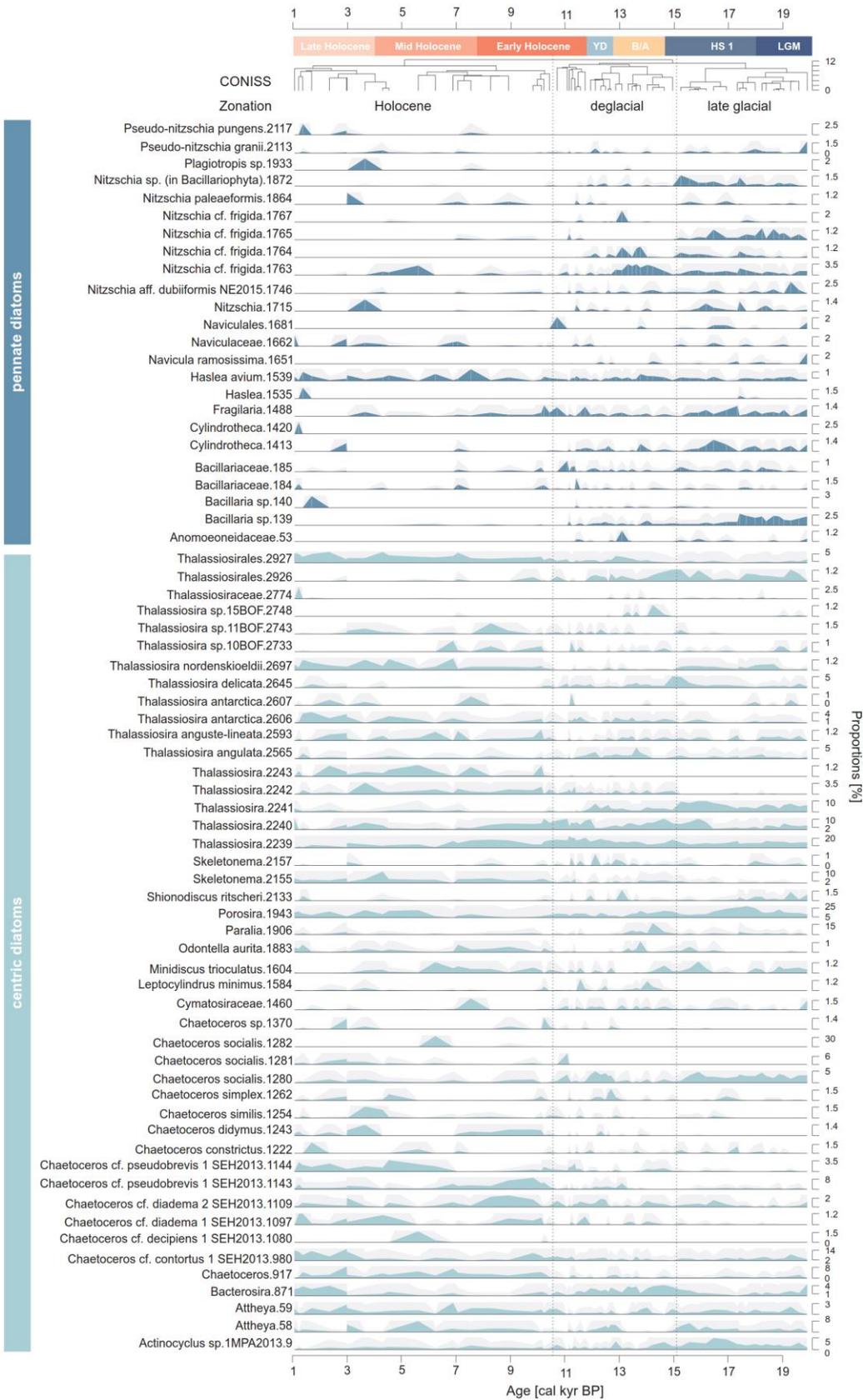
298



299

300 **Figure 2:** Proportions of amplicon sequence variants assigned to centric diatoms (red) and
 301 pennate diatoms (blue) during the glacial/interglacial transition over of the past 19.9 cal kyr BP.

302



304 **Figure 3:** Stratigraphic diagram showing the proportions of the assigned amplicon sequence
 305 variants for each sample through time. Only sequence variants with more than 1% proportion in
 306 one or more samples are shown. If not indicated otherwise, the scales of proportions start with 0%.
 307 Centric diatoms are marked in light blue, while pennate diatoms are marked in dark blue, 5x
 308 exaggeration is shown in gray. The horizontal, dotted lines mark the boundaries of the three
 309 CONISS based stratigraphic zones. The CONISS dendrogram and the total sum of squares are
 310 given on the right side.

311

312 3.2 Relationship of diatom *sed*aDNA composition with environmental variables

313 We used multivariate statistics to determine to what degree abiotic factors have an effect
 314 on the *sed*aDNA composition. Based on variance inflation factors (VIFs) exceeding 10 and thus
 315 collinearity with other variables, we stepwise excluded UK'₃₇-based SSTs, NGRIP $\delta^{18}\text{O}$ and the
 316 BIT-index (branched and isoprenoid tetraether index, measuring the relative input of terrestrial
 317 and marine glycerol dialkyl glycerol tetraethers (GDGTs)) from the set of variables. June
 318 insolation, the sea-ice proxy IP₂₅ and foraminiferal, ice-volume corrected $\delta^{18}\text{O}$ (salinity proxy)
 319 displayed VIF values between 1.003 and 1.009 suggesting low levels of collinearity between each
 320 other. Decomposition of total variance (0.09383) showed that 33.9% of the variance is explained
 321 by the model containing the three explanatory variables (adjusted $R^2=0.321$). RDA axis 1 captures
 322 22.6% of the variance, while RDA axis 2 captures 11.3%. June insolation is highly correlated with
 323 RDA axis 1 (Table 1). Samples located in the right quadrants of the triplot are characterized by
 324 high June insolation and mostly centric diatoms of the genera *Chaetoceros*, *Thalassiosira* and
 325 *Skeletonema*. While the upper left quadrant is characterized by high concentrations of the sea-ice
 326 biomarker IP₂₅, the lower left quadrant is characterized by heavier $\delta^{18}\text{O}_{\text{ivc-sw}}$. Especially in the
 327 genus *Chaetoceros*, our dataset shows several ASVs assigned to the same species. The RDA triplot
 328 shows, for example, that *Chaetoceros socialis* ASV 1280 is located in the upper left quadrant
 329 which is associated with late glacial samples and IP₂₅ whereas *Chaetoceros socialis* ASV 1281 is
 330 located in the lower right quadrant which is associated with Early Holocene samples of the
 331 deglacial stratigraphic zone and high June insolation (Fig. 4). A second example is shown in the
 332 RDA triplot for two ASVs assigned to *Chaetoceros* cf. *pseudobrevis* 1 SHE-2013 with closest

333 match to the same GenBank accession number (KC985654). While ASV 1143 (98% match with
334 reference) is located in the upper right quadrant and has its highest proportions during the Early
335 Holocene and is strongly reduced since the Mid Holocene (Fig. 3), the proportions of ASV 1144

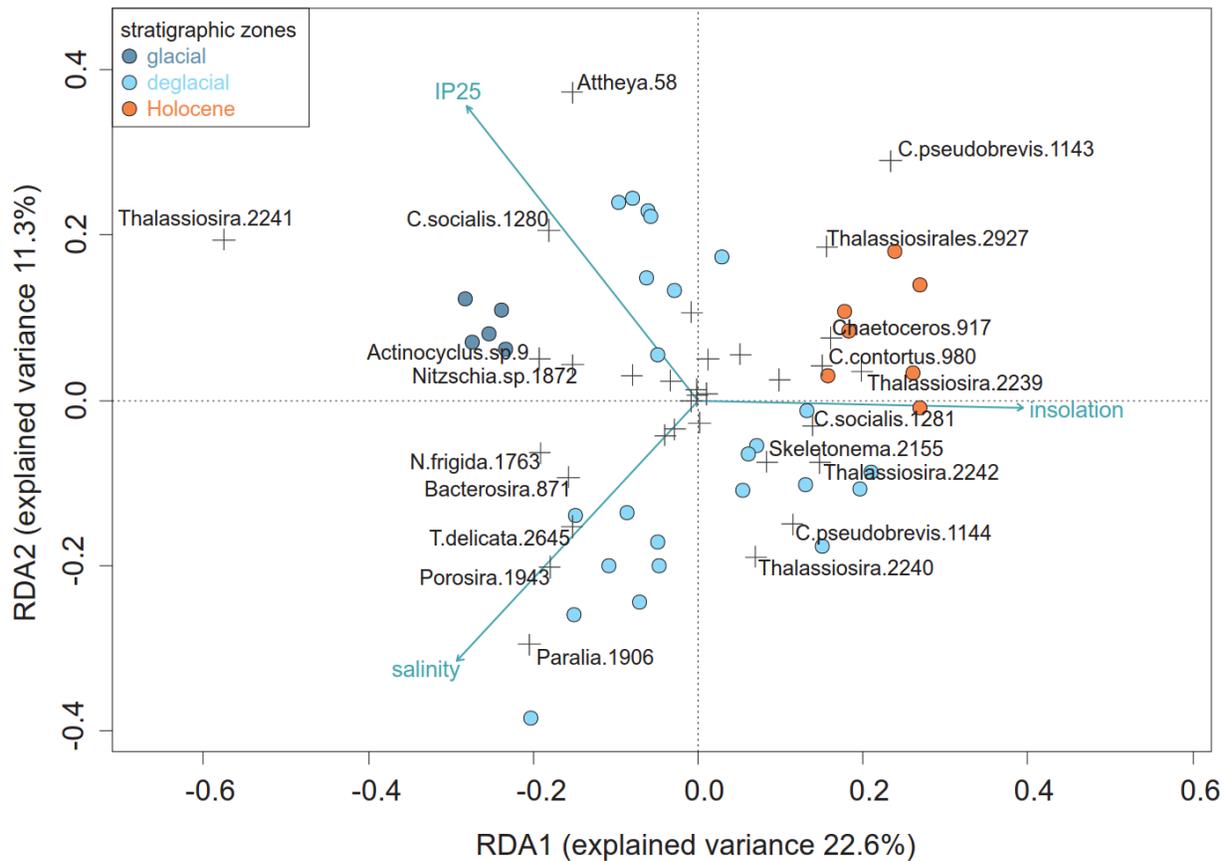
336 (100% match with reference) increase in the early Mid Holocene and remain on a rather similar
 337 level until the Late Holocene.

338

339 **Table 1:** Variation partitioning for the final set of explanatory variables with adjusted R^2 and p-
 340 values for the single and the individual fractions (conditional variance) which remain after
 341 eliminating the effects of the remaining explanatory variables.

	variance	single		conditional	
		adj. R^2	p-value	adj. R^2	p-value
June insolation	0.0116	0.108	0.002	0.040	0.001
IP ₂₅	0.0119	0.104	0.002	0.089	0.001
salinity	0.0113	0.099	0.001	0.023	0.01

342



343

344 **Figure 4:** RDA triplot showing the abbreviated names of the 20 most explaining amplicon
 345 sequence variants (ASVs) and the vector ends of the most abundant ASVs as a gray plus symbol
 346 (+). Samples are marked by colour coded points to the three stratigraphic zones which they
 347 represent (dark blue = late glacial, light blue = deglacial, orange = Holocene). Environmental
 348 variables are shown by blue vectors.

349

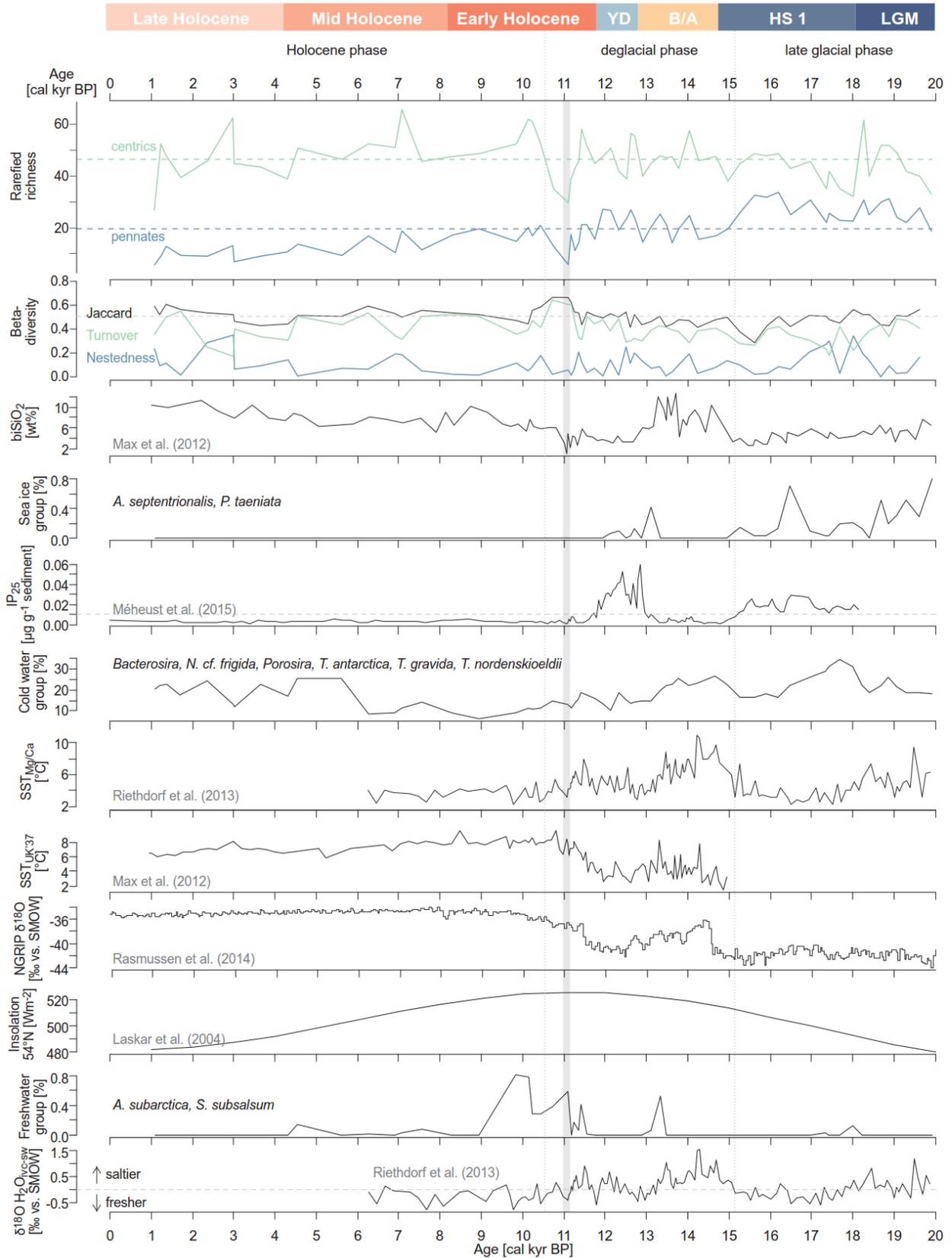
350 3.3 Diatom diversity changes

351 Richness (number of ASVs after rarefaction) was lowest in the youngest sample dated to
 352 1.08 cal kyr BP (37 ASVs) and highest about 18.42 cal kyr BP (106 ASVs) (Fig. 5, Fig. S1). While
 353 the median richness among sequences assigned to centric diatoms varies only slightly over the past
 354 19.9 cal kyr BP, the richness of those assigned to pennate diatoms declines almost continuously
 355 throughout the record, except for an increase during the Younger Dryas (Fig. S1). The richness of

356 pennate diatom ASVs shows a moderate, positive correlation with IP₂₅ (R=0.48, p=0.003), and
357 moderate, negative correlations with June insolation (R=-0.49, p=0.002) and NGRIP $\delta^{18}\text{O}$ (R=-
358 0.68, p<0.001). The richness of centric diatoms does not show any significant correlation with
359 environmental variables Table S1).

360 Beta-diversity, which is here used as a measure of dissimilarity between adjacent samples,
361 is moderate overall (median=0.51). There are four strong declines in diatom richness in our record.
362 The first occurred between 18.24 and 18 cal kyr BP and reduced centric diatom ASVs
363 approximately by half (from 62 to 32 ASVs) and pennate ASVs by about a quarter (from 31 to 23
364 ASVs). Slightly elevated values of beta-diversity (Jaccard dissimilarity=0.56) during this time in
365 comparison to the median Jaccard dissimilarity correspond more to nestedness rather than turnover
366 (Fig. 3). The second decline occurred between 15.6 and 14.24 cal kyr BP and affected pennate
367 diatom ASVs (from 33 to 16 ASVs) more than centric ones (from 49 to 38 ASVs). Beta-diversity
368 increases from 15.6 to 14.95 cal kyr BP and then slightly decreases again. The third decline affects
369 both pennate and centric diatoms strongly and started after 11.95 until 11.1 cal kyr BP for pennate
370 (from 27 to 6 ASVs) and after 11.42 until 11.1 cal kyr BP for centric diatom ASVs (from 58 to 30
371 ASVs). Between 11.1 and 10.5 cal kyr BP, the turnover partition of beta-diversity is highest overall
372 throughout the record. This corresponds predominantly to a loss of ASVs in both pennate and
373 centric diatoms at 11.1 cal kyr BP followed by a gain of ASVs at 10.5 cal kyr BP (Fig.5). The
374 fourth decline is detected between the youngest samples from 1.22 and 1.08 cal kyr BP (from 9 to
375 6 pennate ASVs and from 53 to 27 centric ASVs) and shows slightly increased beta-diversity, of
376 which the turnover partition is only slightly higher than the nestedness one.

377



379 **Figure 5:** Comparison of rarefied richness (horizontal green dashed line marks median richness
380 of centric diatom ASVs, blue dashed line marks median richness of pennate diatom ASVs), beta-
381 diversity (Jaccard dissimilarity and its partitions into turnover and nestedness) and proportions of
382 diatom *sedaDNA* indicator groups with Mg/Ca-based subsurface SSTs (Riethdorf et al., 2013a),
383 alkenone-based surface SST_{UK'37}, biogenic silica (Max et al., 2012), IP₂₅ (Méheust et al., 2015),
384 stable oxygen isotopes from Greenland ice cores (NGRIP $\delta^{18}\text{O}$; Rasmussen et al., 2014), June
385 insolation calculated after Laskar et al. (2004) and sea subsurface salinity approximation ($\delta^{18}\text{O}_{\text{ivc-}}$
386 sw ; Riethdorf et al., 2013a). The vertical dotted lines mark the stratigraphic zonation. The shaded
387 vertical area highlights the sample at ~11.1 cal kyr BP.

388

389 4 Discussion

390 4.1 General relationship between diatom *sedaDNA* composition and environmental
391 change

392 4.1.1 Late glacial phase (19.9–15.26 cal kyr BP)

393 The diatom *sedaDNA* composition during the late glacial phase reflects sea ice as a key
394 variable shaping the past diatom communities. The presence of winter sea ice is indicated by
395 sequences assigned to sea ice-associated species such as the colony-forming pennate diatoms
396 *Pauliella taeniata* (Grunow) Round & Basson (Lovejoy et al., 2002; Syvertsen, 1991) and
397 *Nitzschia cf. frigida* (Grunow) (Hasle & Heimdal, 1998), or the epiphytic living *Attheya*
398 *septentrionalis* (Østrup) R.M.Crawford (von Quillfeldt, 1997). These species are frequently
399 detected in modern surveys of sea ice (Limoges et al., 2018; Lovejoy et al., 2002; Poulin et al.,
400 2011; von Quillfeldt, 2000), but can be absent or only sporadically found in surface sediments due
401 to degradation during sinking through the water column (Limoges et al., 2018; Lopes et al., 2006).
402 Here, the *sedaDNA* has an advantage over morphological data and can complement the picture we
403 perceive of the past. Winter sea-ice conditions with ice-free summers were also reconstructed
404 based on the sea-ice biomarker IP₂₅ and abundant *Fragilariopsis cylindrus* and *F. oceanica* in local
405 microfossil assemblages (Max et al., 2012; Méheust et al., 2016; Smirnova et al., 2015). Low
406 proportions of sequences assigned to the productive genus *Chaetoceros* (Lopes et al., 2006) in our

407 record could reflect overall low productivity as also suggested by low biogenic silica (Max et al.,
408 2012) and low abundance of diatom microfossils during this phase (Smirnova et al., 2015).

409 The high share of sequences assigned to *Bacterosira* (probably *B. bathyomphala*),
410 *Thalassiosira nordenskiöldii*, *T. antarctica* and *Porosira* (probably *P. glacialis*) points to ice-
411 free, but cool, summers. These cold-water species are often found along the winter ice edge in the
412 Bering and Okhotsk Seas and in coastal areas of Kamchatka (Ren et al., 2014), although the neritic
413 species *Bacterosira bathyomphala* could also be associated with shallow slope depths down to 500
414 m in the Sea of Okhotsk (Ren et al., 2014; Sancetta, 1982). In the subarctic North Pacific and
415 adjacent seas, these species achieve maximum abundances where summer SSTs remain between
416 5 to 8 °C, but they can generally be found where summer SSTs remain below 12.5 °C (Ren et al.,
417 2014). This is well within the range of previous reconstructions carried out on this sediment core
418 indicating summer SSTs of about 8 to 9.5 °C during this phase (Meyer et al., 2016b). Late glacial
419 diatom microfossil data of this core are characterized by dominance of the *Thalassiosira trifulta*
420 and *Actinocyclus curvatulus* group. While our *sedaDNA* reveals much more complexity in the
421 diatom assemblages during this time, our method is still restricted by the incompleteness of the
422 reference database, as, for example, *T. trifulta* and *A. curvatulus* are not available and are
423 potentially only resolved to genus level in our record. Still, *Actinocyclus* sp. 1 MPA-2013 displays
424 higher proportions during the late glacial phase in comparison to the deglacial and Holocene
425 phases, which is in agreement with Smirnova et al. (2015) as well as with microfossil records from
426 Bowers Ridge and the western subarctic gyre (Katsuki & Takahashi, 2005).

427 Sequences assigned to pennate, raphid diatoms have the highest proportions until
428 approximately 15.7 cal kyr BP, which may be linked to bottom ice communities (Caissie et al.,
429 2010). Even though pennate diatoms are often poorly preserved as microfossils in sediments
430 (Limoges et al., 2018), they are usually dominant in sea ice (Poulin et al., 2011; von Quillfeldt,
431 2000). The benthic lifestyle, to which pennate diatoms are adapted to, includes the excretion of
432 exopolymers via the raphe, which facilitate their attachment to surfaces and enables them to
433 actively move along surfaces such as ice (Olsen et al., 2019; Round et al., 1990). As summer
434 insolation is an important driver for SSTs in the subarctic North Pacific (Harada et al., 2014), it is
435 conceivable that the low summer insolation during this time (Laskar et al., 2004) promoted pennate
436 diatoms due to a long duration of sea-ice coverage with only a short summer. Short-term summers
437 have also been reconstructed during this phase in the Bering Sea where sea ice was estimated to

438 have covered most of the Bering Sea for more than six months (Caissie et al., 2010; Méheust et
439 al., 2016).

440

441 4.1.2 Deglacial transition phase (14.95–10.73 cal kyr BP)

442 The *sed*aDNA composition of samples dated to the Bølling/Allerød phase (14.7–12.7 cal
443 kyr BP) can likely be related to increased local subsurface salinity conditions in comparison to the
444 glacial phase as implied by an enrichment of foraminifera-derived stable oxygen isotopes
445 ($\delta^{18}\text{O}_{\text{ivc-sw}}$ (Riethdorf et al., 2013a)). Peak proportions of sequences assigned to *Paralia* (*sulcata*)
446 up to ~15% coincide with increasing subsurface salinities (positive $\delta^{18}\text{O}_{\text{ivc-sw}}$) during this phase.
447 *Paralia sulcata* (Ehrenberg) Cleve is a coastal species, which is a common part of benthic
448 communities and increased abundance is often recorded in areas of upwelling (Abrantes, 1988;
449 McQuoid & Nordberg, 2003; Stabell, 1986), or where the water column is less stable and subject
450 to strong wind-driven or tidal mixing (McQuoid & Nordberg, 2003).

451 Sequences assigned to *Thalassiosira delicata*, the cold-water indicator *Porosira*, and
452 *Bacterosira*, but also to the sea-ice associated *Nitzschia* cf. *frigida* suggest a cold-water
453 environment with frequent influence of sea ice despite recorded increases of SSTs by about 3–
454 5 °C (Max, Riethdorf, Tiedemann, Smirnova, Lembke-Jene, et al., 2012; Riethdorf et al., 2013a).
455 Such SSTs are still in the growth range of *B. bathyomphala* and *P. glacialis* which occur in the
456 Bering and Okhotsk Seas in regions with summer SSTs between 5 and 12.5 °C (Ren et al., 2014).
457 These species also occur in the marginal ice zone, where sea-ice concentration varies between 15
458 and 80%, and achieve high abundances during the spring bloom (Hasle, 1990). Hence, proportions
459 of sequences assigned to *Nitzschia* cf. *frigida* of more than 3% in this phase are not necessarily
460 contradictory. While the absence of IP₂₅ suggests ice-free conditions, gravel-sized ice rafted debris
461 at about 13.86 cal kyr BP (560 cm depth; Levitan et al., 2015) point towards the presence of sea
462 ice. As *Nitzschia frigida* is a common inhabitant of landfast ice communities (Medlin & Hasle,
463 1990), a possible explanation could be the occurrence of drifting ice floes, which were transported
464 away from the coast or southward by the East Kamchatka Current, and probably originated from
465 enclosed bights and bays along the eastern coast of Kamchatka (Polyakova, 2007).

466 The diatom *sed*aDNA composition of samples dated to the Younger Dryas (12.7–11.7 cal
467 kyr BP) are characterized by the presence of sea ice-associated ASVs (*Pauliella taeniata*, *Attheya*

468 *septentrionalis*, *Nitzschia* cf. *frigida*) suggesting winter sea-ice coverage during this time. Sea ice-
469 associated ASVs in combination with a higher richness of pennate diatom ASVs in comparison to
470 the Bølling/Allerød phase are in agreement with sea-ice diatom microfossils detected by Smirnova
471 et al. (2015), high concentrations of the sea-ice biomarker IP₂₅ (Méheust et al., 2016) and
472 reconstructed subsurface SSTs between about 2–5 °C with lower seasonal contrasts and a reduced
473 stratification of the mixed layer (Riethdorf et al., 2013a).

474 Increased proportions of *Chaetoceros socialis* (ASV 1280) are characteristic in this phase
475 and show comparable proportions with regard to the late glacial phase. This matches increasing
476 proportions of *Chaetoceros* spp. detected in the microfossil record during this phase (Smirnova et
477 al., 2015). *Chaetoceros socialis* is often dominant in coastal areas with sparse to no sea ice in the
478 Greenland North Water Polynya, where it blooms in late summer; however, it can also achieve
479 substantial abundances in sea ice (Booth et al., 2002; von Quillfeldt, 1997). We therefore assume
480 that this ASV is derived from a lineage adapted to sea ice or marginal ice zone conditions and
481 might actually represent the novel described *C. gelidus* Chamnansinp, Y.Li, Lundholm &
482 Moestrup (Chamnansinp et al., 2013).

483 The composition of samples dated to the onset of the Preboreal (11.7–10.73 cal kyr BP)
484 are marked by a decrease in sequences assigned to cold-water diatoms and highest proportions of
485 sequences assigned to freshwater species throughout the record. From about 11.2 cal kyr BP
486 onwards, sequences assigned to *Skeletonema subsalsum* (Cleve-Euler) Bethge and *Aulacoseira*
487 *subarctica* (Otto Müller) E.Y.Haworth increase strongly. *Skeletonema subsalsum* can be found in
488 rivers, estuaries and coastal areas (Hasle & Evensen, 1975) while *A. subarctica* is a freshwater
489 species occurring in rivers and turbulent, cool lakes on Kamchatka (Lepsкая et al., 2010). Their
490 increased proportions in the *seDaDNA* record suggest an increased runoff from Kamchatka and
491 possibly fresher sea-surface conditions. The timing coincides with the post-glacial opening of the
492 Bering Strait (Jakobsson et al., 2017) and is marked by a high June insolation (Laskar et al., 2004).
493 Fresher subsurface conditions are also indicated by peaks of lighter $\delta^{18}\text{O}_{\text{IVC-SW}}$ in this core
494 (Gorbarenko et al., 2019; Riethdorf et al., 2013a). Even though terrestrial lipid biomarkers and
495 titanium/calcium records of this core suggest low terrestrial input, the sum of terrestrial soil derived
496 branched GDGTs peaks during this time (Meyer et al., 2016b, 2017) and might also point towards
497 increased runoff from Kamchatka.

498

499 4.1.3 Holocene phase (10.43–1.08 cal kyr BP)

500 The *sedDNA* composition of the Early to Mid Holocene is characterized by reduced
501 proportions of sequences assigned to cold-water species and correlate with high June insolation
502 between ~11 and 6 cal kyr BP (Laskar et al., 2004). The timing fits well into the range of the
503 Holocene Thermal Maximum between ~10 and 8 cal kyr BP (Meyer et al., 2017), with maximum
504 SSTs ranging from 6–11 °C (SST_{UK'37}) as reconstructed by (Max et al., 2012) to 10–11 °C
505 (SST_{TEXL86} August temperatures of Meyer et al., 2016b). During this phase, the proportions of
506 sequences assigned to centric diatoms, especially of the genera *Chaetoceros*, *Thalassiosira* and
507 *Skeletonema*, increase at the expense of sequences assigned to pennate diatoms.

508 High proportions of *Chaetoceros* are indicative of high productivity, which is supported
509 by increasing biogenic opal (Max et al., 2012; Riethdorf et al., 2013a) and high diatom microfossil
510 abundances (Smirnova et al., 2015). Sequences assigned as freshwater diatoms and negative
511 $\delta^{18}\text{O}_{\text{ivc-sw}}$ values indicate long-term relatively fresh subsurface conditions (Riethdorf et al., 2013a).

512 From about 6–1 cal kyr BP, increasing proportions of cold-water diatom ASVs reflect
513 cooling, which is in agreement with the climate development recorded in the NGRIP Greenland
514 ice core record (Rasmussen et al., 2014), reconstructed SSTs (Max et al., 2012; Meyer et al., 2016b,
515 2017; Riethdorf et al., 2013a) and the advance of glaciers on Kamchatka (between 4.5 and 3.5 cal
516 kyr BP) (Brooks et al., 2015). A drop in cold-water diatoms at ~3 cal kyr BP, suggests a short-
517 term interruption of this phase of cooling, which is synchronous with a short-term increase of
518 SST_{UK'37} (Max et al., 2012).

519 An increase of sequences assigned to pennate diatoms in the Late Holocene and high
520 proportions of cold-water diatom ASVs including the ice-associated *Nitzschia* cf. *frigida* suggest
521 cooling and potentially an increasing influence of winter sea ice or at least drifting ice floes in the
522 region. In support of this, grain-size analyses of this core suggest increased presence of potentially
523 ice-rafted material (Levitan et al., 2015) and after 6 cal kyr BP the microfossil assemblage contains
524 sea ice-associated species (Smirnova et al., 2015), whereas the sea-ice biomarker IP₂₅ suggests ice-
525 free conditions (Méheust et al., 2016). Thus, *sedDNA* has the potential to complement our
526 understanding of past sea-ice distribution.

527 A second sequence variant of *Chaetoceros socialis* (ASV 1281) is linked with Holocene
528 samples and suggests that different ASV are potentially derived from different lineages adapted to
529 different growth conditions (Hamsher et al., 2013). In several cases, different ASVs which were
530 assigned to the same species co-occurred while a few showed patterns consistent with changes in
531 environmental conditions. Such co-occurrence of several ASVs could arise due to the degraded
532 state of ancient DNA introducing erroneous nucleotides during PCR (Hofreiter et al., 2001), but it
533 is also in agreement with the ecological hypothesis that genetic variation allows species to quickly
534 adapt to changing environmental conditions (Godhe & Rynearson, 2017). The application of
535 ASVs, in comparison to clustering into operational taxonomic units, thus has the advantage of
536 revealing (pseudo-)cryptic species or ecotypes and should be considered more in biogeographic
537 surveys.

538

539 4.2 Changes in diatom *seada*DNA richness and the relationship with environmental 540 changes

541 Richness of pennate diatom ASVs was higher during phases of strong Northern
542 Hemisphere cooling, namely the late LGM, Heinrich Stadial 1 and the Younger Dryas and is
543 significantly positively correlated with sea ice and significantly negatively correlated with June
544 insolation and Northern Hemisphere temperatures. This points towards sea ice as a facilitating
545 factor for pennate diatoms with regard to alpha-diversity in the subarctic NW Pacific. The negative
546 correlation of pennate diatom richness with June insolation and temperatures in comparison to
547 centric diatom richness might be related to the higher sensitivity of pennate diatoms to ultraviolet
548 light in the absence of a thick sea-ice cover, whereas centric diatoms have developed strategies to
549 prevent damage to their photosystems (Enberg et al., 2015).

550 A strong decline of richness accompanied by increased beta-diversity occurred during the
551 Preboreal phase of the Early Holocene around 11.1 cal kyr BP and could be attributed
552 predominantly to turnover by loss of ASVs until 11.1 cal kyr BP and replacement by different
553 ASVs towards 10.5 cal kyr BP. This decline of richness occurs simultaneously with a sharp drop
554 in biogenic opal, suggesting strongly reduced productivity of siliceous organisms such as diatoms
555 (Max, Riethdorf, Tiedemann, Smirnova, Lembke-Jene, et al., 2012). This interval is characterized
556 by a peak in summer insolation, rising SSTs (Max et al., 2012) and strong sea-level rise which

557 culminated in the opening of the Bering Strait (Jakobsson et al., 2017). A sharp increase in
558 proportions of the freshwater group and foraminiferal stable oxygen isotopes points towards
559 increased sea-surface freshening, potentially derived from increased runoff from Kamchatka
560 (Gorbarenko et al., 2019; Riethdorf et al., 2013a) as suggested by a peak in terrestrial lipid
561 biomarkers around 11.2 cal kyr BP (Meyer et al., 2016b, 2017). Fresher surface conditions could
562 have resulted in enhanced vertical stratification limiting the mixing of nutrients from deeper water
563 masses toward the euphotic zone and thus restricting primary productivity, manifested as a sharp
564 drop of biogenic silica during this time (Max et al., 2012).

565 To what extent changes in the diversity and composition of past diatom assemblages
566 affected the past organismal community at the ecosystem level remains to be revealed. Diatoms
567 have been shown to impact marine plankton structure by biotic interactions, for example by
568 competition or selective grazing (Vincent & Bowler, 2020) and exopolymers excreted by sea-ice
569 diatoms have been shown to be hotspots for enhanced bacterial activity (Meiners et al., 2008).
570 Hence, changes might be linked to several trophic levels and could substantially impact marine
571 food-webs and biogeochemical cycling in areas such as the Arctic where food-webs have evolved
572 over millions of years.

573

574 **5 Conclusions**

575 We present the first diatom-derived *sed*aDNA metabarcoding record of the subarctic NW
576 Pacific, which covers the past 19.9 cal kyr.

577 The dominant components of the diatom *sed*aDNA composition fit well within the
578 framework of reconstructed sea-ice dynamics, SSTs and subsurface salinities over the past
579 millennia. High proportions of sea-ice and cold-water associated diatoms and generally high
580 proportions of pennate diatoms point to sea ice as a main driver of diatom composition during the
581 late glacial phase as well as during the Younger Dryas. A positive correlation of pennate diatom
582 richness with the sea-ice biomarker IP₂₅ suggests that an extended winter sea-ice cover during the
583 late glacial and the Younger Dryas potentially acted as a diversifying force.

584 Substantial proportions of *Nitzschia cf. frigida* in the absence of the sea-ice proxy IP₂₅
585 during the early deglacial phase point to a continued influence of drifting sea ice. This suggests

586 either that the ecology of *N. frigida* is not completely understood yet, or that our proxy can
587 occasionally be more sensitive to the past presence of sea ice than IP₂₅.

588 Unfavorable conditions for pennate diatoms at the start of the Holocene is suggested by
589 decreasing proportions and the negative correlation of pennate diatom richness with June
590 insolation and temperature.

591 The inferred loss of 42% of diatom ASVs at ~11.1 cal kyr BP is accompanied by freshwater
592 diatoms implying the influence of runoff from Kamchatka to the local assemblage. It is likely that
593 the freshwater input resulted in enhanced vertical stratification limiting the mixing of nutrients
594 from deeper water masses toward the euphotic zone and thus restricting primary productivity – a
595 scenario which is supported by near-zero biogenic silica during this time. As modern climate
596 warming is anticipated to increase freshwater input into the Arctic from 11 to 30% by river
597 discharge alone (Nummelin et al., 2016), an enhanced stratification with restricted nutrient supply
598 could have tremendous consequences for the biological carbon pump, biogeochemical cycling and
599 the food-web in the Arctic.

600 Finally, our proxy revealed shifts of potential ecotypes of species from the genus
601 *Chaetoceros*, which is in agreement with the concept that intra-specific variation facilitates an
602 adaptive response to changing environmental conditions and is particularly relevant in high-
603 latitudinal ecosystems. Therefore, marine *seadaDNA* has the potential to answer ecological
604 questions regarding selective or adaptive responses at an intra-specific level where morphological
605 studies might be severely limited.

606

607 **Acknowledgments, Samples, and Data**

608 We declare that there is no conflict of interest.

609

610 Datasets for this research are available in: Dryad (Zimmermann et al., 2020a; Zimmermann
611 et al., 2020b). The rarefaction script is available at https://github.com/StefanKruse/R_Rarefaction
612 (Kruse, 2019).

613

614 We thank Sarah Olischläger and Iris Eder for support with the laboratory work, Lars Max
615 for providing the age-depth model, Cathy Jenks for English correction and the captain and crew of

616 the RV Sonne. The KALMAR Project (SO201-2- KALMAR Leg 2) was funded by the Federal
617 German Ministry of Education and Research (BMBF).

618

619 **References**

- 620 Abrantes, F. (1988). Diatom assemblages as upwelling indicators in surface sediments off Portugal. *Marine*
621 *Geology*, 85(1), 15–39. [https://doi.org/10.1016/0025-3227\(88\)90082-5](https://doi.org/10.1016/0025-3227(88)90082-5)
- 622 Baselga, A., & Orme, C. D. L. (2012). betapart: an R package for the study of beta diversity. *Methods in Ecology*
623 *and Evolution*, 3(5), 808–812. <https://doi.org/10.1111/j.2041-210X.2012.00224.x>
- 624 Binladen, J., Gilbert, M. T. P., Bollback, J. P., Panitz, F., Bendixen, C., Nielsen, R., & Willerslev, E. (2007). The
625 use of coded PCR primers enables high-throughput sequencing of multiple homolog amplification products
626 by 454 parallel sequencing. *PLoS ONE*, 2(2), e197. <https://doi.org/10.1371/journal.pone.0000197>
- 627 Booth, B. C., Larouche, P., Bélanger, S., Klein, B., Amiel, D., & Mei, Z.-P. (2002). Dynamics of *Chaetoceros*
628 *socialis* blooms in the North Water. *Deep Sea Research Part II: Topical Studies in Oceanography*, 49(22),
629 5003–5025. [https://doi.org/10.1016/S0967-0645\(02\)00175-3](https://doi.org/10.1016/S0967-0645(02)00175-3)
- 630 Boyer, F., Mercier, C., Bonin, A., Le Bras, Y., Taberlet, P., & Coissac, E. (2016). OBITools: a Unix-inspired
631 software package for DNA metabarcoding. *Molecular Ecology Resources*, 16, 176–182.
632 <https://doi.org/10.1111/1755-0998.12428>
- 633 Brooks, S. J., Diekmann, B., Jones, V. J., & Hammarlund, D. (2015). Holocene environmental change in
634 Kamchatka: A synopsis. *Global and Planetary Change*, 134, 166–174.
635 <https://doi.org/10.1016/j.gloplacha.2015.09.004>
- 636 Caissie, B. E., Brigham-Grette, J., Lawrence, K. T., Herbert, T. D., & Cook, M. S. (2010). Last Glacial Maximum to
637 Holocene sea surface conditions at Umnak Plateau, Bering Sea, as inferred from diatom, alkenone, and
638 stable isotope records. *Paleoceanography*, 25(1). <https://doi.org/10.1029/2008PA001671>
- 639 Chamnansinp, A., Li, Y., Lundholm, N., & Moestrup, Ø. (2013). Global diversity of two widespread, colony-
640 forming diatoms of the marine plankton, *Chaetoceros socialis* (syn. *C. radians*) and *Chaetoceros gelidus*
641 sp. nov. *Journal of Phycology*, 49(6), 1128–1141. <https://doi.org/10.1111/jpy.12121>

- 642 Corinaldesi, C., Beolchini, F., & Dell'anno, A. (2008). Damage and degradation rates of extracellular DNA in
643 marine sediments: implications for the preservation of gene sequences. *Molecular Ecology*, *17*(17), 3939–
644 3951. <https://doi.org/10.1111/j.1365-294X.2008.03880.x>
- 645 Coupel, P., Ruiz-Pino, D., Sicre, M. A., Chen, J. F., Lee, S. H., Schiffrine, N., et al. (2015). The impact of
646 freshening on phytoplankton production in the Pacific Arctic Ocean. *Progress in Oceanography*, *131*, 113–
647 125. <https://doi.org/10.1016/j.pocean.2014.12.003>
- 648 De Schepper, S., Ray, J. L., Skaar, K. S., Sadatzki, H., Ijaz, U. Z., Stein, R., & Larsen, A. (2019). The potential of
649 sedimentary ancient DNA for reconstructing past sea ice evolution. *The ISME Journal*.
650 <https://doi.org/10.1038/s41396-019-0457-1>
- 651 Degerlund, M., Huseby, S., Zingone, A., Sarno, D., & Landfald, B. (2012). Functional diversity in cryptic species of
652 *Chaetoceros socialis* Lauder (Bacillariophyceae). *Journal of Plankton Research*, *34*(5), 416–431.
653 <https://doi.org/10.1093/plankt/fbs004>
- 654 Dulias, K., Stoof-Leichsenring, K. R., Pestryakova, L. A., & Herzsuh, U. (2017). Sedimentary DNA versus
655 morphology in the analysis of diatom-environment relationships. *Journal of Paleolimnology*, *57*(1), 51–66.
656 <https://doi.org/10.1007/s10933-016-9926-y>
- 657 Enberg, S., Piiparinen, J., Majaneva, M., Vähätalo, A. V., Autio, R., & Rintala, J.-M. (2015). Solar PAR and UVR
658 modify the community composition and photosynthetic activity of sea ice algae. *FEMS Microbiology*
659 *Ecology*, *91*(10). <https://doi.org/10.1093/femsec/fiv102>
- 660 Epp, L. S., Zimmermann, H. H., & Stoof-Leichsenring, K. R. (2019). Sampling and Extraction of Ancient DNA
661 from Sediments. In B. Shapiro, A. Barlow, P. D. Heintzman, M. Hofreiter, J. L. A. Paijmans, & A. E. R.
662 Soares (Eds.), *Ancient DNA: Methods and Protocols* (pp. 31–44). New York, NY: Springer New York.
663 https://doi.org/10.1007/978-1-4939-9176-1_5
- 664 Fetterer, F., Knowles, K., Meier, W. N., Savoie, M., & Windnagel, A. K. (2017). Sea Ice Index, Version 3. Monthly
665 and daily GIS compatible shapefiles of median ice extent [Data set]. NSIDC: National Snow and Ice Data
666 Center. <https://doi.org/10.7265/n5k072f8>
- 667 Gebhardt, H., Sarnthein, M., Grootes, P. M., Kiefer, T., Kuehn, H., Schmieder, F., & Röhl, U. (2008). Paleonutrient
668 and productivity records from the subarctic North Pacific for Pleistocene glacial terminations I to V.
669 *Paleoceanography*, *23*(4). <https://doi.org/10.1029/2007PA001513>

- 670 Gorbarenko, S., Shi, X., Zou, J., Velivetskaya, T., Artemova, A., Liu, Y., et al. (2019). Evidence of meltwater pulses
671 into the North Pacific over the last 20 ka due to the decay of Kamchatka Glaciers and Cordilleran Ice Sheet.
672 *Global and Planetary Change*, 172, 33–44. <https://doi.org/10.1016/j.gloplacha.2018.09.014>
- 673 Grimm, E. C. (1987). CONISS : A FORTRAN 77 Program for stratigraphically constrained cluster analysis by the
674 method of incremental sum of squares. *Computers and Geosciences*, 13(I), 13–35.
- 675 Hamsher, S. E., LeGresley, M. M., Martin, J. L., & Saunders, G. W. (2013). A comparison of morphological and
676 molecular-based surveys to estimate the species richness of *Chaetoceros* and *Thalassiosira*
677 (Bacillariophyta), in the Bay of Fundy. *PLOS ONE*, 8(10), e73521.
678 <https://doi.org/10.1371/journal.pone.0073521>
- 679 Harada, N., Katsuki, K., Nakagawa, M., Matsumoto, A., Seki, O., Addison, J. A., et al. (2014). Holocene sea surface
680 temperature and sea ice extent in the Okhotsk and Bering Seas. *Progress in Oceanography*, 126, 242–253.
681 <https://doi.org/10.1016/j.pocean.2014.04.017>
- 682 Harrell Jr, F. E. (2020). *Hmisc: Harrell Miscellaneous. R package version 4.4-0*. [https://CRAN.R-](https://CRAN.R-project.org/package=Hmisc)
683 [project.org/package=Hmisc](https://CRAN.R-project.org/package=Hmisc).
- 684 Hasle, G. R. (1990). Arctic plankton diatoms: dominant species, biogeography. In L. K. Medlin & J. Priddle, *Polar*
685 *Marine Diatoms* (pp. 53–56). British Antarctic Survey, Natural Environment Research Council.
- 686 Hasle, G. R., & Evensen, D. L. (1975). Brackish-water and fresh-water species of the diatom genus *Skeletonema*
687 Grev. I. *Skeletonema subsalsum* (A. Cleve) Bethge. *Phycologia*, 14(4), 283–297.
688 <https://doi.org/10.2216/i0031-8884-14-4-283.1>
- 689 Hasle, G. R., & Heimdal, B. R. (1998). The net phytoplankton in Kongsfjorden, Svalbard, July 1988, with general
690 remarks on species composition of Arctic phytoplankton. *Polar Research*, 17(1), 31–52.
691 <https://doi.org/10.3402/polar.v17i1.6605>
- 692 Hofreiter, M., Jaenicke, V., Serre, D., Haeseler, A., & Pääbo, S. (2001). DNA sequences from multiple
693 amplifications reveal artifacts induced by cytosine deamination in ancient DNA. *Nucleic Acids Research*,
694 29. <https://doi.org/10.1093/nar/29.23.4793>
- 695 Hsieh, T. C., Ma, K. H., & Chao, A. (2016). iNEXT: an R package for rarefaction and extrapolation of species
696 diversity (Hill numbers). *Methods in Ecology and Evolution*, 7(12), 1451–1456.
697 <https://doi.org/10.1111/2041-210X.12613>

- 698 Huang, S., Herzsuh, U., Pstryakova, L. A., Zimmermann, H. H., Davydova, P., Biskaborn, B. K., et al. (2020).
699 Genetic and morphologic determination of diatom community composition in surface sediments from
700 glacial and thermokarst lakes in the Siberian Arctic. *Journal of Paleolimnology*.
701 <https://doi.org/10.1007/s10933-020-00133-1>
- 702 Jakobsson, M., Pearce, C., Cronin, T. M., Backman, J., Anderson, L. G., Barrientos, N., et al. (2017). Post-glacial
703 flooding of the Bering Land Bridge dated to 11 cal ka BP based on new geophysical and sediment records.
704 *Climate of the Past*, 13(8), 991–1005. <https://doi.org/10.5194/cp-13-991-2017>
- 705 Juggins, S. (2012). *rioja: Analysis of Quaternary science data, R package version 0.7-3*.
- 706 Kanz, C., Aldebert, P., Althorpe, N., Baker, W., Baldwin, A., Bates, K., et al. (2005). The EMBL nucleotide
707 sequence database. *Nucleic Acids Research*, 33(DATABASE ISS.), 29–33.
708 <https://doi.org/10.1093/nar/gki098>
- 709 Katsuki, K., & Takahashi, K. (2005). Diatoms as paleoenvironmental proxies for seasonal productivity, sea-ice and
710 surface circulation in the Bering Sea during the late Quaternary. *Deep Sea Research Part II: Topical
711 Studies in Oceanography*, 52(16), 2110–2130. <https://doi.org/10.1016/j.dsr2.2005.07.001>
- 712 Kruse, S. (2019). *R code for resampling and thus normalizing of count data to the minimum number of counts
713 across a set of samples (e.g. sedaDNA sequence/pollen taxa counts per sample along a sediment core)*.
714 Retrieved from https://github.com/StefanKruse/R_Rarefaction
- 715 Laskar, J., Robutel, P., Joutel, F., Gastineau, M., Correia, A. C. M., & Levrard, B. (2004). A long-term numerical
716 solution for the insolation quantities of the Earth. *Astronomy and Astrophysics*, 428, 261–285.
717 <https://doi.org/10.1051/0004-6361:20041335>
- 718 Lepskaya, E. V., Jewson, D. H., & Usoltseva, M. V. (2010). *Aulacoseira subarctica* in Kurilskoye Lake,
719 Kamchatka: a deep, oligotrophic lake and important Pacific salmon nursery. *Diatom Research*, 25(2), 323–
720 335. <https://doi.org/10.1080/0269249X.2010.9705853>
- 721 Levitan, M. A., Kuz'mina, T. G., Luksha, V. L., Roshchina, I. A., Syromyatnikov, K. V., Max, L., et al. (2015).
722 Evolution of sedimentation on the continental slope of the Kronotskii Peninsula (Eastern Kamchatka) over
723 the last 20 ka. *Lithology and Mineral Resources*, 50(4), 249–269.
724 <https://doi.org/10.1134/S0024490215040045>

- 725 Li, W. K. W., McLaughlin, F. A., Lovejoy, C., & Carmack, E. C. (2009). Smallest algae thrive as the Arctic Ocean
726 freshens. *Science*, 326(5952), 539–539. <https://doi.org/10.1126/science.1179798>
- 727 Limoges, A., Massé, G., Weckström, K., Poulin, M., Ellegaard, M., Heikkilä, M., et al. (2018). Spring succession
728 and vertical export of diatoms and IP₂₅ in a seasonally ice-covered high Arctic fjord. *Frontiers in Earth
729 Science*, 6. <https://doi.org/10.3389/feart.2018.00226>
- 730 Lopes, C., Mix, A. C., & Abrantes, F. (2006). Diatoms in northeast Pacific surface sediments as paleoceanographic
731 proxies. *Marine Micropaleontology*, 60(1), 45–65. <https://doi.org/10.1016/j.marmicro.2006.02.010>
- 732 Lovejoy, C., Legendre, L., Martineau, M.-J., Bâcle, J., & von Quillfeldt, C. H. (2002). Distribution of phytoplankton
733 and other protists in the North Water. *Deep Sea Research Part II: Topical Studies in Oceanography*,
734 49(22), 5027–5047. [https://doi.org/10.1016/S0967-0645\(02\)00176-5](https://doi.org/10.1016/S0967-0645(02)00176-5)
- 735 Matul', A. G., Saidova, Kh. M., Smirnova, M. A., Khusid, T. A., Kazarina, G. Kh., & Chekhovskaya, M. P. (2015).
736 Rapid diachronous paleoceanographic changes in the Far East marginal areas of the Pacific Ocean at the
737 last glaciation-to-holocene transition. *Doklady Earth Sciences*, 463(2), 873–877.
738 <https://doi.org/10.1134/S1028334X1508022X>
- 739 Max, L., Riethdorf, J.-R., Tiedemann, R., Smirnova, M., Lembke-Jene, L., Fahl, K., et al. (2012a). Biogenic opal of
740 sediment core SO201-2-12 [Data set]. In *supplement to: Max, L et al. (2012): Sea surface temperature
741 variability and sea-ice extent in the subarctic northwest Pacific during the past 15,000 years.*
742 *Paleoceanography*, 27(3), PA3213, <https://doi.org/10.1029/2012PA002292>. PANGAEA.
743 <https://doi.org/10.1594/PANGAEA.786197>
- 744 Max, L., Riethdorf, J.-R., Tiedemann, R., Smirnova, M., Lembke-Jene, L., Fahl, K., et al. (2012b). Sea surface
745 temperature calculated from alkenones of sediment core SO201-2-12 [Data set]. In *supplement to: Max, L
746 et al. (2012): Sea surface temperature variability and sea-ice extent in the subarctic northwest Pacific
747 during the past 15,000 years.* *Paleoceanography*, 27(3), PA3213, <https://doi.org/10.1029/2012PA002292>.
748 PANGAEA. <https://doi.org/10.1594/PANGAEA.786132>
- 749 Max, L., Riethdorf, J.-R., Tiedemann, R., Smirnova, M., Lembke-Jene, L., Fahl, K., et al. (2012). Sea surface
750 temperature variability and sea-ice extent in the subarctic northwest Pacific during the past 15,000 years.
751 *Paleoceanography*, 27(3). <https://doi.org/10.1029/2012PA002292>

- 752 McQuoid, M. R., & Nordberg, K. (2003). The diatom *Paralia sulcata* as an environmental indicator species in
753 coastal sediments. *Estuarine, Coastal and Shelf Science*, 56(2), 339–354. [https://doi.org/10.1016/S0272-](https://doi.org/10.1016/S0272-7714(02)00187-7)
754 [7714\(02\)00187-7](https://doi.org/10.1016/S0272-7714(02)00187-7)
- 755 Medlin, L. K., & Hasle, G. R. (1990). Some *Nitzschia* and related diatom species from fast ice samples in the Arctic
756 and Antarctic. *Polar Biology*, 10(6). <https://doi.org/10.1007/BF00233693>
- 757 Méheust, M., Stein, R., Fahl, K., Max, L., & Riethdorf, J.-R. (2015). High-resolution IP25 of sediment core SO201-
758 2-12 [Data set]. In supplement to: Méheust, M et al. (2015): High-resolution IP25-based reconstruction of
759 sea-ice variability in the western North Pacific and Bering Sea during the past 18,000 years. *Geo-Marine*
760 *Letters*, <https://doi.org/10.1007/s00367-015-0432-4>. PANGAEA.
761 <https://doi.org/10.1594/PANGAEA.855451>
- 762 Méheust, M., Stein, R., Fahl, K., Max, L., & Riethdorf, J.-R. (2016). High-resolution IP25-based reconstruction of
763 sea-ice variability in the western North Pacific and Bering Sea during the past 18,000 years. *Geo-Marine*
764 *Letters*, 36(2), 101–111. <https://doi.org/10.1007/s00367-015-0432-4>
- 765 Meiners, K., Krembs, C., & Gradinger, R. (2008). Exopolymer particles: microbial hotspots of enhanced bacterial
766 activity in Arctic fast ice (Chukchi Sea). *Aquatic Microbial Ecology*, 52(2), 195–207.
767 <https://doi.org/10.3354/ame01214>
- 768 Meyer, V. D., Max, L., Hefter, J., Tiedemann, R., & Mollenhauer, G. (2016a). Fractional abundances of isoprenoid
769 and branched glycerol dialkyl glycerol tetraethers (GDGT) of sediment core SO201-2-12KL [Data set]. In
770 supplement to: Meyer, VD et al. (2016): Glacial-to-Holocene evolution of sea surface temperature and
771 surface circulation in the subarctic northwest Pacific and the Western Bering Sea. *Paleoceanography*, 31,
772 12 pp, <https://doi.org/10.1002/2015PA002877>. PANGAEA. <https://doi.org/10.1594/PANGAEA.862964>
- 773 Meyer, V. D., Max, L., Hefter, J., Tiedemann, R., & Mollenhauer, G. (2016b). Glacial-to-Holocene evolution of sea
774 surface temperature and surface circulation in the subarctic northwest Pacific and the Western Bering Sea.
775 *Paleoceanography*, 31(7), 916–927. <https://doi.org/10.1002/2015PA002877>
- 776 Meyer, V. D., Hefter, J., Lohmann, G., Max, L., Tiedemann, R., & Mollenhauer, G. (2017). Summer temperature
777 evolution on the Kamchatka Peninsula, Russian Far East, during the past 20 000 years. *Climate of the Past*,
778 13(4), 359–377. <https://doi.org/10.5194/cp-13-359-2017>

- 779 Nagano, A., Wakita, M., & Watanabe, S. (2016). Dichothermal layer deepening in relation with halocline depth
780 change associated with northward shrinkage of North Pacific western subarctic gyre in early 2000s. *Ocean*
781 *Dynamics*, 66(2), 163–172. <https://doi.org/10.1007/s10236-015-0917-8>
- 782 Nummelin, A., Ilicak, M., Li, C., & Smedsrud, L. H. (2016). Consequences of future increased Arctic runoff on
783 Arctic Ocean stratification, circulation, and sea ice cover. *Journal of Geophysical Research: Oceans*,
784 121(1), 617–637. <https://doi.org/10.1002/2015JC011156>
- 785 Oksanen, J., Blanchet, F. G., Kindt, R., Legendre, P., Minchin, P. R., O'Hara, R. B., et al. (2011). *vegan*:
786 *Community Ecology Package. R package version 2.0-2.*
- 787 Olsen, L. M., Duarte, P., Peralta-Ferriz, C., Kauko, H. M., Johansson, M., Peeken, I., et al. (2019). A red tide in the
788 pack ice of the Arctic Ocean. *Scientific Reports*, 9. <https://doi.org/10.1038/s41598-019-45935-0>
- 789 Pääbo, S. (1989). Ancient DNA: extraction, characterization, molecular cloning, and enzymatic amplification.
790 *Proceedings of the National Academy of Sciences of the United States of America*, 86(6), 1939–1943.
- 791 Parkinson, C. L., Cavalieri, D. J., Gloersen, P., Zwally, H. J., & Comiso, J. C. (1999). Arctic sea ice extents, areas,
792 and trends, 1978–1996. *Journal of Geophysical Research: Oceans*, 104(C9), 20837–20856.
793 <https://doi.org/10.1029/1999JC900082>
- 794 Pawłowska, J., Łacka, M., Kucharska, M., Pawłowski, J., & Zajączkowski, M. (2020). Multiproxy evidence of the
795 Neoglacial expansion of Atlantic Water to eastern Svalbard. *Climate of the Past*, 16(2), 487–501.
796 <https://doi.org/10.5194/cp-16-487-2020>
- 797 Pistone, K., Eisenman, I., & Ramanathan, V. (2014). Observational determination of albedo decrease caused by
798 vanishing Arctic sea ice. *Proceedings of the National Academy of Sciences*, 111(9), 3322–3326.
799 <https://doi.org/10.1073/pnas.1318201111>
- 800 Polyakova, A. M. (2007). Extreme supply of floating ice to the northwestern part of the Pacific Ocean. *Oceanology*,
801 47(1), 1–4. <https://doi.org/10.1134/S0001437007010018>
- 802 Poulin, M., Daughjerg, N., Gradinger, R., Ilyash, L., Ratkova, T., & Quillfeldt, C. von. (2011). The pan-Arctic
803 biodiversity of marine pelagic and sea-ice unicellular eukaryotes: a first-attempt assessment. *Marine*
804 *Biodiversity*, 41(1), 13–28. <https://doi.org/10.1007/s12526-010-0058-8>
- 805 von Quillfeldt, C. H. (1997). Distribution of diatoms in the Northeast Water Polynya, Greenland. *Journal of Marine*
806 *Systems*, 10(1), 211–240. [https://doi.org/10.1016/S0924-7963\(96\)00056-5](https://doi.org/10.1016/S0924-7963(96)00056-5)

- 807 von Quillfeldt, C. H. (2000). Common diatom species in Arctic spring blooms: their distribution and abundance.
808 *Botanica Marina*, 43(6). <https://doi.org/10.1515/BOT.2000.050>
- 809 R Core Team. (2018). *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for
810 Statistical Computing. Retrieved from <https://www.R-project.org/>
- 811 Rasmussen, S. O., Bigler, M., Blockley, S. P., Blunier, T., Buchardt, S. L., Clausen, H. B., et al. (2014). A
812 stratigraphic framework for abrupt climatic changes during the Last Glacial period based on three
813 synchronized Greenland ice-core records: refining and extending the INTIMATE event stratigraphy.
814 *Quaternary Science Reviews*, 106, 14–28. <https://doi.org/10.1016/j.quascirev.2014.09.007>
- 815 Ren, J., Jiang, H., Seidenkrantz, M.-S., & Kuijpers, A. (2009). A diatom-based reconstruction of Early Holocene
816 hydrographic and climatic change in a southwest Greenland fjord. *Marine Micropaleontology*, 70(3–4),
817 166–176. <https://doi.org/10.1016/j.marmicro.2008.12.003>
- 818 Ren, J., Gersonde, R., Esper, O., & Sancetta, C. (2014). Diatom distributions in northern North Pacific surface
819 sediments and their relationship to modern environmental variables. *Palaeogeography, Palaeoclimatology,*
820 *Palaeoecology*, 402, 81–103. <https://doi.org/10.1016/j.palaeo.2014.03.008>
- 821 Reschke, M., Kunz, T., & Laepple, T. (2019). Comparing methods for analysing time scale dependent correlations
822 in irregularly sampled time series data. *Computers & Geosciences*, 123, 65–72.
823 <https://doi.org/10.1016/j.cageo.2018.11.009>
- 824 Riethdorf, J.-R., Max, L., Nürnberg, D., Lembke-Jene, L., & Tiedemann, R. (2013a). Deglacial development of
825 (sub) sea surface temperature and salinity in the subarctic northwest Pacific: Implications for upper-ocean
826 stratification: deglacial subsurface temperature and salinity in the NW Pacific. *Paleoceanography*, 28(1),
827 91–104. <https://doi.org/10.1002/palo.20014>
- 828 Riethdorf, J.-R., Max, L., Nürnberg, D., Lembke-Jene, L., & Tiedemann, R. (2013b). Stable isotopes measured on
829 benthic foraminifers of sediment core SO201-2-12 [Data set]. *In supplement to: Riethdorf, J-R et al.*
830 *(2013): Deglacial development of (sub) sea surface temperature and salinity in the subarctic northwest*
831 *Pacific: Implications for upper-ocean stratification. Paleoceanography*, 28(1), 91-104,
832 <https://doi.org/10.1002/palo.20014>. PANGAEA. <https://doi.org/10.1594/PANGAEA.786246>

- 833 Rimet, F., Gusev, E., Kahlert, M., Kelly, M. G., Kulikovskiy, M., Maltsev, Y., et al. (2019). Diat.barcode, an open-
834 access curated barcode library for diatoms. *Scientific Reports*, 9(1), 15116. [https://doi.org/10.1038/s41598-](https://doi.org/10.1038/s41598-019-51500-6)
835 019-51500-6
- 836 Rotatore, C., Colman, B., & Kuzma, M. (1995). The active uptake of carbon dioxide by the marine diatoms
837 *Phaeodactylum ticornutum* and *Cyclotella* sp. *Plant, Cell & Environment*, 18(8), 913–918.
838 <https://doi.org/10.1111/j.1365-3040.1995.tb00600.x>
- 839 Round, F. E., Crawford, R. M., & Mann, D. G. (1990). *Diatoms: Biology and morphology of the genera*. Cambridge
840 University Press.
- 841 Sancetta, C. (1979). Oceanography of the North Pacific during the last 18,000 years: Evidence from fossil diatoms.
842 *Marine Micropaleontology*, 4, 103–123. [https://doi.org/10.1016/0377-8398\(79\)90009-4](https://doi.org/10.1016/0377-8398(79)90009-4)
- 843 Sancetta, C. (1982). Distribution of diatom species in surface sediments of the Bering and Okhotsk Seas.
844 *Micropaleontology*, 28(3), 221–257. <https://doi.org/10.2307/1485181>
- 845 Sancetta, C., & Silvestri, S. (1986). Pliocene-Pleistocene evolution of the North Pacific Ocean-Atmosphere system,
846 interpreted from fossil diatoms. *Paleoceanography*, 1(2), 163–180.
847 <https://doi.org/10.1029/PA001i002p00163>
- 848 Schlitzer, R. (2002). Interactive analysis and visualization of geoscience data with Ocean Data View,
849 <https://odv.awi.de>. *Computers & Geosciences*, 28(10), 1211–1218. [https://doi.org/10.1016/S0098-](https://doi.org/10.1016/S0098-3004(02)00040-7)
850 3004(02)00040-7
- 851 Schnell, I. B., Bohmann, K., & Gilbert, M. T. P. (2015). Tag jumps illuminated – reducing sequence-to-sample
852 misidentifications in metabarcoding studies. *Molecular Ecology Resources*, 15(6), 1289–1303.
853 <https://doi.org/10.1111/1755-0998.12402>
- 854 Shu, Q., Qiao, F., Song, Z., Zhao, J., & Li, X. (2018). Projected freshening of the Arctic Ocean in the 21st century.
855 *Journal of Geophysical Research: Oceans*, 123(12), 9232–9244. <https://doi.org/10.1029/2018JC014036>
- 856 Smirnova, M. A., Kazarina, G. Kh., Matul, A. G., & Max, L. (2015). Diatom evidence for paleoclimate changes in
857 the northwestern Pacific during the last 20000 years. *Oceanology*, 55(3), 383–389.
858 <https://doi.org/10.1134/S0001437015030157>
- 859 Stabell, B. (1986). A diatom maximum horizon in upper Quaternary deposits. *Geologische Rundschau*, 75(1), 175–
860 184. <https://doi.org/10.1007/BF01770186>

- 861 Stabeno, P. J., & Reed, R. K. (1994). Circulation in the Bering Sea Basin Observed by Satellite-Tracked Drifters:
862 1986–1993. *Journal of Physical Oceanography*, 24(4), 848–854. <https://doi.org/10.1175/1520->
863 0485(1994)024<0848:CITBSB>2.0.CO;2
- 864 Stabeno, P. J., Schumacher, J. D., & Ohtani, K. (1999). The physical oceanography of the Bering Sea. In T. R.
865 Loughlin & K. Ohtani (Eds.), *Dynamics of the Bering Sea: A summary of physical, chemical, and*
866 *biological characteristics, and a synopsis of research on the Bering Sea* (pp. 1–28). University of Alaska
867 Sea Grant, Fairbanks: North Pacific Marine Science Organization. Retrieved from
868 <https://www.pmel.noaa.gov/pubs/outstand/stab1878/stab1878.shtml>
- 869 Stoof-Leichsenring, K. R., Epp, L. S., Trauth, M. H., & Tiedemann, R. (2012). Hidden diversity in diatoms of
870 Kenyan Lake Naivasha: a genetic approach detects temporal variation. *Molecular Ecology*, 21(8), 1918–
871 1930. <https://doi.org/10.1111/j.1365-294X.2011.05412.x>
- 872 Syvertsen, E. E. (1991). Ice algae in the Barents Sea: types of assemblages, origin, fate and role in the ice-edge
873 phytoplankton bloom. *Polar Research*, 10(1), 277–288. <https://doi.org/10.1111/j.1751->
874 8369.1991.tb00653.x
- 875 Tremblay, J.-É., & Gagnon, J. (2009). The effects of irradiance and nutrient supply on the productivity of Arctic
876 waters: a perspective on climate change. In J. C. J. Nihoul & A. G. Kostianoy (Eds.), *Influence of Climate*
877 *Change on the Changing Arctic and Sub-Arctic Conditions* (pp. 73–93). Dordrecht: Springer Netherlands.
878 https://doi.org/10.1007/978-1-4020-9460-6_7
- 879 Vincent, F., & Bowler, C. (2020). Diatoms are selective segregators in global ocean planktonic communities.
880 *MSystems*, 5(1). <https://doi.org/10.1128/mSystems.00444-19>
- 881 Walsh, J. E., Fetterer, F., Stewart, J. S., & Chapman, W. L. (2017). A database for depicting Arctic sea ice variations
882 back to 1850. *Geographical Review*, 107(1), 89–107. <https://doi.org/10.1111/j.1931-0846.2016.12195.x>
- 883 Zimmermann, H. H., Raschke, E., Epp, L. S., Stoof-Leichsenring, K. R., Schwamborn, G., Schirrmeister, L., et al.
884 (2017). Sedimentary ancient DNA and pollen reveal the composition of plant organic matter in Late
885 Quaternary permafrost sediments of the Buor Khaya Peninsula (north-eastern Siberia). *Biogeosciences*,
886 14(3), 575–596. <https://doi.org/10.5194/bg-14-575-2017>
- 887 Zimmermann, H. H., Stoof-Leichsenring, K. R., Kruse, S., Müller, J., Stein, R., Tiedemann, R., & Herzsuh, U.
888 (2019). Changes in the composition of marine and sea-ice diatoms derived from sedimentary ancient DNA

889 of the eastern Fram Strait over the past 30,000 years. *Ocean Science Discussions*, 1–25.
890 <https://doi.org/10.5194/os-2019-113>

891 Zimmermann, H. H., Stoof-Leichsenring, K. R., Kruse, S., Nuernberg, D., Tiedemann, R., & Herzsuh, U. (2020).
892 Diatom sedimentary ancient DNA metabarcoding from Kronotsky Peninsula. *Dryad, Dryad Dataset*.
893 <https://doi.org/10.5061/dryad.qnk98sfdh>

894 Zimmermann, H. H., Stoof-Leichsenring, K. R., Kruse, S., Müller, J., Stein, R., Tiedemann, R., & Herzsuh, U.
895 (2020). Diatom sedimentary ancient DNA metabarcoding from western Fram Strait and Kronotsky
896 Peninsula. *Dryad, Dryad Dataset*. <https://doi.org/10.5061/dryad.bnzs7h481>

897 Zimmermann, J., Abarca, N., Enk, N., Skibbe, O., Kusber, W.-H., & Jahn, R. (2014). Taxonomic reference libraries
898 for environmental barcoding: A best practice example from diatom research. *PLoS ONE*, 9(9), e108793.
899 <https://doi.org/10.1371/journal.pone.0108793>

900