

# **COVID-19 hypothesis: Exosomes of mesenchymal stem cells as nano-cargos for anti-SARS-CoV-2 asRNAs**

Running title: **Anti-SARS-CoV-2 asRNAs' MSC exosomes**

Alireza Afshar<sup>1\*</sup>, Masoud Zare<sup>1\*</sup>, Zohreh Farrar<sup>1\*</sup>, Alireza Hashemi<sup>1\*</sup>, Arezoo Khoradmehr<sup>1</sup>, Hassan Habibi<sup>2</sup>, Mohammad Amin Behzadi<sup>3†</sup>, Amin Tamadon<sup>1†</sup>

1- The Persian Gulf Marine Biotechnology Research Center, The Persian Gulf Biomedical Sciences Research Institute, Bushehr University of Medical Sciences, Bushehr, Iran

2- Department of Animal Sciences, Agriculture and Natural Resources College, Persian Gulf University, Bushehr, Iran

3- Department of Microbiology, Icahn School of Medicine at Mount Sinai, New York, NY, USA

\* These authors have same contribution as first authors.

† Corresponding authors:

Amin Tamadon, DVM, PhD; The Persian Gulf Marine Biotechnology Research Center, The Persian Gulf Biomedical Sciences Research Institute, Bushehr University of Medical Sciences, Bushehr, Iran; Postal Code: 7514633196; Tel/fax: +98-77-3332-8724; Email: [amintamaddon@yahoo.com](mailto:amintamaddon@yahoo.com)

Mohammad Amin Behzadi, DVM, PhD; Department of Microbiology, Icahn School of Medicine at Mount Sinai, New York, NY, USA; Postal Code: 10029; Tel/fax: +1-212-241-7318; Email: [ma.behzadi@mssm.edu](mailto:ma.behzadi@mssm.edu)

**Authors emails:**

Alireza Afshar, [alireza.af2017@gmail.com](mailto:alireza.af2017@gmail.com)

Masood Zare, [md.zare77@gmail.com](mailto:md.zare77@gmail.com)

Zohre Farrar, [zohrehfarrar@gmail.com](mailto:zohrehfarrar@gmail.com)

Alireza Hashemi, [alireza\\_hashemi89@yahoo.com](mailto:alireza_hashemi89@yahoo.com)

Arezo Khoradmehr, [mehrarezoo@gmail.com](mailto:mehrarezoo@gmail.com)

Hassan Habibi, [dr.h.habibi@gmail.com](mailto:dr.h.habibi@gmail.com)

Mohammad Amin Behzadi, [ma.behzadi@mssm.edu](mailto:ma.behzadi@mssm.edu)

Amin Tamadon, [amintamaddon@yahoo.com](mailto:amintamaddon@yahoo.com)

## **Abstract**

The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) emerged in December 2019 is rapidly spreading worldwide. Scientists are searching to find an effective treatment for coronavirus disease 2019 (COVID-19). Several antiviral drugs are currently undergoing clinical trial studies to evaluate their safety and efficacy in the treatment of COVID-19. SARS-CoV-2 is a positive-sense single-stranded RNA virus. Previous studies showed the efficacy of anti-RNA virus, single strand RNA inhibiting antisense RNAs (asRNAs), on silencing of virus replication, *in vitro*. To transfer the anti-SARS-CoV-2 asRNAs to human respiratory epithelium, exosomes can be suggested as a promising candidate. Mesenchymal stem cells (MSCs) secrete exosomes and they can be loaded by anti-RNA virus asRNAs. MSCs-secreted exosomes as a nano-cargo of anti-SARS-CoV-2 asRNAs have other therapeutic potentials such as immunomodulatory effects of their cytokine contents, affinity to respiratory epithelial attachment, anti-fibrotic activity in lung, non-toxicity for normal cells, and do not trigger an immune response. Inhalation of anti-SARS-CoV-2 asRNAs may stop SARS-CoV-2 replication. Producing a specific anti-SARS-CoV-2 asRNAs by targeting the genome of virus and their delivery by MSCs exosomes is suggested and discussed. This approach potentially sheds light on gene therapy of the other human lung diseases via inhalational delivery using exosomes in future.

**Keywords:** SARS-CoV-2, COVID-19, antisense RNA, exosome, mesenchymal stem cells, treatment

## Introduction

Infections with the majority of respiratory viruses are a global health concern and are known as one of the main causes of death among high-risk population (1). Coronavirus disease 2019 (COVID-19) is one of the respiratory viral infections caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), emerging from Wuhan, China and shortly led to a catastrophic worldwide epidemic health problem (2). The virus belongs to the family of coronaviruses such as severe acute respiratory syndrome (SARS) and Middle-East respiratory syndrome (MERS) (3-5). SARS-CoV-2 is a positive-sense single-stranded RNA virus showing a high rate of pathogenesis (2). The symptoms of COVID-19 range from mild to severe and may include fever, cough, chills, shortness of breath and bilateral pneumonia in end-stage patients (4). Although, some therapeutics are suggested for curing some symptoms of COVID-19 and other coronaviruses but to the best of our knowledge, there is no effective antiviral to treat COVID-19 (6).

Anti-inflammatory and immunomodulatory effects of MSCs as a therapeutic approach for various types of respiratory diseases have been confirmed by clinical studies (7). Different important sources of MSCs including Wharton's jelly, umbilical cord, umbilical cord blood, dental pulp, menstrual blood, are used in these clinical trials (7). Compatibility of MSCs-derived exosomes with host immune system, their nano-size and low-toxicity making them a very efficient drug carrier (8-10). As in patients of COVID-19, SARS and MERS, serum concentrations of pro-inflammatory cytokines (IL-1, IL-6, and TNF- $\alpha$ ) and chemokines (IL-8) increase (11), using MSCs-derived exosomes for transportation of asRNA may modulate these signs, as well.

As the target of this therapeutic approach is the infected epithelial cells of respiratory tract, drug administration by inhalation has been suggested. Lung cancer therapy by inhalation of MMP2/9-triggered-release micelles had appropriate effects (12). Using serum-derived exosomes as a vehicle to small RNA molecules via inhalational delivery into the lung macrophages *in vivo* has been shown in mice for lung inflammation treatment (13). In our hypothesis exosomes containing asRNAs can introduce to patients with inhalation, so the asRNA may reach to the respiratory system with more efficacy.

We hypothesized MSCs-derived exosomes can perform as a carrier to deliver anti-SARS-CoV-2 miR-5197-3p to infected cells to decrease the viruses' replication and transcription *in vivo* (14). Table 1 summarized locations and products of single strand RNA of SARS-CoV-2 can be inhibited by asRNAs. It may be useful to exploit the MSCs-derived exosomes to efficiently deliver anti-SARS-CoV-2 asRNAs for targeted drug delivery (8). Also, previous studies have demonstrated that exosome based drug delivery can protect exosome-encapsulating RNAs from RNase enzymes (15). Different exosomes with cell-specific surface proteins may have distinct routes and circulation pattern all over the body (16). Accordingly, it can be predicted that MSCs-derived exosomes contain the targeted asRNA for those viruses. To our knowledge, the function of these asRNA filled exosomes derived from MSCs has not been investigated.

Human Wharton jelly-derived MSCs is one of the most important sources of primary MSCs that can be obtained from umbilical cord with standard methods (17). MSCs and hematopoietic stem cells CD markers can be analyzed by flow cytometry to confirm purity of MSCs isolation. In order to appraise the differentiation capabilities of MSCs, the cells will be test for osteogenic, adipogenic, and chondrogenic differentiation. The conditioned medium (CM) will be prepared using MSCs (18). The exosomes will be isolated from CM by commercial kit

(19). Transmission electron microscopy test will be used for morphological assessment of the isolated exosomes (20).

### **Transportation of antisense RNA inhibiting RNAs (asRNAs)**

RNA viruses use intracellular host cell machinery for DNA replication, RNA transcription and protein synthesis (21). The pathogenesis of such viruses is mainly depends on the rate of genome replication and cell destruction following host immune responses (22). Thus, blocking the viral genome replication and transcription can be a promising approach to combat the infection (23). To selective inhibition of RNA virus replication, many targets exist (23). However, anti-replication effect of antisense RNA inhibiting RNAs (asRNAs) as a non-coding RNAs (ncRNAs) has not been investigated for coronaviruses. Although, significant improvements have been made in the field of nucleic acid-based therapies and numerous carriers have been used to transport molecules such as asRNA, studies on finding suitable carriers for these kinds of molecules are still ongoing (24). The other challenge of designing an anti-SARS-CoV-2 asRNA is transportation of them into the replication site, host respiratory epithelial cells. As a consequence of the nanometer size of asRNAs, they can go through the vessels and affect other parts of the host's body (25). One of the main problems in the asRNA transfer, related to their instability and negative charge, and hence, even in the presence of a suitable transfection reagent, it cannot effectively penetrate the hydrophobic cell membranes (8, 26).

To minimize the side effects of asRNAs on other tissues, we can use targeted drug delivery system. For this purpose, exosome-based drug delivery approach can be one of the best candidates (27). On the other hand, the mesenchymal stem cells (MSCs) have been extensively studied for their immunosuppressive effects.

The target asRNAs will be produced using a cost-effective DGCR8-independent stable asRNA expression (DISME) strategy (28). To increase the expression level of asRNAs, units of a U6 promoter-driven expression cassette will be inserted in the vector (28). The exosomes will be loaded with anti-SARS-CoV-2 miR-4778-3p, using electroporation method (8). The loaded exosomes were reisolated using commercial kit (19). The amount of anti-SARS-CoV-2 asRNAs oligonucleotides in MSCs-derived exosomes will be estimated as previously described (8). Then, the asRNA loaded exosomes will be exposed to SARS-CoV-2 culture in transmembrane protease, serine 2 (TMPRSS2)-expressing VeroE6 cell line and virus replication will be analyzed (29).

To test the therapeutic method in animal model, two approved model of rhesus macaques and ferrets can be used (30). These models can be infected with SARS-CoV-2 and they show virus replication and shed virus (30). After confirming of the treatment in animal model, asRNAs efficiency analyzing, the asRNAs encapsulated with exosomes will be introduced to the patient with inhalation (14). Figure 1 summarized this therapeutic approach.

## **Discussion**

Exosomes have been introduced as a new alternative to the transmission of therapeutic molecules (27). Exosomes are nano-sized lipid package that are produced in the most eukaryotic cells (31). They are capable of being drug carriers since they are composed of cell membranes, rather than synthetic polymers (27). Exosomes have many functions in human bodies including cell to cell communication, wound healing, tissue regeneration and even cell death (32). They have some information from their primary parental cells (33). Exosomes with 60-80 nm size are the smallest particles between entire exosomes (34). They can interact with macromolecules and that they can

serve as distributors for proteins, lipids, mRNA, miRNA, and DNA (35). Exosomes may interact with the host's immune system, so selection of the parent cell for exosome production need for careful (36). Application of human exosomes for treatment of diseases are tested in clinical trials (37) and have a developing market (38).

MSCs have been used in many clinical trials of transplant rejection, autoimmune disorders, and inflammation- associated diseases (39). It has been shown that MSC-secreted factors suppress T-cell proliferation (40). Therefore, MSCs-derived exosomes have the same properties as their parent cell (41). With immunomodulatory characteristics, MSCs-derived exosomes suppress the secretion of tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and interleukin-1 (IL-1) in peripheral blood mononuclear cells (41). Some of the earliest exosome research indicated that they can carry the MHC-peptide complexes that are recognized by T lymphocytes (42) in order to more activate cell immune system and destroy the tumor and other cell contaminant (for example viruses) (43).

The asRNAs therapy is suggested as a new method for controlling viral infections (44). Here, we hypothesized MSC derived exosomes containing anti- SARS-CoV-2 asRNA to help finding a specific treatment for COVID-19. Since the SARS-CoV-2 genome replicates in respiratory epithelium of host cells, the developed asRNAs should have characters in order to not damage genome of host cells. Over the past decade, discovery of ncRNAs including microRNA, small interfering RNA (siRNA), PIWI-interacting RNA (piRNA) and long noncoding RNA (lncRNA) is a well-defined research topic. Application of asRNAs considering the required purpose undergoes two challenges: 1) The asRNA binding to the exact site of the RNA virus genome. 2) Side effects of the randomly binding of these asRNAs to human mRNAs due to the subjective set of nucleotides in synthetic asRNAs those bind with complete complementarity

(45-48). It is necessary to select asRNAs that can most effectively bind to the nucleotide sequence of the RNA genomes of these coronaviruses. Therefore, constructing specific asRNAs for coronaviruses of COVID-19, SARS, and MERS that will not affect the expression of human or animal genes is crucial (14).

In this case, asRNA with the ability of specific binding to viral genome, must be selected (14, 49). *In-silico* analysis showed that miR-5197-3p (cc-miR, fully complementary miRNA) has affinity to ssRNA of SARS-CoV-2 and also two miR-6864-5p and miR-4778-5p can strongly bind to MERS (cc-miRm) and SARS (cc-miRs) genomes, respectively (14) (Table 1). The miR-5197-3p can bind to a few human genes which they have similar characteristic (14). In order to avoid the potential side effects, this asRNA has sustained some changes in the length and nucleotide sequences and called cc-miR2 (fully complementary miRNA). This new asRNA is more efficient to bind with ssRNA of SARS-CoV-2 without reacting with human protein-coding genes and presenting related side effects (14). The next concern of this therapeutic approach is related to the possibility of SARS-CoV-2 genomic mutations during worldwide spreading of the virus. Fortunately, comparative sequence analyses of SAR-CoV2 genomes isolated from different geographical locations have unique features (50). Therefore, designing a conserved cc-miRNA is possible.

The SARS-CoV-2 binds to epithelial cells of the nasal cavity after inhalation of virus and replicates there (51). Then along the conducting airways, SARS-CoV-2 migrates down the respiratory tract (52). Studies utilizing exosomes for treatment of different kind of lung diseases are mainly administer by systemically into a vein or direct tissue injection. But treatment of the lungs by inhalation is the minimally invasive and most direct route of delivery (53). Nonetheless, innovative methods must be developed for the scale up of exosome production and isolation.

## **Conclusions**

The asRNAs can inhibit ssRNA of SARS-CoV-2 will be effective on silencing of virus replication. MSCs secret exosomes and they can be loaded by anti-RNA virus asRNA. Producing a specific anti-SARS-CoV-2 by targeting the viral genome can be suggested for developing a novel therapeutic candidate tor the disease.

## **Authors' contributions**

A.A., M.Z., Z.F., A.H., A.K., and H.H. collected information and co-wrote the draft. K.V., I.N., M.A.B., and A.T. designed the idea, and critically revised the paper.

## **Competing interests**

There is no conflict of interest.

## **Acknowledgements**

Not applicable.

## **References**

1. Ferkol T, Schraufnagel D. The global burden of respiratory disease. *Ann Am Thorac Soc*. 2014;11(3):404-6.
2. Gorbalenya AE, Baker SC, Baric RS, Groot RJd, Drosten C, Gulyaeva AA, et al. The species Severe acute respiratory syndrome-related coronavirus: classifying 2019-nCoV and naming it SARS-CoV-2. *Nat Microbiol*. 2020;5:536–44.

3. Liu J, Zheng X, Tong Q, Li W, Wang B, Sutter K, et al. Overlapping and discrete aspects of the pathology and pathogenesis of the emerging human pathogenic coronaviruses SARS-CoV, MERS-CoV, and 2019-nCoV. *Journal of Medical Virology*. 2020.
4. World Health Organization. Key Messages and Actions for COVID-19 Prevention and Control in Schools. 2020.
5. Sardar T, Ghosh I, Rodó X, Chattopadhyay J. A realistic two-strain model for MERS-CoV infection uncovers the high risk for epidemic propagation. *PLoS neglected tropical diseases*. 2020;14(2):e0008065.
6. Behzadi MA, Leyva-Grado VH. Overview of current therapeutics and novel candidates against influenza, respiratory syncytial virus and Middle East respiratory syndrome coronavirus infections. *Frontiers in microbiology*. 2019;10:1327.
7. Golchin A, Seyedjafari E, Ardeshirylajimi A. Mesenchymal Stem Cell Therapy for COVID-19: Present or Future. *Stem Cell Reviews and Reports*. 2020:1-7.
8. Naseri Z, Oskuee RK, Jaafari MR, Moghadam MF. Exosome-mediated delivery of functionally active miRNA-142-3p inhibitor reduces tumorigenicity of breast cancer in vitro and in vivo. *Int J Nanomedicine*. 2018;13:7727.
9. van Dommelen SM, Vader P, Lakhal S, Kooijmans S, van Solinge WW, Wood MJ, et al. Microvesicles and exosomes: opportunities for cell-derived membrane vesicles in drug delivery. *Journal of Controlled Release*. 2012;161(2):635-44.
10. Vlassov AV, Magdaleno S, Setterquist R, Conrad R. Exosomes: current knowledge of their composition, biological functions, and diagnostic and therapeutic potentials. *Biochimica et Biophysica Acta*. 2012;1820(7):940-8.

11. Qin C, Zhou L, Hu Z, Zhang S, Yang S, Tao Y, et al. Dysregulation of immune response in patients with COVID-19 in Wuhan, China. *Clinical Infectious Diseases*. 2020.
12. Wang X, Chen Q, Zhang X, Ren X, Zhang X, Meng L, et al. Matrix metalloproteinase 2/9-triggered-release micelles for inhaled drug delivery to treat lung cancer: preparation and in vitro/in vivo studies. *International journal of nanomedicine*. 2018;13:4641-59.
13. Zhang D, Lee H, Wang X, Rai A, Groot M, Jin Y. Exosome-mediated small RNA delivery: a novel therapeutic approach for inflammatory lung responses. *Molecular Therapy*. 2018;26(9):2119-30.
14. Ivashchenko A, Rakhmetullina A, Aisina D. How miRNAs can protect humans from coronaviruses COVID-19, SARS-CoV, and MERS-CoV. *Research Square*. 2020:10.21203/rs.3.rs-16264/v1.
15. Lee H, Abston E, Zhang D, Rai A, Jin Y. Extracellular vesicle: an emerging mediator of intercellular crosstalk in lung inflammation and injury. *Frontiers in immunology*. 2018;9:924.
16. Samanta S, Rajasingh S, Drosos N, Zhou Z, Dawn B, Rajasingh J. Exosomes: new molecular targets of diseases. *Acta Pharmacologica Sinica*. 2018;39(4):501-13.
17. Rezaeian L, Hosseini SE, Dianatpour M, Edalatmanesh MA, Tanideh N, Mogheiseh A, et al. Intrauterine xenotransplantation of human Wharton jelly-derived mesenchymal stem cells into the liver of rabbit fetuses: A preliminary study for in vivo expression of the human liver genes. *Iranian J Basic Med Sci*. 2018;21(1):89.
18. Chudickova M, Vackova I, Machova Urdzikova L, Jancova P, Kekulova K, Rehorova M, et al. The effect of Wharton jelly-derived mesenchymal stromal cells and their conditioned media in the treatment of a rat spinal cord injury. *International journal of molecular sciences*. 2019;20(18):4516.

19. Lobb RJ, Becker M, Wen Wen S, Wong CS, Wiegman AP, Leimgruber A, et al. Optimized exosome isolation protocol for cell culture supernatant and human plasma. *J Extracell Vesicles*. 2015;4(1):27031.
20. Lv C-X, Duan H, Wang S, Gan L, Xu Q. Exosomes derived from human umbilical cord mesenchymal stem cells promote proliferation of allogeneic endometrial stromal cells. *Reproductive Sciences*. 2020:1-10.
21. Walsh D, Mathews MB, Mohr I. Tinkering with translation: protein synthesis in virus-infected cells. *Cold Spring Harbor perspectives in biology*. 2013;5(1):a012351.
22. Ryan EL, Hollingworth R, Grand RJ. Activation of the DNA Damage Response by RNA Viruses. *Biomolecules*. 2016;6(1):2-.
23. Leyssen P, De Clercq E, Neyts J. Molecular strategies to inhibit the replication of RNA viruses. *Antiviral research*. 2008;78(1):9-25.
24. Burnett JC, Rossi JJ. RNA-based therapeutics: current progress and future prospects. *Chemistry & biology*. 2012;19(1):60-71.
25. Sohel MH. Extracellular/circulating microRNAs: release mechanisms, functions and challenges. *Achiev Life Sci*. 2016;10(2):175-86.
26. Aagaard L, Rossi JJ. RNAi therapeutics: principles, prospects and challenges. *Advanced drug delivery reviews*. 2007;59(2-3):75-86.
27. Lakhal S, Wood MJ. Exosome nanotechnology: an emerging paradigm shift in drug delivery: exploitation of exosome nanovesicles for systemic in vivo delivery of RNAi heralds new horizons for drug delivery across biological barriers. *Bioessays*. 2011;33(10):737-41.

28. Wang X-W, Hao J, Guo W-T, Liao L-Q, Huang S-Y, Guo X, et al. A DGCR8-independent stable microRNA expression strategy reveals important functions of miR-290 and miR-183-182 families in mouse embryonic stem cells. *Stem Cell Rep.* 2017;9(5):1618-29.
29. Matsuyama S, Nao N, Shirato K, Kawase M, Saito S, Takayama I, et al. Enhanced isolation of SARS-CoV-2 by TMPRSS2-expressing cells. *Proc Natl Acad Sci U S A.* 2020;117(13):7001-3.
30. WHO COVID-19 modelling ad hoc expert working group. COVID-19 Animal Models. 2020.
31. Van Niel G, d'Angelo G, Raposo G. Shedding light on the cell biology of extracellular vesicles. *Nat Rev Mol Cell Biol.* 2018;19(4):213.
32. Panagiotou N, Davies RW, Selman C, Shiels PG. Microvesicles as vehicles for tissue regeneration: changing of the guards. *Curr Pathobiol Rep.* 2016;4(4):181-7.
33. Maia J, Caja S, Strano Moraes MC, Couto N, Costa-Silva B. Exosome-based cell-cell communication in the tumor microenvironment. *Front Cell Dev Biol.* 2018;6:18.
34. Lane RE, Korbie D, Anderson W, Vaidyanathan R, Trau M. Analysis of exosome purification methods using a model liposome system and tunable-resistive pulse sensing. *Scientific reports.* 2015;5:7639.
35. Mulcahy LA, Pink RC, Carter DRF. Routes and mechanisms of extracellular vesicle uptake. *J Extracell Vesicles.* 2014;3(1):24641.
36. Gilligan KE, Dwyer RM. Extracellular vesicles for cancer therapy: Impact of host immune response. *Cells.* 2020;9(1):224.
37. Chen Y-S, Lin E-Y, Chiou T-W, Harn H-J. Exosomes in clinical trial and their production in compliance with good manufacturing practice. *Tzu-chi Med J.* 2019;32(2):113-20.

38. Gimona M, Pachler K, Laner-Plamberger S, Schallmoser K, Rohde E. Manufacturing of human extracellular vesicle-based therapeutics for clinical use. *International journal of molecular sciences*. 2017;18(6):1190.
39. Gomzikova M, James V, Rizvanov A. Therapeutic application of mesenchymal stem cells derived extracellular vesicles for immunomodulation. *Frontiers in Immunology*. 2019;10:2663.
40. Di Nicola M, Carlo-Stella C, Magni M, Milanese M, Longoni PD, Matteucci P, et al. Human bone marrow stromal cells suppress T-lymphocyte proliferation induced by cellular or nonspecific mitogenic stimuli. *Blood*. 2002;99(10):3838-43.
41. Mokarizadeh A, Delirez N, Morshedi A, Mosayebi G, Farshid A, Mardani K, et al. Microvesicles Derived from Mesenchymal Stem Cells: Potent Organelles for Induction of Tolerogenic Signaling. *Cell J*. 2013;15.
42. Raposo G, Nijman HW, Stoorvogel W, Liejendekker R, Harding CV, Melief C, et al. B lymphocytes secrete antigen-presenting vesicles. *Journal of experimental medicine*. 1996;183(3):1161-72.
43. Zitvogel L, Regnault A, Lozier A, Wolfers J, Flament C, Tenza D, et al. Eradication of established murine tumors using a novel cell-free vaccine: dendritic cell derived exosomes. *Nature medicine*. 1998;4(5):594-600.
44. Latini A, Borgiani P, Novelli G, Ciccacci C. miRNAs in drug response variability: potential utility as biomarkers for personalized medicine. *Pharmacogenomics*. 2019;20(14):1049-59.

45. Wu F, Lu F, Fan X, Chao J, Liu C, Pan Q, et al. Immune-related miRNA-mRNA regulation network in the livers of DHAV-3-infected ducklings. *BMC genomics*. 2020;21(1):1-14.
46. Trobaugh DW, Klimstra WB. MicroRNA regulation of RNA virus replication and pathogenesis. *Trends in molecular medicine*. 2017;23(1):80-93.
47. Cui H, Zhang C, Zhao Z, Zhang C, Fu Y, Li J, et al. Identification of cellular microRNA miR-188-3p with broad-spectrum anti-influenza A virus activity. *Virology journal*. 2020;17(1):12.
48. Kemp V, Laconi A, Cocciolo G, Berends AJ, Breit TM, Verheije MH. miRNA repertoire and host immune factor regulation upon avian coronavirus infection in eggs. *Archives of Virology*. 2020;165:835–43.
49. Zheng B, Zhou J, Wang H. Host microRNAs and exosomes that modulate influenza virus infection. *Virus Research*. 2020:197885.
50. Sardar R, Satish D, Birla S, Gupta D. Comparative analyses of SAR-CoV2 genomes from different geographical locations and other coronavirus family genomes reveals unique features potentially consequential to host-virus interaction and pathogenesis. *bioRxiv*. 2020:DOI: 10.1101/2020.03.21.001586.
51. Hoffmann M, Kleine-Weber H, Schroeder S, Krüger N, Herrler T, Erichsen S, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell*. 2020.
52. Mason RJ. Pathogenesis of COVID-19 from a cell biology perspective. *European Respiratory Journal*. 2020;55:2000607.

53. Dinh P-UC, Paudel D, Brochu H, Popowski KD, Gracieux MC, Cores J, et al. Inhalation of lung spheroid cell secretome and exosomes promotes lung repair in pulmonary fibrosis. *Nature communications*. 2020;11(1):1-14.

**Table 1**

Table 1. Locations and products of single strand RNA (ssRNA) of SARS-CoV-2, MERS, and SARS can be inhibited by antisense RNAs (asRNA) (14)

	SARS-CoV-2	MERS	SARS
asRNA name	miR-5197-3p	miR-6864-5p	miR-4778-3p
asRNA sequence	UAAGCUACUGAGUCAGAGAAGA A	CCGUAUAGACUGAACAGGGAAGUU	AGUUGAGACGUUCCCUUC-UUCU
Matched sequence of ssRNA	AUUCGAAGACCCAGUCCCUACUU	GGCGUUUCUGACUUGUCCCUCAAA	UCGACUCCGCAAGGGAGGUAGGA
Matching (%)	89	88	91
asRNA length	23	24	22
Gene name	spike glycoprotein	orf1ab	orf1ab
Location	21874-21896	1188-1211	1450-1472
Product of complete gene	Surface glycoprotein	1AB polypeptide	Counterpart of MHV p65

**Figure 1**

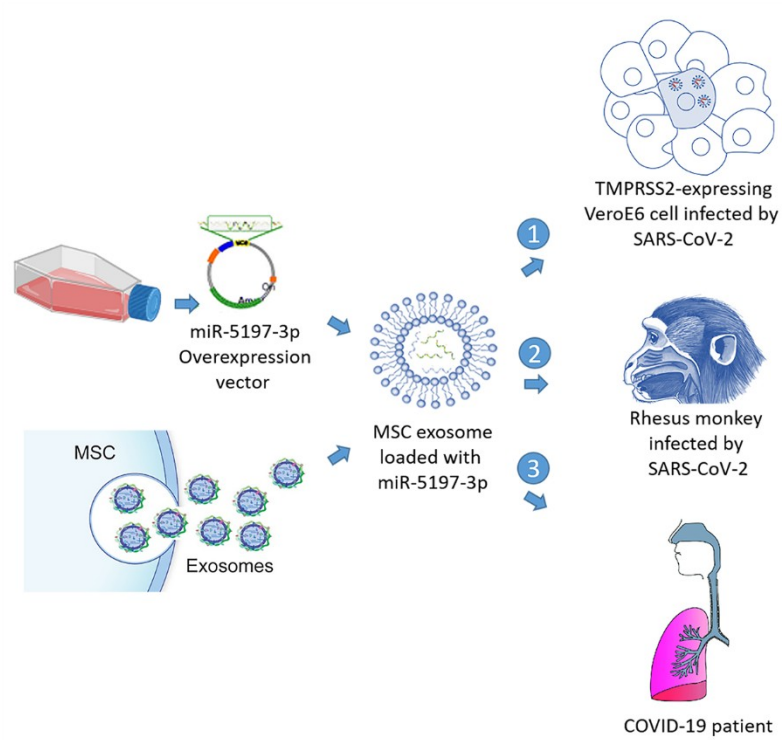


Figure 1. Schematic diagram of developing a therapeutic method for COVID-19 using exosomes of mesenchymal stem cells as nano-cargos for antisense RNAs for inhibiting single strand RNA (ssRNA) of SARS-CoV-2